

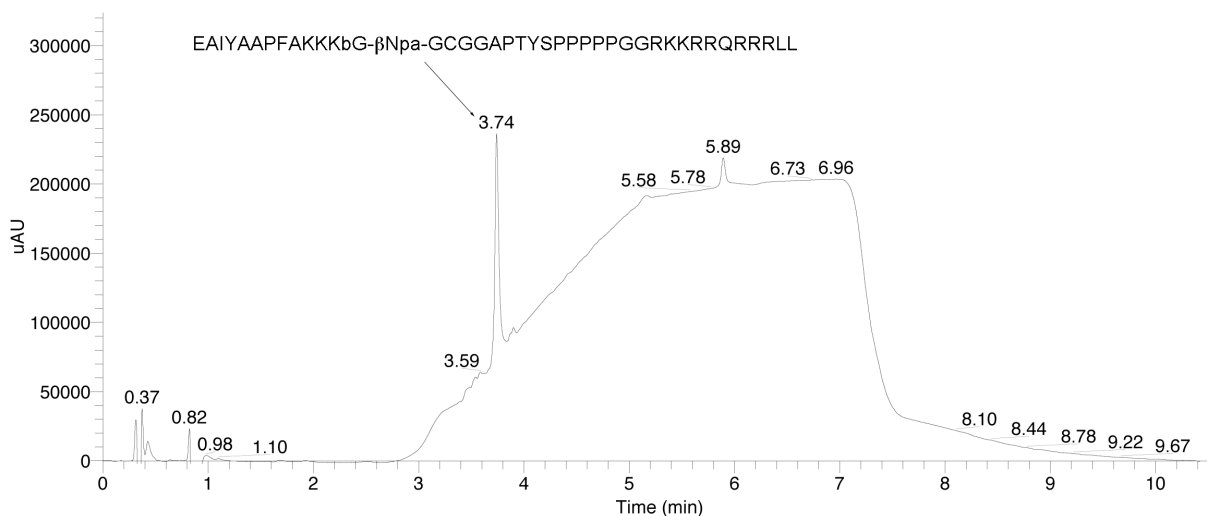
Supporting information for “A multiple reaction monitoring (MRM) method to detect Bcr-Abl kinase activity in CML using a peptide biosensor,” Yang et al.

Figure S1. LC/MS characterization of Abl biosensor peptide. Used in assays and also spiked into K562 lysate for digestion and use as MRM calibration standard for EAIYAAPFAK fragment.

E:\Xcalibur\...\03_FL_Abltide

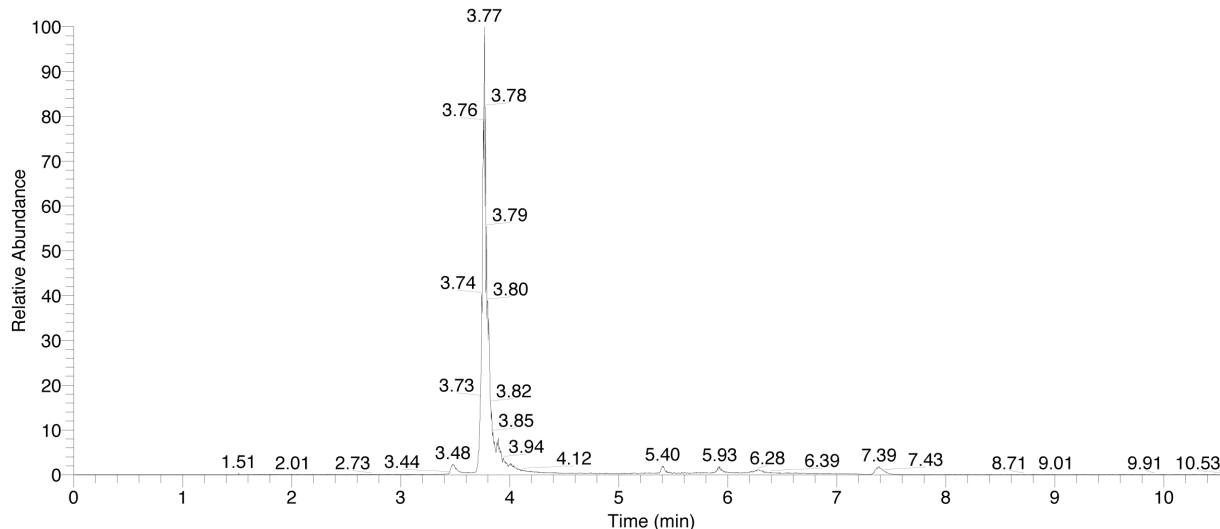
6/23/2011 7:08:38 PM

RT: 0.00 - 10.58



NL:
3.24E5
Channel A
UV
03_FL_Ablti
de

RT: 0.00 - 10.60



NL:
1.33E8
TIC MS
03_FL_Ablti
de

03_FL_Abltide #1101-1163 RT: 3.70-3.90 AV: 63 NL: 4.12E5
T: ITMS + p ESI Full ms [400.00-2000.00]

