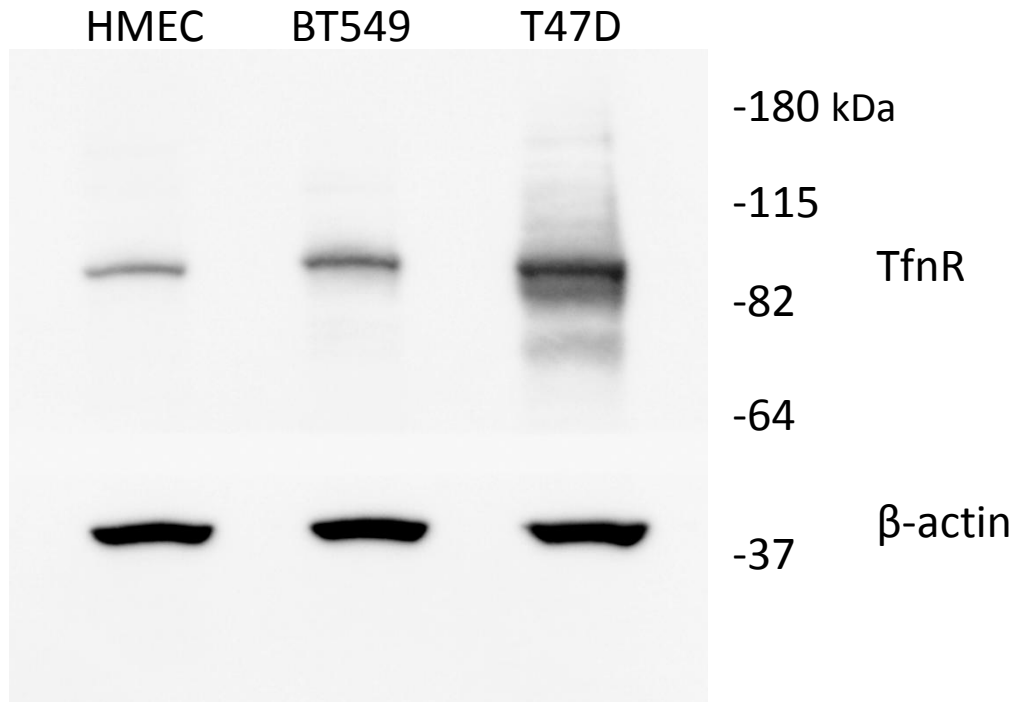
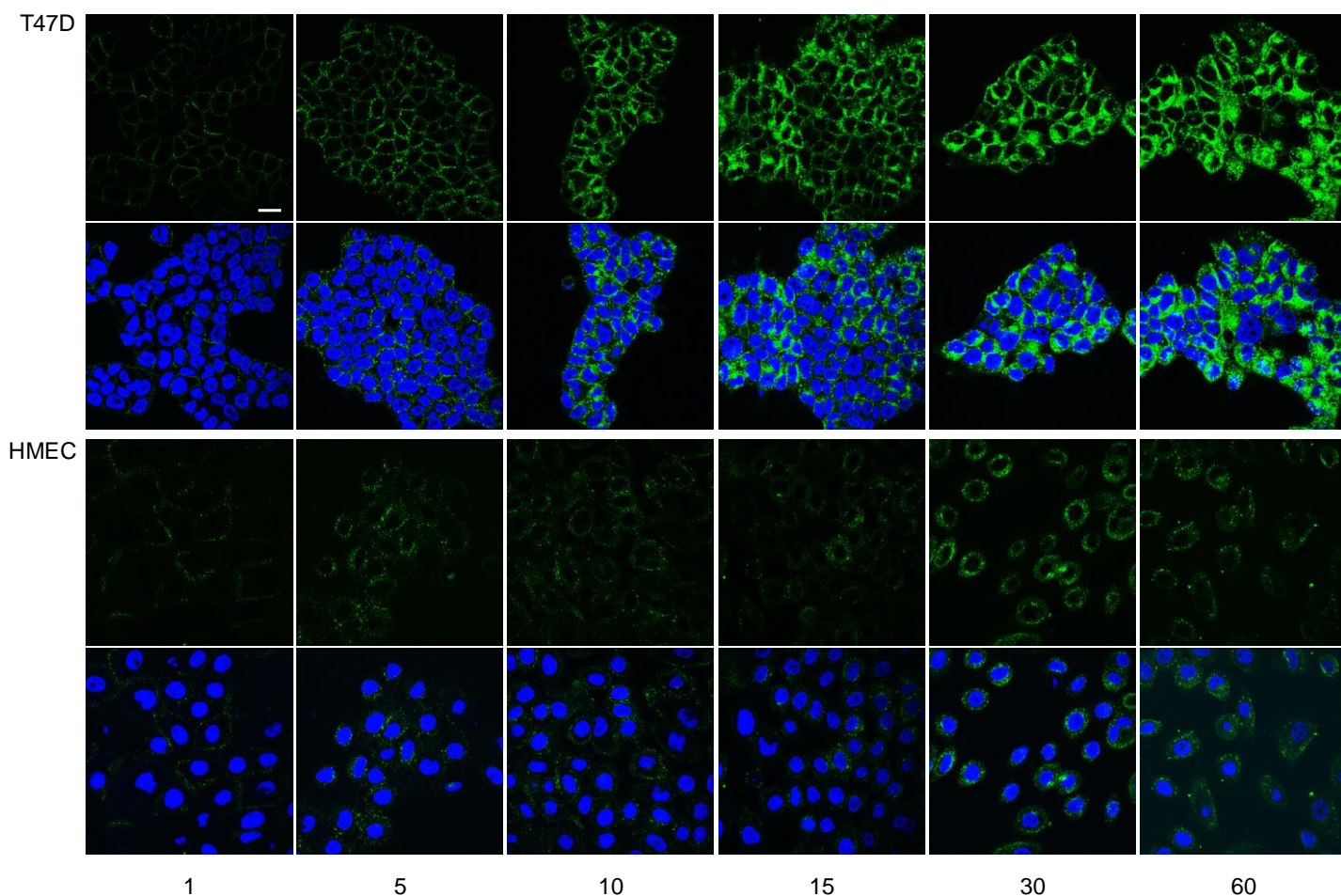


