

RESEARCH ARTICLE

Location Is Everything: Evaluating the Effects of Terrestrial and Marine Resource Subsidies on an Estuarine Bivalve

Joel M. S. Harding^{1,2*}, Michelle R. Segal¹, John D. Reynolds^{1,2}

1 Earth to Ocean Research Group, Department of Biological Sciences, Simon Fraser University, Burnaby, BC, Canada, **2** The Hakai Institute, Heriot Bay, BC, Canada

* joelmsharding@gmail.com



OPEN ACCESS

Citation: Harding JMS, Segal MR, Reynolds JD (2015) Location Is Everything: Evaluating the Effects of Terrestrial and Marine Resource Subsidies on an Estuarine Bivalve. PLoS ONE 10(5): e0125167. doi:10.1371/journal.pone.0125167

Academic Editor: Simon Thrush, University of Auckland, NEW ZEALAND

Received: October 21, 2014

Accepted: March 21, 2015

Published: May 18, 2015

Copyright: © 2015 Harding et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: If accepted, data for this study will be available online at: <http://summit.sfu.ca/item/14866>.

Funding: This research was supported by the Natural Sciences and Engineering Research Council and the Tom Buell BC Leadership Chair endowment funded by the Pacific Salmon Foundation and the BC Leading Edge Endowment Fund. Support was also received from the Tula Foundation, including a scholarship to Joel Harding through the Hakai Network for Coastal Peoples, Ecosystems and Management. Funding sources did not have any role

Abstract

Estuaries are amongst the world's most productive ecosystems, lying at the intersection between terrestrial and marine environments. They receive substantial inputs from adjacent landscapes but the importance of resource subsidies is not well understood. Here, we test hypotheses for the effects of both terrestrial- and salmon-derived resource subsidies on the diet (inferred from stable isotopes of muscle tissue), size and percent nitrogen of the soft-shell clam (*Mya arenaria*), a sedentary estuarine consumer. We examine how these relationships shift across natural gradients among 14 estuaries that vary in upstream watershed size and salmon density on the central coast of British Columbia, Canada. We also test how assimilation and response to subsidies vary at smaller spatial scales within estuaries. The depletion and enrichment of stable isotope ratios in soft-shell clam muscle tissue correlated with increasing upstream watershed size and salmon density, respectively. The effects of terrestrial- and salmon-derived subsidies were also strongest at locations near stream outlets. When we controlled for age of individual clams, there were larger individuals with higher percent nitrogen content in estuaries below larger watersheds, though this effect was limited to the depositional zones below river mouths. Pink salmon exhibited a stronger effect on isotope ratios of clams than chum salmon, which could reflect increased habitat overlap as spawning pink salmon concentrate in lower stream reaches, closer to intertidal clam beds. However, there were smaller clams in estuaries that had higher upstream pink salmon densities, possibly due to differences in habitat requirements. Our study highlights the importance of upstream resource subsidies to this bivalve species, but that individual responses to subsidies can vary at smaller scales within estuaries.

Introduction

Cross-ecosystem resource linkages can structure and stabilize recipient communities [1,2]. Resource linkages, or subsidies, can be driven by abiotic mechanisms [3,4], and biological processes [5,6]. The effects of subsidies can also vary among ecosystems [7,8], individuals [9,10],

in study design, data collection, analysis, decision to publish or manuscript preparation.

Competing Interests: The authors have declared that no competing interests exist.

and with the timing, quality or quantity of resource inputs [10,11]. Interface and hydrologically-linked landscapes such as estuaries have a particularly high potential to benefit from subsidies as upstream resources are conveyed downstream, providing nutrient inputs to these low-lying recipient ecosystems [12–14]. Estuaries are at the intersection of terrestrial, freshwater and marine ecosystems and provide a conduit for the movement of resources among landscapes [1,15]. They are productive, depositional and open ecosystems [16,17], capable of receiving substantial resource inputs from external sources [18–20]. However, the importance of resource subsidies in estuarine ecology remains less clear.