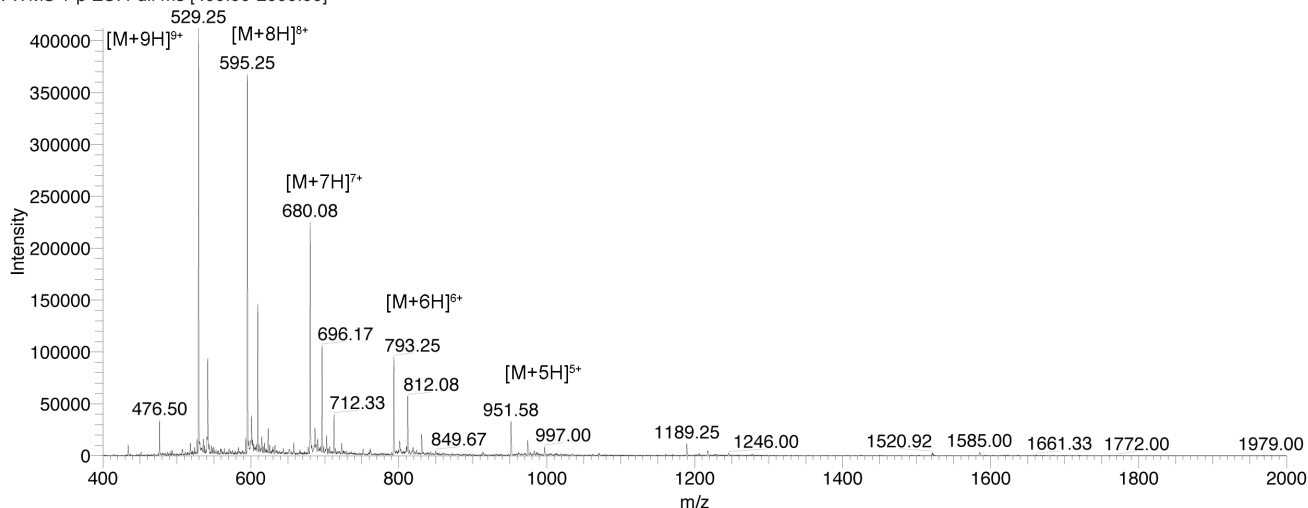
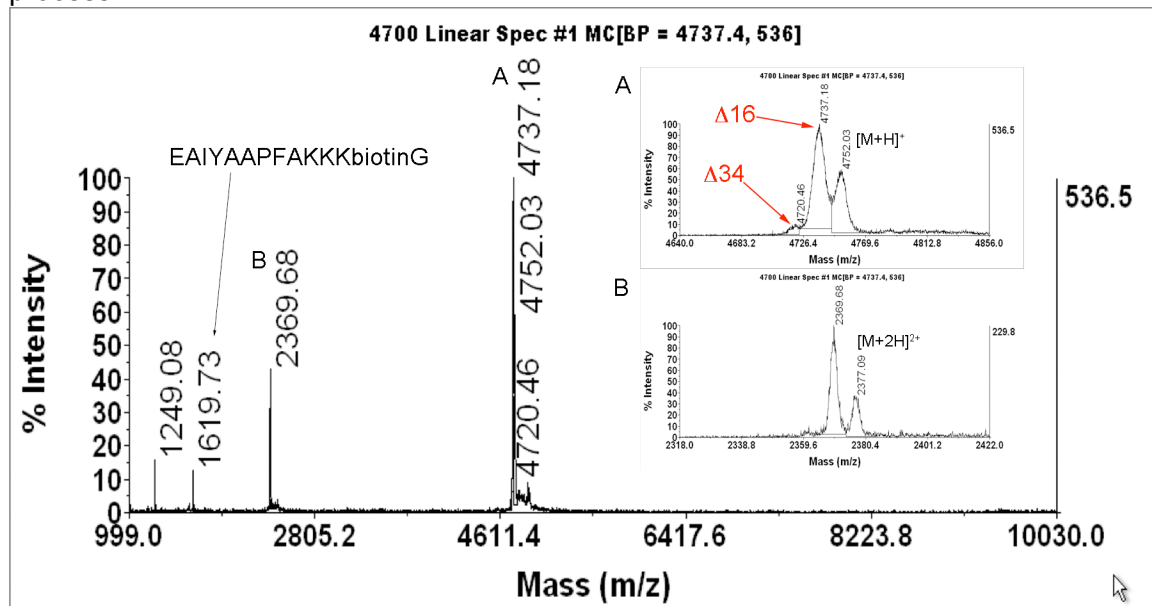


Figure S2. MALDI-TOF characterization of Abl biosensor; shows what appear to be laser-induced photolytic degradation peaks that may represent the intermediates proposed in Scheme S1.¹⁻² While the resolution at this mass range makes the m/z differences appear to be 15-17 and 32-34, similar peaks in spectra from other fragments (including Fig. S5 and Fig. S6) show that these losses are more likely to be 16 and 34, and these values are consistent with a plausible proposed mechanism for their formation (Scheme 1). Photolysis peaks are not observed in the LC/ESI-MS spectrum given in S1, indicating that this is a laser desorption mediated process.



1. Gaplovsky, M.; Il'ichev, Y. V.; Kamdzhilov, Y.; Kombarova, S. V.; Mac, M.; Schworer, M. A.; Wirz, J., Photochemical reaction mechanisms of 2-nitrobenzyl compounds: 2-Nitrobenzyl alcohols form 2-nitroso hydrates by dual proton transfer. *Photoch Photobio Sci* **2005**, 4 (1), 33-42.
2. Il'ichev, Y. V.; Schworer, M. A.; Wirz, J., Photochemical reaction mechanisms of 2-nitrobenzyl compounds: Methyl ethers and caged ATP. *Journal Of The American Chemical Society* **2004**, 126 (14), 4581-4595.