## Supplementary Figures and Tables

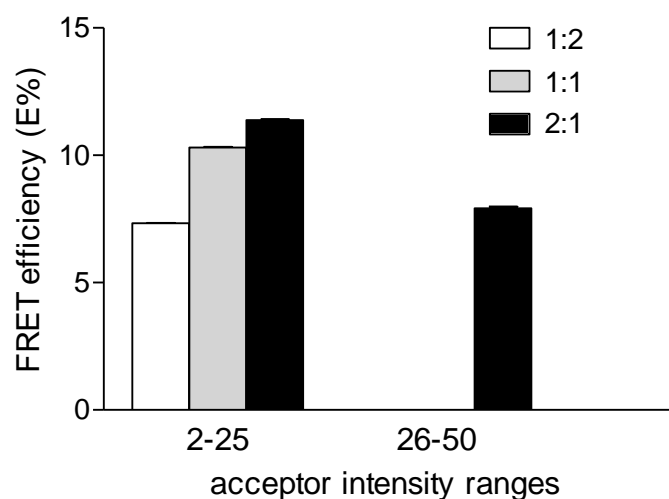
**Abbreviations:** Tfn: transferrin, TfnR: transferrin receptor, A: acceptor, D: donor, AF: AlexaFluor, FRET: Förster Resonance Energy Transfer, E%: energy transfer %, EPR, , HMEC: human mammary epithelial cell, FD: FRET donor SD: standard deviation



**Supplementary Figure S1.** Western Blot of TfnR in HMECs, BT-549, and T47D cells. Blots indicate higher expression of TfnR in human breast cancer cell lines BT549 and T47D compared to normal HMECs



**Supplementary Figure S2.** HMEC and T47D cells are incubated with Tfn conjugated to AF488 (green) for the times as indicated (minutes) and imaged under confocal microscopy with DAPI staining (blue). Scale bar = 20  $\mu\text{m}$ . Tfn-AF488 internalization into HMEC and T47D cells is clearly detected between 1 and 10 min, showing plasma membrane localization at 1-5 min and intracellular peri-nuclear staining at 10-60 min.



**Supplementary Figure S3.** HMEC cells internalized with A:D ratios of 1:2, 1:1, and 2:1 and plotted with binned acceptor intensity per pixel and E% values. Error bars indicate 95% confidence intervals. Absence of higher acceptor intensity ranges reflect decreased uptake of Tfn. A:D ratios of 1:2 and 1:1 in the 6-50 range is likely due to higher ratio of acceptor quenching donor. Similar increase in E% as A:D ratios increase (at acceptor range of 2-25) is observed compared to results found in Fig. 1B (main text).