Locally-derived resources within estuaries have been thought to be of primary importance [21,22]. However, more recent work has shown that externally-derived resources can form a major component of available estuarine resources [23–25]. The magnitude of resource subsidy influx can also scale with the size of upstream ecosystems and stream flow [19,26]. Many of the previously mentioned studies have centered largely on the use of stable isotopes to investigate subsidy effects. Although they are a powerful tool in ecology, enabling us to trace resource pathways and relative contributions of potential energy sources [27,28], they are limited beyond confirmation of resource assimilation [11]. Consequently less is known about the biological importance of subsidies in estuaries.

Terrestrial-derived resources are often thought to be of lower quality than estuarine or marine sources [21]. Although they can elevate organic content in estuaries [19,29,30], few studies have attempted to detect productivity responses from them [31–33] and even fewer have tested the degree to which responses can vary across landscapes [26]. Although estuaries are one of the most productive habitats globally [16], this likely varies even at regional scales as a result of the complex interactions with surrounding landscapes.

Around the Northern Pacific Rim, many estuaries also receive pulsed ‘counter-flow’ inputs of enriched marine-derived material from the annual migration of Pacific salmon (*Oncorhynchus* spp.). Because Pacific salmon die in streams after spawning, they can function as one-way nutrient vectors, acquiring the majority of their body mass at sea [34], then transporting this mass back to natal streams. Their carcasses, which are relatively rich in nitrogen and phosphorus, are scattered throughout streams and riparian habitats by predators, scavengers and water flow. Due to the higher trophic level of salmon, and contrasts in biochemistry between marine and terrestrial systems, salmon nutrient subsidies can be differentiated from terrestrial sources using stable isotope ratios of nitrogen and carbon ($\delta^{15}\text{N}$ and $\delta^{13}\text{C}$); where salmon-derived material is enriched and terrestrial-derived material is depleted in heavy isotopes [26,27,35]. The net effects of Pacific salmon in coastal ecosystems can vary [26,36], ranging from nutrient subsidies through excretion and deposition of eggs and carcasses [35,37,38], to benthic disturbance and nutrient export from juvenile salmon emigration and adults digging and defending nests [39–41]. In addition to streams and forests, estuaries also receive substantial amounts of salmon-derived nutrients from upstream watersheds [42–44]. Although dissolved nutrient concentrations increase in estuaries during salmon spawning [44,45] and many estuarine organisms are known to consume carcasses [46], few studies have investigated the importance of salmon subsidies in these communities [26,44,47].

Sedentary consumers such as bivalves not only provide an opportunity to investigate the importance of terrestrial- and salmon-derived resource subsidies in estuarine food webs, but also how these relationships might change spatially within, and across, landscapes. Bivalves integrate isotopes over time and can thus reveal resource contributions in relation to proximity of resource inputs [22,48,49]. Suspension feeders such as the soft-shell clam (*Mya arenaria*) are widespread in estuaries of the Pacific Northwest and have recently been shown to assimilate terrestrial-derived resources [50]. Similar to the river continuum concept of Vannote et al. [51], we hypothesize that estuarine organisms, such as the soft-shell clam, are influenced by

Table 1. Hypotheses for soft-shell clam stable isotope ratios ($\delta^{15}\text{N}$ and $\delta^{13}\text{C}$).