Scheme S1. Proposed potential photodegradation intermediates (as observed in MALDI-TOF spectra). The observed ions exhibit m/z values of 16 amu (major) and 34 amu (minor) less than the expected m/z . The major species is likely formed via gas phase ion chemistry involving protonation of one of the oxygens in the nitro group by initial photo-tautomerization (which is well-characterized in the literature, see 1. and 2. above), additional protonation by acid and elimination as water. For the minor product, the second loss of water (18 amu) to give a total net loss of 34 amu overall could occur—however, the mechanism for this is not clear and while it is very interesting, its absolute determination would be outside the scope of this work. However, this phenomenon suggests that MALDI-MS might be a useful way to study gas phase photochemistry, and this could be the subject of future investigation.

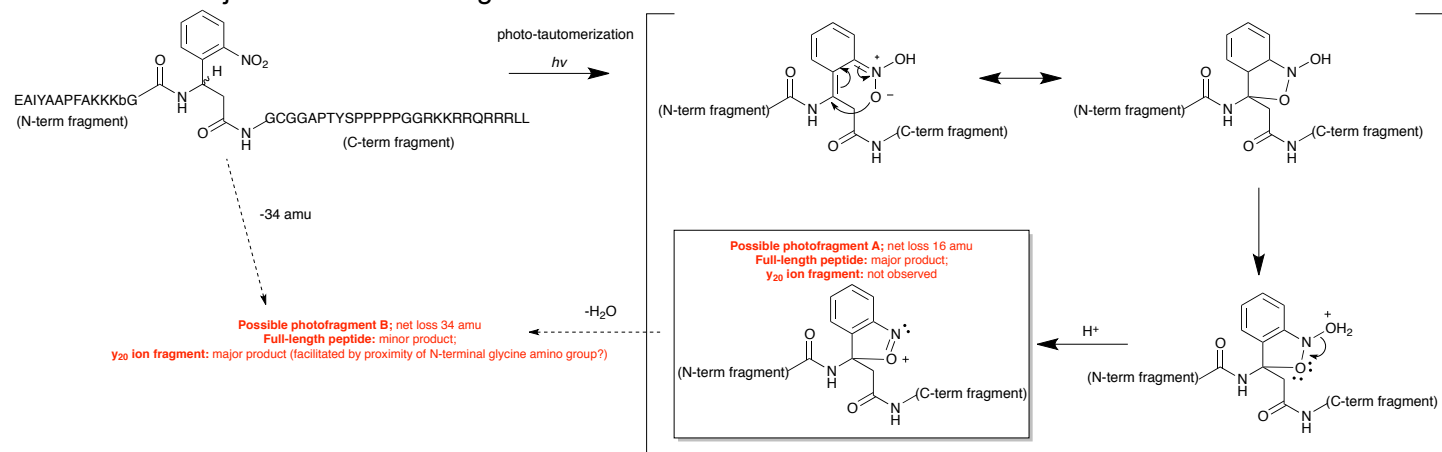
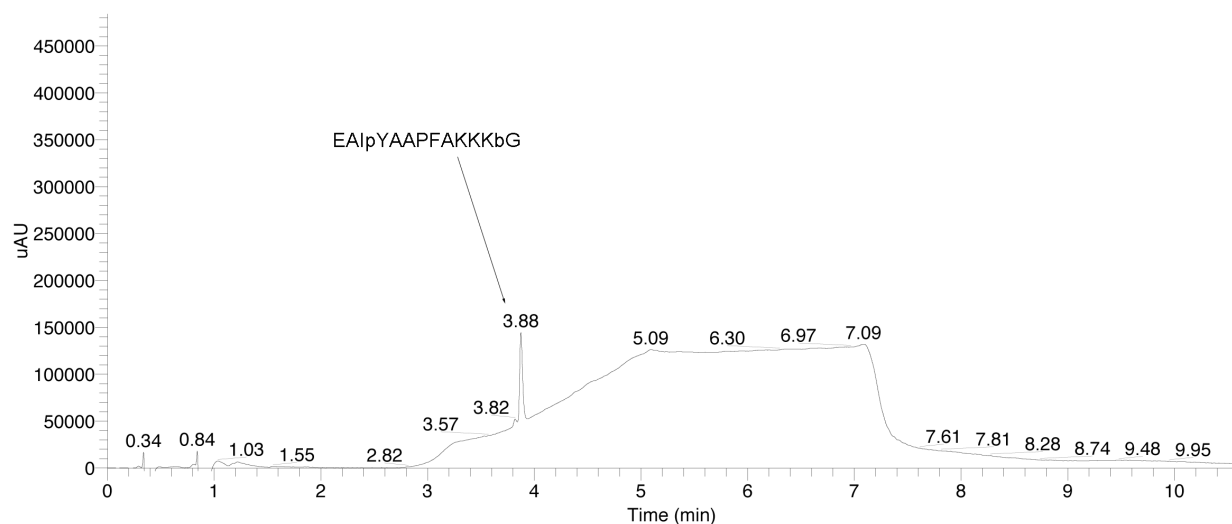


Figure S3. Synthetic phosphorylated Abl biosensor reporter module. Added to K562 lysate followed by digestion and use as MRM calibration standard.

