# Non-invasive Predictors of Human Cortical Bone Mechanical Properties: T<sub>2</sub>-Discriminated <sup>1</sup>H NMR Compared with High Resolution X-ray

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## Abstract

Recent advancements in magnetic resonance imaging (MRI) have enabled clinical imaging of human cortical bone, providing a potentially powerful new means for assessing bone health with molecular-scale sensitivities unavailable to conventional X-ray-based diagnostics. To this end, <sup>1</sup>H nuclear magnetic resonance (NMR) and high-resolution X-ray signals from human cortical bone samples were correlated with mechanical properties of bone. Results showed that <sup>1</sup>H NMR signals were better predictors of yield stress, peak stress, and pre-yield toughness than were the X-ray derived signals. These <sup>1</sup>H NMR signals can, in principle, be extracted from clinical MRI, thus offering the potential for improved clinical assessment of fracture risk.

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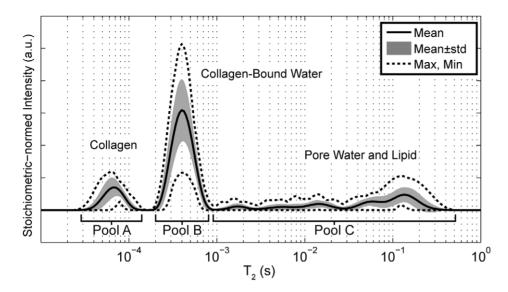
## Introduction

Current bone diagnostics are incomplete. The estimate of areal bone mineral density (BMD) by dual energy x-ray absorptiometry (DXA) does not fully predict fracture risk: for a given DXA score, there is an unexplained increase in fracture risk with age [1,2], as well as with progression of various disease states, such as diabetes [3]. The limitations of DXA related to BMD depending on bone size [4] may be somewhat overcome by quantitative computed tomography imaging, but, ultimately, any X-ray based diagnostic is only sensitive to the mineral portion of the bone, which accounts for only  $\approx 43\%$  of bone by volume. The remaining soft-tissue components of bone, including collagen and collagen-bound water, are essentially invisible to DXA and quantitative computed tomography. In contrast, clinical magnetic resonance imaging (MRI), which is based on the <sup>1</sup>H NMR signal, cannot directly detect bone mineral but is sensitive to the soft tissue of bone. Further, a recent study has demonstrated that <sup>1</sup>H NMR transverse relaxation time constants (T<sub>2</sub>) distinguishes proton signals from collagen, collagen-bound water, and pore water [5]. With this technology and the idea that the presence and hydration-state of collagen play a critical role in dissipating energy in bone [6], we hypothesized that <sup>1</sup>H NMR can report on the material strength of bone, and we present here compelling experimental observations of <sup>1</sup>H NMR, X-ray CT and mechanical tests of cadaveric bone samples which indicate that MRI has the potential to better diagnose fracture risk than DXA.

## Results

Figure 1 shows the mean (and standard deviation and range) spectrum of <sup>1</sup>H NMR transverse relaxation time constants (T<sub>2</sub>) spectrum) from 40 cadaveric bone samples. In this mean spectrum and in each individual sample spectrum, signals from three distinct domains of  $T_2$  were readily identified, as previously found [5]: 50  $\mu$ s<T<sub>2</sub><150  $\mu$ s, defined as pool A, due primarily to collagen methylene protons; 150 µs<T<sub>2</sub><1 ms, pool B, due primarily to collagen-bound water protons; and 1 ms<T<sub>2</sub><1 s, pool C, due to water protons in pores in lipid protons. From these three signals, six parameters were extracted: 3 signal amplitudes ( $S_A$ ,  $S_B$ ,  $S_C$ , in absolutes units of mole <sup>1</sup>H per liter bone) and 3 corresponding mean relaxation rate constants  $(R_{2,A}, R_{2,B}, R_{2,C} \text{ in s}^{-1})$ . Note that while the signal amplitudes are computed in absolute units of concentration, the correspondence between signal amplitudes,  $S_A$ ,  $S_B$ , and  $S_C$ , and actual concentrations of collagen methylene protons, bound water protons, and pore-water or lipid protons, respectively, is potentially affected by a number of factors, including the line shape of the methylene protons, the magnetization exchange rate between bound and methylene protons, and overlap of T<sub>2</sub> components from different sources.