Variable	Mechanism	Metric	Level	Response	Reference
Salmon density	Salmon tissues are enriched in stable isotopes.	2006–2007 mean pink salmon biomass density (kg m^{-2})	Site	Positive	[37,38,47,49,96]
Watershed size	Terrestrial-derived material is depleted in stable isotopes.	Watershed size principal component axis 1 (PC1)	Site	Negative	[19,48–50,97]
Size	Larger individuals grow more slowly and have slower tissue turnover rates, which reflect dietary sources over longer time periods.	Mass (g)	Individual	Positive	[98]
Age	Older individuals have more time to accumulate stable isotope ratios from enriched dietary sources.	Age (years)	Individual	Positive	[85,99]
Intertidal height	Individuals higher in intertidal will have lower isotopic discrimination as a result of more limited feeding opportunities.	Height above datum depth (m)	Within-site	Positive	[100,101]
Temperature	Energy requirements for maintenance and growth increase with temperature, reducing isotopic discrimination.	Maximum weekly average temperature (MWAT $^{\circ}\text{C}$)	Site	Positive	[101]
Clam bed zone	1) Moving outward from upper to lower zones (increasing distance from stream outlet) will reduce the effect of both salmon density and watershed size. 2) Moving outward from upper to lower zones will also correspond with increased dominance of marine resources and enrich isotopes.	Upper, middle and lower clam bed locations.	Within-site	1) Negative (in interaction with salmon and watershed size). 2) Positive as main effect.	[44,47–49]
Location	1) The effect of salmon and watershed size will increase going from control to below stream locations. 2) Clams below streams will experience increased influx of terrestrial resources, and therefore have depleted isotopes, compared to control locations.	Below stream and control sites.	Within-site	1) Positive (in interaction with salmon and watershed size). 2) Negative as main effect	[44,47–49]

doi:10.1371/journal.pone.0125167.t001

resources derived from both upstream and marine landscapes, and that the importance of these resources will vary spatially with landscape traits. We further hypothesize that responses of sedentary consumers can vary based on their proximity to resource subsidies and local habitat conditions.

Here, we test hypotheses on how terrestrial and salmon resource subsidies, in addition to individual traits, explain the diet (inferred from stable isotopes of nitrogen and carbon), size, and percent nitrogen of soft-shell clams (Tables 1 and 2). We test for the effects of these covariates across 14 estuaries that span natural gradients in watershed size, spawning salmon density and other attributes (Table 3). Prior to our main analyses we tested metrics of chum (*Oncorhynchus keta*), pink (*O. gorbuscha*) and total (chum and pink combined) salmon density for their ability to explain isotope ratios of soft-shell clam muscle tissue. We hypothesized pink salmon may have a disproportionately large effect on bivalves because they spawn further downstream than chum salmon, including upper reaches of estuaries, and thus closer to bivalve habitats.

Study Area

We studied estuaries within 45 km of Bella Bella (52°9'N, 128°8'W) on the central coast of British Columbia, Canada (Fig 1). This region lies within the Coastal Western Hemlock biogeoclimatic zone and receives some of the highest levels of precipitation on the continent [52].

Although selective logging occurred in many areas during the first half of the 20th century, this

Table 2. Hypotheses for soft-shell clam size and tissue %N.

Variable	Mechanism	Metric	Level	Response	Reference
Salmon density	Salmon tissues are higher quality than other sources, resulting in larger individuals and higher N content in tissues.	2006–2007 mean pink salmon biomass density (kg m ⁻²)	Site	Positive	[35,42,73,76,89,102]
Watershed size	Terrestrial-derived material can enhance organic content in estuaries, which could increase clam size and %N in tissues. It is also thought to be of lower quality and may displace higher-quality estuarine resources, reducing size and %N.	Watershed size principal component axis 1 (PC1)	Site	Positive/no effect/ negative	[19,21,29,49,75,76]
Size (for %N only)	Larger individuals grow more slowly and have slower tissue turnover rates, which will reflect higher-quality dietary sources over longer time periods.	Mass (g)	Individual	Positive	[73,89]
Age	1) Size: Older individuals are larger. 2) %N: Younger individuals grow faster, resulting in higher percentages of nitrogen in their tissues.	Age (years)	Individual	1) Positive (for size) 2) Negative (for %N)	[73,89,103]
Intertidal height	Individuals located higher in intertidal will have limited feeding opportunities This should result in smaller sizes and reduced N content (energy stores) in tissues.	Height above datum depth (m)	Within-site	Negative	[100,101]
Temperature	Energy requirements for maintenance and growth increase with temperature, reducing opportunity for growth and energy stores.	Maximum weekly average temperature (MWAT°C)	Site	Negative	[101]
Clam bed zone	1) Moving outward from upper to lower zones (increasing distance from stream outlet) will reduce the effect of both salmon density and watershed size on mass and % N. 2) Moving outward from upper to lower zones will correspond with an increase in size and %N as marine resource availability increases.	Upper, middle and lower clam bed locations.	Within-site	1) Negative (in interaction with salmon and watershed size). 2) Positive as main effect	[104,105]
Location	1) The effect of salmon and watershed size will increase going from control to below stream locations. 2) Clams below streams will be smaller and have less %N compared to control locations as a result of shifting from marine- to terrestrial-dominated resources. However, reduced habitat quality in control sites may offset this effect.	Below stream and control sites.	Within-site	1) Positive (in interaction with salmon and watershed size). 2) Positive/ Negative as main effect	[104,105]