E:\Xcalibur\...110811_02_pAbtide1

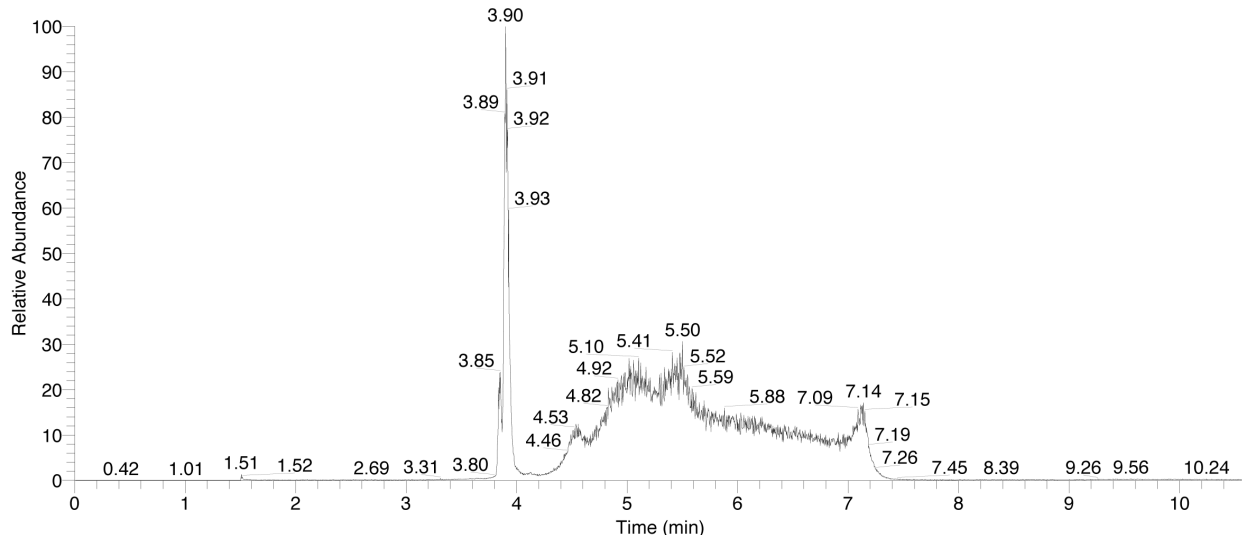
8/11/2011 12:15:30 PM

RT: 0.00 - 10.58



NL:
4.84E5
Channel A
UV
110811_02
_pAbtide1

RT: 0.00 - 10.60



NL:
3.14E7
TIC MS
110811_02
_pAbtide1

110811_02_pAbtide1 #1151-1170 RT: 3.87-3.93 AV: 20 NL: 6.23E5

T: ITMS + p ESI Full ms [400.00-2000.00]

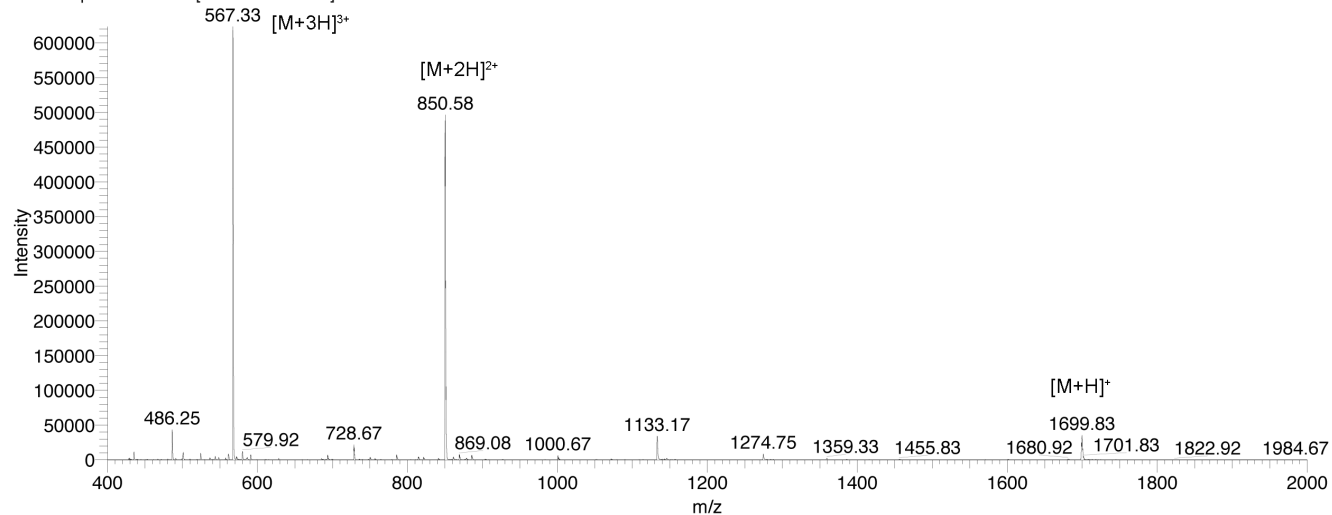
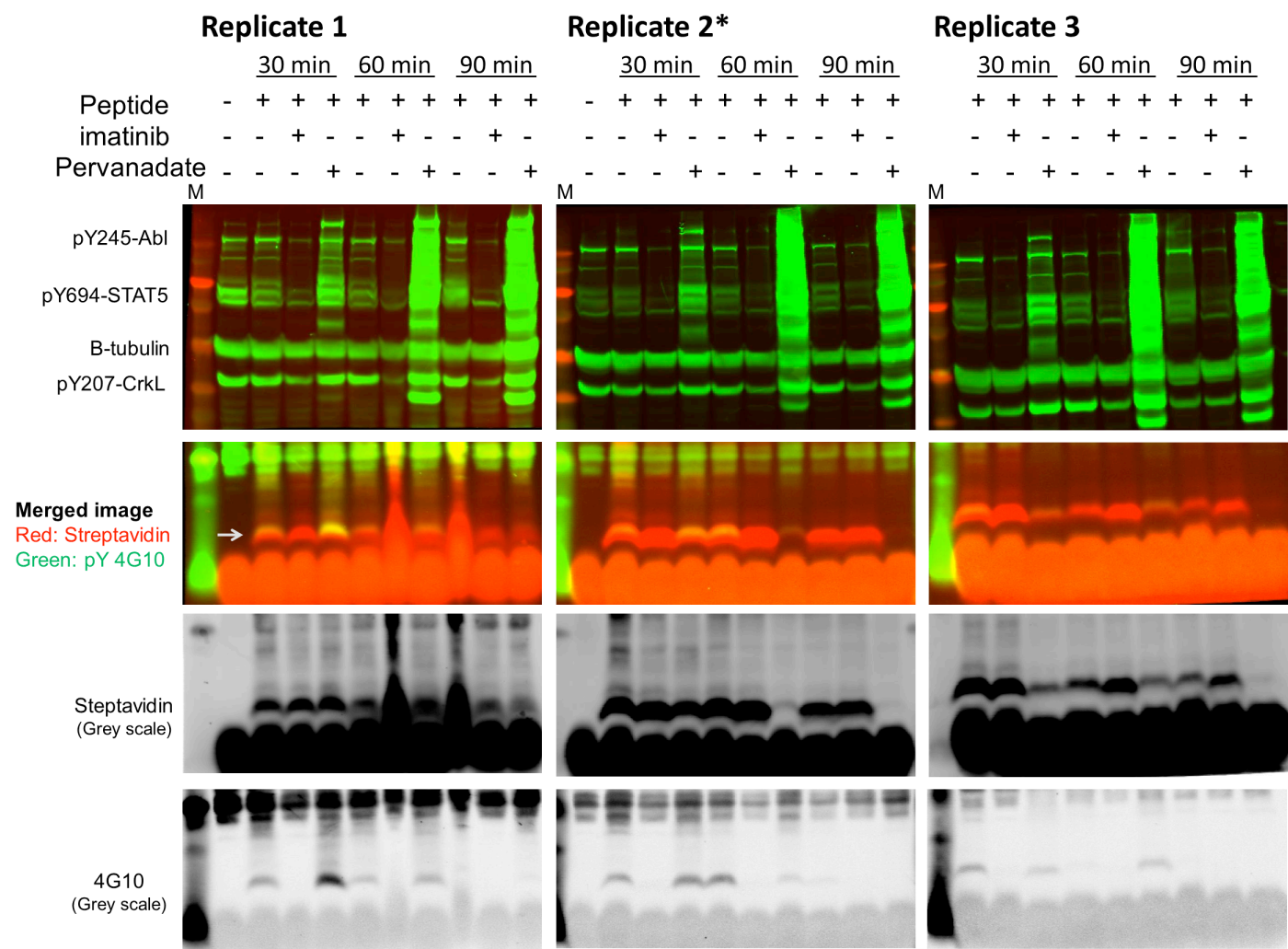