Each of the three NMR signal amplitudes ( $S_A$ ,  $S_B$ ,  $S_C$ ) was found to linearly correlate ( $r^2 = 0.34$ , 0.68, 0.61, p<0.05) with peak stress (Fig. 2), but note that the sum of all three signals did not ( $r^2 = 0.06$ , p>0.05). Similar pair-wise linear correlations (and lack thereof) also existed between NMR signal amplitudes and the other three



**Figure 1. Summary of T<sub>2</sub> spectra measured from 40 human cortical bone samples.** All spectra exhibited a short-T<sub>2</sub> component ( $T_2 \approx 60 \ \mu s$ ), derived primarily from collagen protons, an intermediate T<sub>2</sub> components ( $T_2 \approx 400 \ \mu s$ ), derived primarily from collagen-bound water protons, and a broad distribution of long-T<sub>2</sub> components (1 ms<T<sub>2</sub><1 s), derived from a combination of pore water and lipid protons. doi:10.1371/journal.pone.0016359.g001

measured mechanical properties. These findings indicate that peak cortical bone stress, and the other measured mechanical properties, are directly related to the amount of collagen and collagen-bound water in bone, and inversely related to the bone pore volume. Micro-computed tomography ( $\mu$ CT)-derived measures of bone porosity and the apparent volumetric bone mineral density (avBMD, akin to DXA) were also found to linearly correlate with mechanical properties, but  $S_A$  and  $S_B$  were better predictors (i.e., higher r<sup>2</sup> values) than  $\mu$ CT-porosity for three of four mechanical properties (flexural modulus being the exception), and better predictors than avBMD (i.e., DXA) for all four mechanical properties. Table 1 summarizes the pairwise linear correlations between imaging measure (<sup>1</sup>H NMR and X-ray) and the four mechanical properties.

Note that without the two apparent outlier data (peak stress  $\approx 100$  MPa), the predictive power of  $S_B$  and  $S_C$  decreased to  $r^2$ 

values of 0.52 and 0.49, respectively, but the  $r^2$  of avBMD with peak stress decreased to a greater extent (to 0.16). That is, the relative predictive power of  $S_B$  and  $S_C$  compared with avBMD *increased* without these two data points. Also note that multiple linear regression analysis told a similar story: combination of NMR signal parameters ( $R_B$  and  $S_B$ ) best predicted of three of four mechanical properties (adjusted  $\mathbb{R}^2$ : 0.56-0.70, again, flexural modulus was the exception), and better predicted all four mechanical properties than did avBMD.

#### Discussion

As a surrogate to radiation-based CT, MRI has been developed to characterize trabecular volume and architecture as a means to assess fracture risk [7,8]. For example, such MRI-derived measurements of bone volume fraction and trabecular thickness

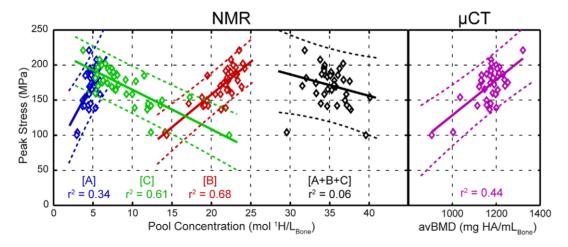


Figure 2. Correlations of measured peak stress and  $T_2$  spectral component amplitudes (NMR, left) and avBMD measured by  $\mu$ CT (right). Blue, red, and green data show integrated amplitudes ( $S_A$ ,  $S_B$ , and  $S_C$ ) of the  $T_2$ -discriminated signals from pools A, B, and C, respectively. The black data show the total <sup>1</sup>H NMR signal ( $S_A+S_B+S_C$ ), and the purple data are derived from  $\mu$ CT-based measures of avBMD. Each of the NMR signals amplitudes shows a significant linear correlation with peak stress and both  $S_B$  and  $S_C$  correlate more strongly with peak stress than does avBMD. Note that the total <sup>1</sup>H NMR signal does not correlate well with peak stress. doi:10.1371/journal.pone.0016359.q002

**Table 1.** A summary of Pearson's r<sup>2</sup> for pairwise correlations between imaging measures (<sup>1</sup>H NMR and X-ray) and mechanical properties.