**Supplementary Table S1 (for Figure 1B)**

Cell Type	T47D	HMEC
Slope	3.770 ± 0.3906	2.410 ± 1.226
95% CI for Slope	-1.193 to 8.732	-13.17 to 17.99
r <sup>2</sup>	0.9894	0.7943
p-value	0.401 (n.s.)	
analysis method	ANCOVA	

Statistical analysis of slopes of FRET efficiency (E%) vs. A:D ratios between T47D and HMECs. No statistical significance is detected between T47D and HMEC slopes using Ancova analysis, suggesting that TfnR-Tfn complexes show similar FRET behavior in HMEC vs. T47D cells. n.s. = not statistically significant (p>0.05)

**Supplementary Table S2 (for Figure 1C)**

Acceptor Range (pixel intensity)	0-25			26-50			51-75			76-100		
A:D ratios	1:2	1:1	2:1	1:2	1:1	2:1	1:2	1:1	2:1	1:2	1:1	2:1
Average (E%)	8.183	11.682	16.613	6.725	10.230	14.400	ND	9.434	13.476	ND	9.871	12.857
95% CI	0.140	0.149	0.391	0.376	0.138	0.168	ND	0.376	0.275	ND	1.807	0.559
Count	894	934	325	34	766	930	ND	67	312	ND	4	72
p-value	<0.0001*			<0.0001*			<0.0001*			p<0.05*		
Analysis Method	one-way ANOVA			one-way ANOVA			t-test			t-test		

Statistical analysis comparing Acceptor ranges among A:D ratios per binned acceptor range. Each acceptor Range column (0-25, 26-50, 51-75, 76-100) includes statistical analysis among the A:D ratios (1:2, 1:1, 2:1). One-way ANOVA is used for analysis in binned acceptor ranges of 0-25, 26-50. t-test analysis is performed for binned acceptor ranges of 51-75 and 76-100. The results show that at a fixed range of acceptor intensities, E% is positively dependent on A:D ratios, as expected for intra-molecular FRET between Tfn molecules bound to homodimeric TfnR; inter-molecular FRET between clustered TfnR-Tfn complexes cannot be excluded (8). p-values < 0.05 are considered significant (\*).

**Supplementary Table S3 (for Figure 2A)**

A:D ratio	T47D Short lifetime±SD (ps)	HMEC Short lifetime±SD (ps)
0:1	333±36	321±36
1:4	287±33	328±45
1:3	303±44	314±33
1:2	298±48	301±4
1:1	319±31	285±24
2:1	305±35	320±33
3:1	295±40	304±13

Short component lifetimes are estimated by biexponential fitting for both T47D and HMEC at A:D ratio from 0:1 to 3:1. The mean short lifetime for both T47 and HMEC is around 300ps. Small SDs are obtained (~32ps). These results indicate a high sensitivity and uniform detection despite the heterogeneity of donor intensities in the sample.

**Supplementary Table S4 (Figure 2B)**

Cell Type	T47D	HMEC
Slope	7.767 ± 0.5321	6.896 ± 0.5845
95% CI for Slope	6.290 to 9.244	5.274 to 8.519
r <sup>2</sup>	0.9581	0.9530
p-value (slope comparison)	0.3029 (n.s.)	
analysis method	ANCOVA	

Statistical analysis of slopes of FD% vs. A:D ratios between T47D and HMECs. No statistical significance is detected between T47D and HMEC slopes using Ancova analysis, suggesting that TfnR-Tfn complexes show similar FRET behavior in HMEC vs. T47D cells, as shown in Supplementary Table 1. n.s. = not statistically significant (p>0.05).

**Supplementary Table S5 (Figure 2C)**

Acceptor Ranges (μg/mL)	2.5			5			10			20		
A:D ratios	1:2	2:1	3:1	1:2	2:1	3:1	1:2	2:1	3:1	1:2	2:1	3:1
FD%	14	25	40	14	24	32	14	25	31	14	25	31
SD	4	4	8	4	5	5	3	5	5	3	4	3
p-value	<0.001*			<0.01*			<0.01*			<0.001*		
Analysis Method	1-way ANOVA			1-way ANOVA			1-way ANOVA			1-way ANOVA		

Statistical analysis comparing % FD among A:D ratios using acceptor ranges of 2.5, 5, 10, and 20 μg/ml Tfn. p-value is the comparison of the % FD among all A:D ranges (1:2, 2:1, 3:1). The results show that at a fixed range of acceptor concentrations, FD% is positively dependent on A:D ratios, as expected for intra-molecular FRET between Tfn molecules bound to homodimeric TfnR; inter-molecular FRET between clustered TfnR-Tfn complexes cannot be excluded (8). p-values < 0.05 are considered significant (\*).