Figure S4. Additional Western blots from all replicates for data shown in Figure 2 in the main manuscript.



*A grey scale version of replicate 2 was shown in fig 2A.

Figure S5. Expanded view of MALDI-TOF spectrum for degradation products.

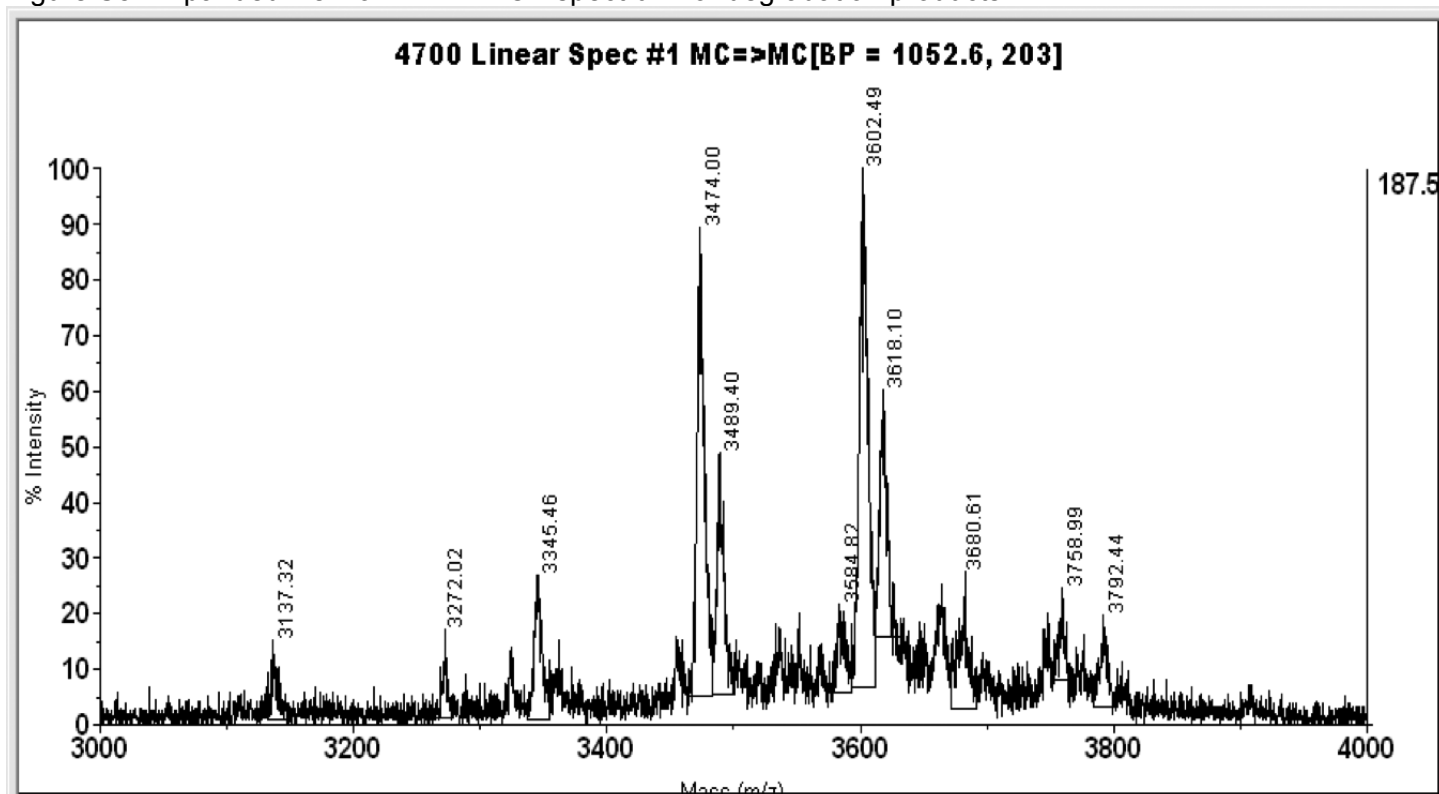
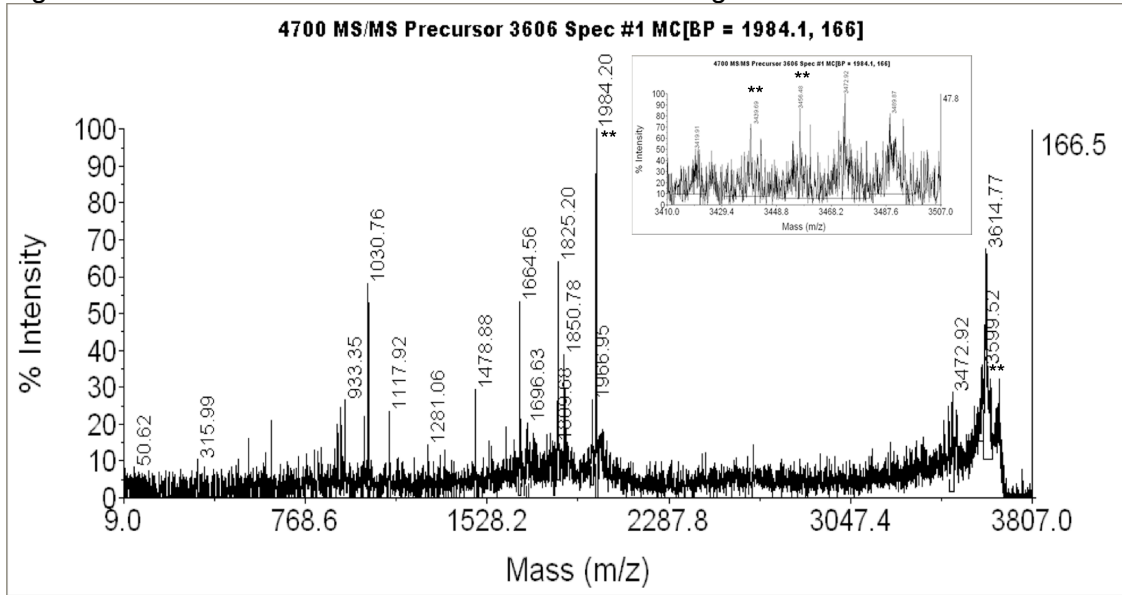


Figure S6. MALDI-TOF/TOF MS/MS for 3618/3602 fragment



y ion	sequence	m/z	b ion	sequence	m/z
32	EAIYAAPFAKKK _{biotin} GβNpaGCGGAP TYSPPPPPGGRKK-OH	3490.9 6111	1	H2N-E	130.12 317
31	AIYAAPFAKKK _{biotin} GβNpaGCGGAPT YSPPPPPGGRKK-OH	3419.8 8251	2	H2N-EA	201.20 177
30	IYAAPFAKKK _{biotin} GβNpaGCGGAPT SPPPPPGGRKK-OH	3306.7 2351	3	H2N-EAI	314.36 077
29	YAAPFAKKK _{biotin} GβNpaGCGGAPT SPPPPPGGRKK-OH	3143.5 4791	4	H2N-EAIY	477.53 637
28	AAPFAKKK _{biotin} GβNpaGCGGAPT PPPPPGGRKK-OH	3072.4 6931	5	H2N-EAIYA	548.61 497