	Yield Stress	Peak Stress	Flexural Modulus	Pre-Yield Toughness
R <sub>2,A</sub>	0.10	0.12	0.04*	0.12
R <sub>2,B</sub>	0.19	0.22	0.12	0.19
R <sub>2,C</sub>	0.00*	0.01*	0.01*	0.00*
S <sub>A</sub>	0.41	0.34	0.39	0.34
S <sub>B</sub>	0.62	0.68	0.48	0.57
S <sub>C</sub>	0.57	0.61	0.49	0.49
$S_A + S_B + S_C$	0.05*	0.06*	0.06*	0.03*
AVBMD	0.43	0.44	0.46	0.33
POROSITY	0.58	0.60	0.59	0.46

All correlations were significant (p<0.05) *except* those indicated with \*. The imaging measure that was most predictive (highest  $r^2$ ) of each mechanical measure is indicated with boldface type.

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correlated with the compressive strength of human trabecular bone, although the correlations were not as strong as that between CTderived BMD and strength [9]. These MRI techniques do not assess the inherent quality of the bone tissue, and this is a significant shortcoming given the importance of ultrastructural characteristics of the extracellular matrix (e.g., collagen integrity) to the fracture resistance of bone [10]. From ex vivo studies of bone, various quantifications of water by NMR have been correlated with the mechanical competence of bone. In a rabbit model of diet-induced hypomineralization, a <sup>1</sup>H NMR-derived measurement of water content was directly related to the bending strength of cortical bone [11]; however, in a study of ovariectomized and treated mice, only group-mean total water <sup>1</sup>H NMR signal correlated with mechanical properties-no correlation was found across pooled data from 60 bones, which may be explained by the findings of total <sup>1</sup>H signal shown here (Fig. 2). Also, an NMR technique known as "decay from diffusion in an internal field" (DDIF) found an inverse correlation between this NMR-derived pore water parameter and the yield stress of bovine trabecular bone in compression [12], in rough agreement with the present observations of pore-water. Prior to the present study though, only one study attempted to correlate NMR measurements of both pore water and water bound to the

extracellular matrix to the mechanical properties of human bone [13]. That study used  $T_2^*$ -discriminated rather than  $T_2$ -discriminated (used herein) <sup>1</sup>H NMR signals at low static magnetic field, and while a direct relationship existed between the so-called  $T_2^*$ -defined bound water and peak stress, it described a much lower fraction of the peak stress variance ( $r^2 = 0.36$ , compared to 0.68, above). Also, the translation of  $T_2^*$  based discrimination to clinical imaging may be problematic due to the presence of lipid in bone [5,11], and the inability of  $T_2^*$  to discriminate bone <sup>1</sup>H pools at clinical field strengths (no discrimination was found at 4.7T [5] and no discrimination has been reported at clinical fields strengths ( $\geq 1.5$  T)).

Current uTE protocols on human MRI systems use echo times  $<100 \ \mu s$  [14] (and references therein), more than short enough to capture the majority of the bound water signal and some of the collagen proton signal, but the translation of the present findings to clinical MRI will require practical imaging methods of distinguishing these short- $T_2$  signals from the longer- $T_2$  pore water and lipid signals. There are numerous strategies for integrating  $T_2$ selective magnetization preparation into a clinically practical uTEtype sequence [15,16,17], and the optimal approach for bone imaging has not yet been determined. However, Fig. 3 shows two T<sub>2</sub> spectra from one bone specimen. The solid line shows the normal T<sub>2</sub> spectrum, as used in the above analysis, while the dotted line shows the spectrum that results following the complex average of two CPMG signals, with and without the preceding hyperbolic secant radiofrequency (RF) pulse. This RF pulse effectively inverts only the long T2 signals while largely saturating the collagen proton and bound-water signal, so the complex average cancels only the long T<sub>2</sub> signals and results in a net NMR signal that is  $\approx 95\%$  derived from protons with T<sub>2</sub><1 ms. This result demonstrates in principle that a simple RF pre-pulse, which can be readily integrated into a standard uTE pulse sequence, can distinguish pore water from collagen protons and collagen bound water protons in bone. Once implemented on clinical scanners, such an MRI method can then assess both the contribution of structure to whole bone strength as well as the contributions of collagen integrity and porosity, thus proving a more complete assessment of fracture risk than current X-ray based methods.

