PCR-reactions took place in 96-well plates using 1 μL of cDNA, 10 μL of Power SYBR Green PCR Master Mix (Applied Biosystems), and primers at a final concentration of 900 nM in a total volume of 20 μL. The PCR reaction had the standard amplification scheme: one cycle of 2 minutes at 50°C (AmpErase UNG activation), one cycle of 10 minutes at 95°C (Gold AmpliTaq activation, AmpErase UNG inactivation), followed by 40 cycles of denaturation for 15 seconds at 95°C and annealing/extension for 1 minute at 60°C in a ABI 7900 HT Sequence Detection System (Applied Biosystems). CT cycle values were correlated to a standard curve. The resulting mRNA levels relative to the 18S rRNA or rpl32 were calculated according to the standard formula 2-ΔΔCT, where ΔΔCT= (CTTarget\_sample - CTendogenous control \_sample)-(CTTarget\_calibrator –CTendogenous control\_calibrator) as described by user bulletin #2; Applied Biosystems.