27	APFAKKK _{biotin} GβNpaGCGGAPT PPPPGGRKK-OH	3001.3 9071	6	H2N-EAIYAA	619.69 357
26	PFAKKK _{biotin} GβNpaGCGGAPT PPPGGRKK-OH	2904.2 7431	7	H2N-EAIYAAP	716.80 997
25	FAKKK _{biotin} GβNpaGCGGAPT PPGGRKK-OH	2757.0 9811	8	H2N-EAIYAAPF	863.98 617
24	AKKK _{biotin} GβNpaGCGGAPT PGGRKK-OH	2686.0 1951	9	H2N-EAIYAAPFA	935.06 477
23	KKK _{biotin} GβNpaGCGGAPT GGRKK-OH	2557.8 4591	10	H2N-EAIYAAPFAK	1063.2 3837
22	KK _{biotin} GβNpaGCGGAPT GRKK-OH	2429.6 7231	11	H2N-EAIYAAPFAKK	1191.4 1197
21	K _{biotin} GβNpaGCGGAPT RKK-OH	2075.2 0231	12	H2N-EAIYAAPFAKKK _{biotin}	1545.8 8197
20	GβNpaGCGGAPT -OH** (observed m/z: 1984.20)	2018.1 5051	13	H2N-EAIYAAPFAKKK _{biotin} G	1602.9 3377
19	bNpaGCGGAPT OH	1825.0 9051	14	H2N-EAIYAAPFAKKK _{biotin} GβNpa	1795.9 9377
18	GCGGAPT OH	1768.0 3871	15	H2N-EAIYAAPFAKKK _{biotin} GβNpaG	1853.0 4557
17	CGGAPT OH	1664.9 0011	16	H2N-EAIYAAPFAKKK _{biotin} GβNpaGC	1956.1 8417

