**Enzyme measurements**

Serum cholesteryl ester transfer protein (CETP) activity was analyzed by a radiometric method as the transfer/exchange of radiolabeled [14C] cholesteryl oleate (Amersham Biosciences) between exogenously added human LDL and HDL, as described previously [1]. Radioactivity in HDL as a measure of transfer activity was determined by liquid scintillation counting. The activity of CETP was expressed as nmol/mL/h. Phospholipid transfer protein (PLTP) activity (nmol/mL/h) and -mass (μg/mL) were determined with radiometric and enzyme-immunoassays as described [2]. Lecithin-cholesterol acyltransferase (LCAT) activity was analyzed by a radiometric assay using a proteoliposome substrate [3]. Briefly, apoA-I -egg lecithin-cholesterol proteoliposome substrate containing radioactive cholesterol tracer was prepared by the cholate dialysis method. The plasma sample and the substrate were incubated at +37C for 30 min after which the LCAT-mediated cholesterol fatty acyl esters were extracted and quantified using thin-layer chromatography for lipid separation and the labelled cholesterol esters were then analyzed for radioactivity. Paraoxonase 1 (PON-1) activity was measured with a chromogenic method [4]. Briefly, in the PON-1 assay, paraoxon (paraoxon-ethyl, D9286-1G, Sigma, St. Louis, MO, USA) was used as the substrate. PON-1 converts paraoxon into *p*-nitrophenol, a yellow compound that can be measured spectrophotometrically at 405 nm. The intra-assay and inter-assay coefficient of variations for PON-1 measurements are 10% and 7%, respectively.

**References**

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