doi:10.1371/journal.pone.0125167.t002

region remains relatively intact due to its remoteness, restricted access and strengthening First Nations governance and conservation coalitions [53]. This remote region provides access to a wide range of relatively pristine watersheds that are ideal systems to test for the effects of terrestrial and salmon resource subsidies in estuaries.

We sampled 14 small- to medium-sized estuaries, which hosted soft-shell clam populations and varied in upstream catchment area, stream channel size, estuary area, upstream salmon spawning density, and red alder (*Alnus rubra*) dominance (Table 3). All streams were dominated by chum (*O. keta*) and pink (*O. gorbuscha*) salmon, which accounted for 90–100% of total adult salmon spawners, with much smaller numbers of coho (*O. kisutch*) and a limited presence of sockeye (*O. nerka*) and Chinook salmon (*O. tshawytscha*). Salmon spawn in streams throughout BC’s central and north coasts, which can produce over half of the wild salmon stocks in this region, and account for over 30% of total populations within BC and the Yukon [54].

Methods

Ethics Statement

Salmon counts and measurements were conducted to meet the requirements of the Canadian Council on Animal Care (approval number 1031B-11). Bivalve sampling and associated protocols at all locations were approved by Fisheries and Oceans Canada (Scientific Licence numbers XHAB 318 2008; XMCFR 11 2009).

Sampling

We collected soft-shell clams during the summers of 2008 and 2009 prior to salmon spawning. Samples were collected during tide heights less than, or equal to 1m above chart datum depth (0 m tidal height). Depths of sample locations ranged between 0.47 and 2.1 m above chart datum. At each site three systematic locations were sampled representing upper, middle, and lower zones of the clam bed spanning the vertical width of the clam bed (Fig 2). These three zones were sampled directly below stream outlets and adjacent to the main channel within each estuary tidal flat. At each location, 5 soft-shell clams were sampled haphazardly by digging to a depth of 30 cm at each sample location and piling the sediment on the beach surface. The excavated sediment was then searched where we retained the first 5 clams encountered. This method helped reduce depth biases in sampling smaller clams in surficial sediments. Additional holes were excavated adjacent to the original if fewer than 5 clams were present. For each clam collected, we immediately recorded shell length, width, depth and wet weight. Clams were then wrapped in aluminum foil and frozen at -20°C in sealed containers until processing. Sampling time and height above water were recorded for each location to enable depth corrections to chart datum. Height above water was measured by viewing a metre stick, located at the water's edge, through a clinometer from each sample location. The height above water was equal to the height on the metre stick, at zero degrees, minus the height of the observer's viewpoint. In 2009, additional within-site control locations were sampled laterally down shore from stream outlets and outside the depositional deltas of each estuary (Table 3). These control locations were located in 9 of our 14 study sites and limited to the upper and lower clam bed zones (Fig 2). Age data were collected by sectioning shell chondrophores (encased in Loctite Hysol epoxy) using a Buehler Isomet Low-speed saw with diamond wafering blades. Chondrophore sections were mounted on glass slides and polished sequentially with 30, 9 and 3 micron lapping film. Sections were aged by counting annual growth lines following the methods of MacDonald and Thomas [55] using light manipulations and a digital camera mounted to a dissecting microscope.