#### Materials and Methods

#### Human cortical bone processing

The Musculoskeletal Tissue Foundation (Edison, NJ), a nonprofit tissue allograft bank, and the Vanderbilt Donor Program

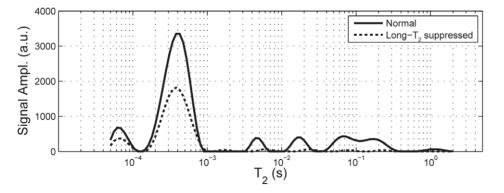


Figure 3. Solid line shows a the  $T_2$  spectrum from a typical bone sample, and the dotted line shows the spectrum that results following the complex average of two signals, with and without an adiabatic full passage magnetization preparation. The total integrated signal from this long- $T_2$  suppressed spectrum is 95% from signals with  $T_2 < 1$  ms, thereby demonstrating in principle a simple and practical method for generating a MRI contrast dominated by  $S_A+S_B$ . doi:10.1371/journal.pone.0016359.g003

(Nashville, TN) supplied human femurs from 40 cadaveric donors (26 male, 14 female, aged 21-105 years old, mean ± standard deviation:  $67\pm24$  years) under instruction to not provide tissue from donors who had tested positive for a blood borne pathogen (e.g., HIV or Hepatitis C). One human cortical bone sample per donor was extracted from the medial quadrant of the mid-shaft and was machined to  $70 \times 5 \times 2$  mm<sup>3</sup> dimensions via end mill. During dimensioning, care was taken to remove endosteal and periosteal surfaces such that the final specimens were pure cortical bone. From each milled sample, three specimens were extracted for NMR, µCT, and mechanical testing (Fig. 4). Specimens were stored in phosphate-buffered saline at  $-20^{\circ}$ C then that  $4^{\circ}$ C for approximately 18 hours prior to NMR measurements. (No more than three freeze-thaw cycles were involved for a given specimen, and separate experiments found that up to six freezethaw cycles had negligible impact on the NMR properties.) Final specimen dimensions were measured with digital caliper for volume determination.

## NMR

From one of the three specimens per donor sample, <sup>1</sup>H NMR transverse relaxation (T<sub>2</sub>) characteristics were measured and reduced to three independent signal components, which we have recently identified as being primarily derived from collagen methylene protons, collagen-bound water protons, and water protons in pores [5]. All NMR measurements were performed in a Varian/Magnex 4.7 T horizontal bore magnet with a Direct Drive Receiver. An in-house loop-gap style RF coil with Teflon structural support was used (similar to the coil described in [18]), which provided 90°/180° RF pulses of ≈8 µs/16 µs duration and contributed negligible background <sup>1</sup>H signal (≈1% of net HCB signal).

Carr-Purcell-Meiboom-Gill (CPMG) measurements with a total of 10000 echoes were collected at 100  $\mu$ s echo spacing, which was empirically determined to be a suitable minimum threshold for both maximizing the range of T<sub>2</sub> detection while minimizing spinlocking effects. Echo magnitudes were fitted to a sum of 128 decaying exponential functions (with time constants log-spaced between 20  $\mu$ s and 10 sec) in a non-negative least-squares sense, subject to a minimum curvature constraint, which produced a socalled T<sub>2</sub> spectrum [19]. In order to quantitatively compare the absolute signal amplitudes of T<sub>2</sub> components across days, a

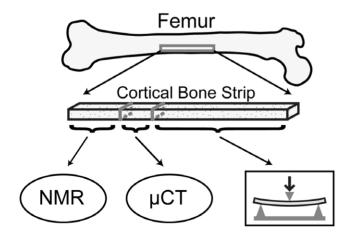
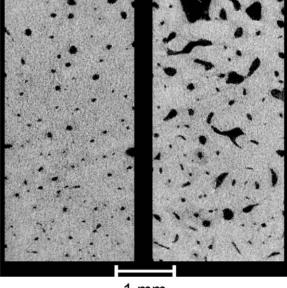


Figure 4. From each cadaveric bone studied, one strip of cortical bone was extracted, three separate pieces of which were used for NMR,  $\mu$ CT, and mechanical testing. doi:10.1371/journal.pone.0016359.g004

reference sample with long  $T_2 (\approx 2 \text{ s})$  and known proton content was included in each CPMG measurement. The presence of the reference sample, together with the known specimen volumes, enabled the calculation of proton concentrations in the bulk bone specimens for each CPMG relaxation component by comparing integrated areas of each  $T_2$  spectral component to the area of the marker. As a simple demonstration of the potential for acquiring signal from a specific  $T_2$  domain without the full CPMG acquisition, from one bone specimen, an additional CPMG measurement was acquired with a preceding a 10-ms duration, 3500 Hz bandwidth hyperbolic secant inversion pulse [20], so chosen to selectively invert the long- $T_2$  <sup>1</sup>H signal.