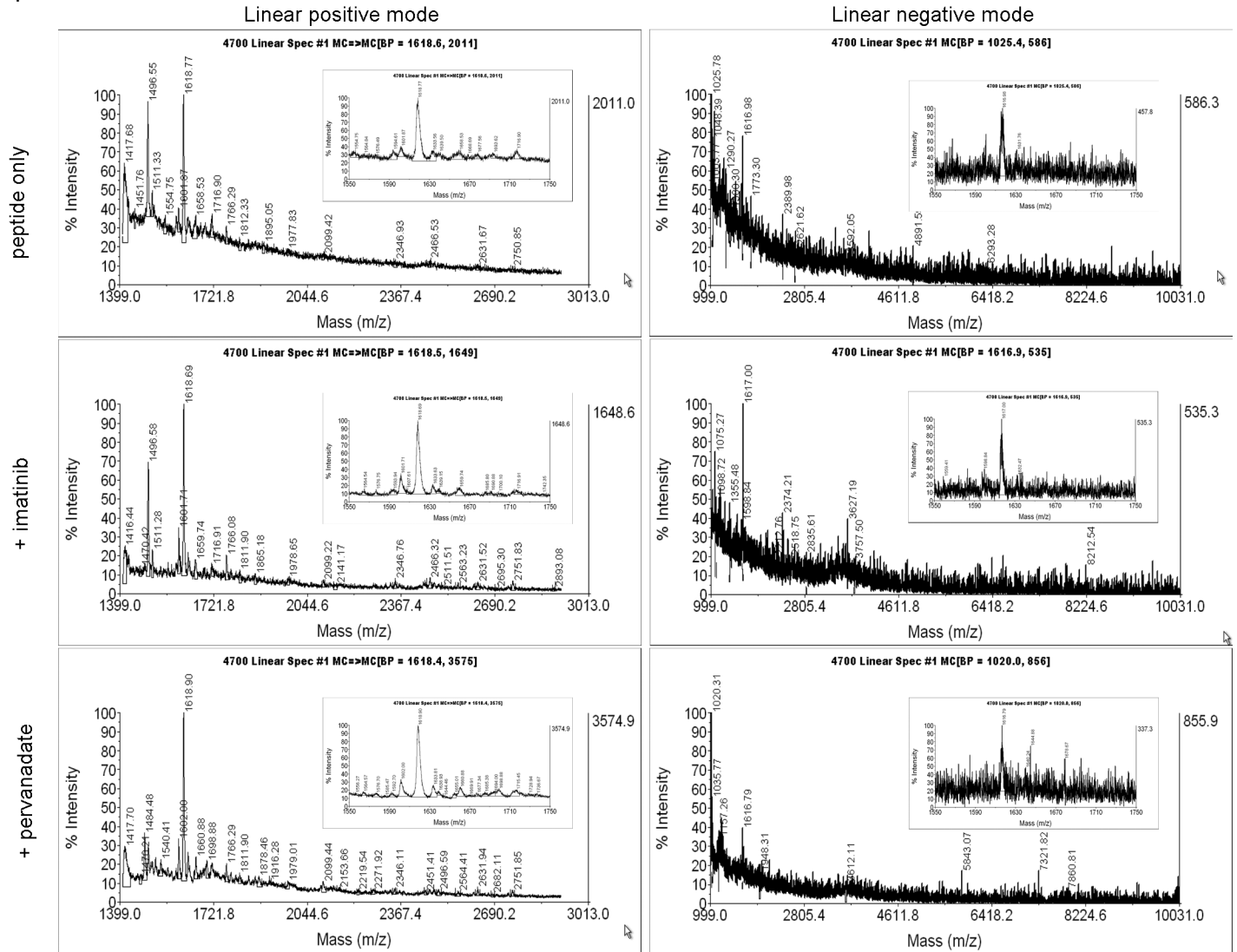
16	GGAPTYSPPPPPGGRKK-OH	1607.8 4831	17	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG	2013.2 3597
15	GAPTYSPPPPPGGRKK-OH	1550.7 9651	18	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG	2070.2 8777
14	APTYSPPPPPGGRKK-OH	1479.7 1791	19	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG A	2141.3 6637
13	PTYSPPPPPGGRKK-OH	1382.6 0151	20	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG AP	2238.4 8277
12	TYSPPPPPGGRKK-OH	1281.4 9671	21	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APT	2339.5 8757
11	YSPPPPPGGRKK-OH	1118.3 2111	22	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTY	2502.7 6317
10	SPPPPPGGRKK-OH	1031.2 4311	23	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYS	2589.8 4117
9	PPPPPGGRKK-OH	934.12 671	24	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPP	2686.9 5757
8	PPPPGGRKK-OH	837.01 031	25	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPP	2784.0 7397
7	PPPGGRKK-OH	739.89 391	26	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPPP	2881.1 9037
6	PPGGRKK-OH	642.77 751	27	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPPPP	2978.3 0677
5	PGGRKK-OH	545.66 111	28	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPPPPP	3075.4 2317
4	GGRKK-OH	488.60 931	29	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPPPPPG	3132.4 7497
3	GRKK-OH	431.55 751	30	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPPPPPGG	3189.5 2677
2	RKK-OH	275.37 051	31	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPPPPPGGR	3345.7 1377
1	KK-OH	147.19 691	32	H2N- EAIYAAPFAKKK _{biotin} GβNpaGCGG APTYSPPPPPGGRK	3473.8 8737