**Supplementary Table S6 (Figure 2D)**

Cell Number	T47D (1x10 <sup>5</sup> cells)	T47D (1x10 <sup>4</sup> cells)
Slope	9.631 ± 0.7624	7.767 ± 0.5321
95% CI for Slope	7.514 to 11.75	6.290 to 9.244
r <sup>2</sup>	0.9755	0.9816
p-value (slope comparison)	0.08 (n.s.)	
analysis method	ANCOVA	

Statistical analysis of slopes of FD% vs. A:D ratios in T47D cells at 1x10<sup>5</sup> and 1x10<sup>4</sup> cells. No statistical significance is detected using Ancova analysis, suggesting sensitivity of % FD is consistent across 1 x10<sup>5</sup> to 1 x10<sup>4</sup> T47D cells. n.s. = not statistically significant (p>0.05).

**Supplementary Table S7 (for Figure 3A)**

<b>A:D ratios</b>	<b>20 µg/mL Short lifetime±SD (ps)</b>	<b>40 µg/mL Short lifetime±SD (ps)</b>
0:1	333±35	322±42
2:1	320±38	336±18

Short component lifetimes measured in picoseconds are shown, indicating a high sensitivity and uniform detection despite the heterogeneity of donor intensities in the sample.

**Supplementary Table S8 (for Figure 3B)**

<b>Amount of Tfn</b>	<b>20 µg/mL</b>		<b>40 µg/mL</b>	
<b>A:D ratios</b>	0:1 (donor only)	2:1	0:1 (donor only)	2:1
<b>FD%</b>	11	39	13	37
<b>SD</b>	4	3	4	3
<b>p-value</b>	<0.001*		<0.005*	
<b>Analysis Method</b>	t-test		t-test	

Statistical analysis comparing % FD between 0:1 and 2:1 A:D ratios for each amount of Tfn (20 µg/ml and 40 µg/ml). Statistical significance is reached comparing 0:1 vs. 2:1 A:D ratios for all Tfn amounts. The results show that at a fixed range of acceptor concentrations, FD% is positively dependent on A:D ratios, as shown in supplementary table 5. p-values < 0.05 are considered significant (\*).

**Supplementary Table 9 (for Figure 3B)**

<b>A:D ratios</b>	<b>0:1 (donor only)</b>		<b>2:1</b>	
<b>Amount of Tfn</b>	20 µg/mL	40 µg/mL	20 µg/mL	40 µg/mL
<b>FD%</b>	11	13	39	37
<b>SD</b>	4	3	3	3
<b>p-value</b>	0.5734 (n.s.)		0.4601 (n.s.)	
<b>Analysis Method</b>	t-test		t-test	

Statistical analysis comparing % FD between 20 µg/ml and 40 µg/ml at either 0:1 or 2:1 A:D ratios. No statistical significance between each comparison is detected, suggesting sensitivity of % FD is consistent across internalization of 20 µg/mL and 40 µg/mL of Tfn. n.s. = not statistically significant (p>0.05).

**Supplementary Table S10 (for Figure 3B)**

<b>Source of Variation</b>	<b>Interaction</b>	<b>A:D Ratio</b>	<b>Tfn amount</b>
% Total Variation	0.56	94.77	0.00
p-value	0.3559 (n.s.)	< 0.0001*	1.0000 (n.s.)
Analysis Method	two-way ANOVA		

Two-way ANOVA table to determine which parameter, A:D ratio or Tfn amount accounts for the variation in results. As shown, Acceptor:Donor ratios demonstrate the strongest influence in variation of data. n.s. = not statistically significant (p>0.05); p-values < 0.05 are considered significant (\*).

**Supplementary Table S11 (for Figure 4A)**

<b>A:D ratios</b>	<b>Short lifetime±SD (ps)</b>
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0:1	318±34
1:2	317±50
1:1	306±44
2:1	322±39

Short component lifetimes are estimated by biexponential fitting for live mice at A:D ratio from 0:1 to 2:1. The mean short lifetime is around 300ps. Small standard deviations are obtained (~40ps). These *in vivo* results are consistent with the *in vitro* data and demonstrate that our imaging platform is sensitive and quantitatively matches *in vitro* results generated by confocal microscopy.

**Supplementary Table S12 (for Figure 4B)**

A:D ratios	1:2	2:1
FD%	24	36
SD	4	3
p-value	p< 0.0001*	
Analysis Method	t-test	

FD% values are listed for A:D ratios of 1:2 and 2:1. Analysis indicates statistical significance between low and high values supporting a linear increase in FD% as A:D ratios increase. p-values < 0.05 are considered significant (\*).