#### μCT

The second specimen from each donor sample ( $\sim$  volume of 40 mm<sup>3</sup>) was studied at high resolution (6  $\mu$ m), with low noise micro-CT (µCT) to quantify apparent volumetric bone mineral density (avBMD) and intracortical porosity (for pores  $\geq 6 \ \mu m$  in diameter). Note that for a given specimen size avBMD is a volumetric analog to areal BMD as measured by DXA, and intracortical porosity at this resolution is not readily determined from clinical radiographs or QCT including high-resolution peripheral QCT scanners (which obtain resolutions of 80-100 µm) [21]. The specimen was scanned by acquiring 1000 projections per  $180^{\circ}$  at 70 keV using a Scanco, model  $\mu$ CT-40. From an hydroxyapatite (HA) phantom image (acquired weekly), linear attenuation coefficients derived from the  $\mu$ CT images were equated to volumetric bone mineral density (vBMD) in units of mg-HA/cm<sup>3</sup>. Using the Scanco software, the outer perimeter of the sample was defined to determine the total bone volume. The avBMD was defined as the mean of vBMD for all voxels within the total bone volume. The bone tissue volume was segmented from air or soft tissue at a threshold of 800 mg-HA/cm<sup>3</sup> to determine the porosity (=1 minus bone tissue volume per total bone volume) (Fig. 5).



1 mm

Figure 5. Axial  $\mu$ CT images are shown for cortical bone specimens from a 48 y.o. male donor (left) and an 82 y.o. male donor (right). For the 48 and 82 y.o. donors, respectively, avBMD was 1222 and 1135 mg-HA/cm<sup>3</sup>, and porosity was 4% and 11.3%.

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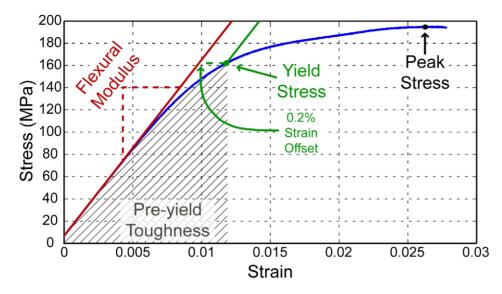


Figure 6. A representative stress vs. strain curve for cortical bone is shown (blue) along with graphical depictions of mechanical parameters. Flexural modulus is the slope of the initial linear mechanical response, yield stress is defined at 0.2% offset from the flexural modulus line, and peak stress is the maximum observed stress. Pre-yield toughness (see text for definition) is proportional to the area under the curve, up to the yield stress.

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#### Mechanical

Finally, we subjected the third, parallelpiped specimen (nominal dimensions of 2 mm×5 mm×40 mm) from each donor sample to a three point bending test, and measured four mechanical properties relevant to fracture risk in bone: yield stress, peak stress, flexural modulus, and pre-yield or elastic toughness. A material testing system (Dynamight 8841, Instron, Canton, OH) recorded the force-displacement data (Fig. 6) from a 100 N load cell and the linear variable differential transformer, respectively, at 50 Hz, as the hydrated bone was loaded to failure at 5 mm/min. The span was 35 mm, and all tests were performed at room temperature. Applying the flexure formula to the yield force, as determined by the 0.2% offset, or to the peak force endured by the

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bone specimen, and applying the deflection equation to the slope of the linear section of the force-displacement curve provided the material properties, yield stress, peak stress, and flexural modulus, respectively [6]. Pre-yield or elastic toughness was the area under the force-displacement curve from zero displacement to the yield displacement divided by the cross-sectional area of the bone sample to account for slight differences in specimen dimensions.

### **Author Contributions**

Conceived and designed the experiments: RAH DFG JSN MDD. Performed the experiments: RAH. Analyzed the data: RAH. Contributed reagents/materials/analysis tools: RAH DFG JSN MDD. Wrote the paper: JSN MDD. Supervised the project: JSN MDD.

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