Predicted fragmentation for 3618/3602 degradation product

*Observed/tentatively assigned

**Ion observed (m/z 1984.2) has m/z consistent with MS/MS fragment y₂₀ exhibiting loss of 34 amu, which may predominate due to some effect of the proximity of the N-terminal glycine in this particular fragment (which, when protonated, could act as a proton donor); other potential photodegradation peaks also marked as ** in spectrum. For further discussion, see Fig. S2 and Scheme S1.

Figure S7. MALDI-TOF to analyze enriched reporter segment.
Replicate 1:



Replicate 2:

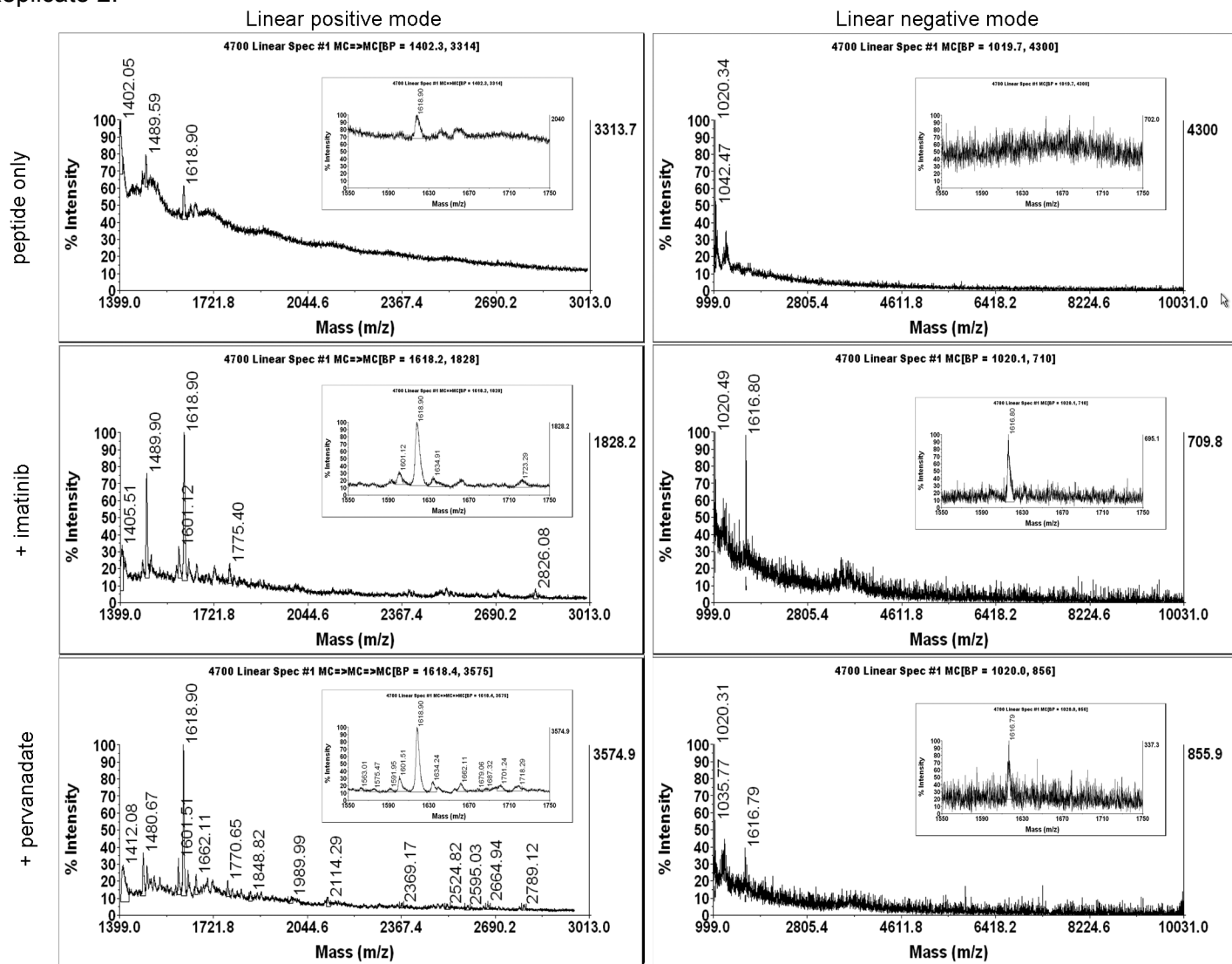


Table S8. Table of raw MRM data. See additional .csv file.

Table S9. Table of peak integrations and calibration. See attached Excel file.