Watershed Data

Stream and riparian canopy (% alder) data were collected during the summer of 2007 as part of an extensive survey in the region. Temperature was measured continuously using waterproofed temperature loggers (iButtons DS1922L) anchored to rebar below chart datum depth (0m tide) and set to record every 2 hours spanning the study period. Stream measurements occurred at 12 randomly selected transects along a study reach equal to 30 times the mean bankfull width of each stream [56]. Alder basal area was estimated by measuring the diameter at breast height of all trees greater than 5 cm in diameter within six belt transects that extended perpendicular from each stream and were 35 m long by 10 m wide [35]. Percent alder was calculated for each site as:

$$\%A = \frac{B_{alder}}{B_{total}} \times 100$$

Table 3. Site-level covariates used to create watershed size PC1 (catchment area, bankfull, depth and bank height), percent alder, pink salmon density, temperature and distances between clam sampling locations.

Site	Catchment area (km ²)	Mean bankfull width (m)	Mean depth (m)	Mean bank height (m)	Watershed Size PC1	Percent riparian alder	Mean 2006–07 pink salmon adult biomass density (kg/m ²)	Maximum weekly average temperature (°C)	Distance between clam bed zones below streams (m)	Distance between zones in control locations (m)	Lateral distance between control and below stream locations (m)	Latitude	Longitude
Ada	10.1	11.1	0.12	0.34	-0.91	3.26	0.047	16.00	1 location	NA	NA	52.0553	-128.0507
Bullock Main	3.3	10.9	0.08	0.26	-2.18	3.31	0.078	19.93	35	70	140	52.4029	-128.0785
Clatsse	32.1	22.8	0.16	0.30	0.53	26.08	0.264	23.38	102.5	205	280	52.3455	-127.8476
Codville	2.4	3.3	0.18	0.24	-2.50	0.00	0.004	18.92	15.5	NA	NA	52.0790	-127.8633
Fannie Left	35.0	12.8	0.16	0.39	0.39	1.74	0.090	18.57	57.5	115	60	52.0426	-128.0668
Fell Creek	7.0	10.9	0.19	0.41	-0.38	1.16	0.229	21.41	1 location	1 location	74	52.4336	-128.0790
Hooknose	18.4	16.9	0.18	0.46	0.67	3.08	0.057	18.88	56.5	113	155	52.1249	-127.8370
Kunsoot Main	5.7	13.1	0.04	0.22	-2.20	0.00	0.259	17.16	35	NA	NA	52.1569	-128.0435
Mosquito Bay	5.2	9.7	0.11	0.21	-1.84	6.33	0.081	20.48	10	1 location	70	52.3968	-128.1660
Neekeas	17.6	17.7	0.16	0.40	0.33	13.35	0.413	22.84	85	170	70	52.4509	-128.1569
Quartcha	40.9	34.1	0.24	0.55	3.28	17.95	0.010	18.77	82	1 location	375	52.5155	-127.8421
Rainbow	13.7	15.1	0.23	0.47	0.77	20.34	0.001	24.86	35	NA	NA	52.4512	-127.7280
Roscoe Main	33.6	23.5	0.28	0.56	2.70	54.77	0.000	24.63	62.5	125	240	52.4696	-127.7448
Sagar	36.6	15.5	0.25	0.43	1.34	0.21	0.013	18.13	1 location	NA	NA	52.0959	-127.8388

doi:10.1371/journal.pone.0125167.t003

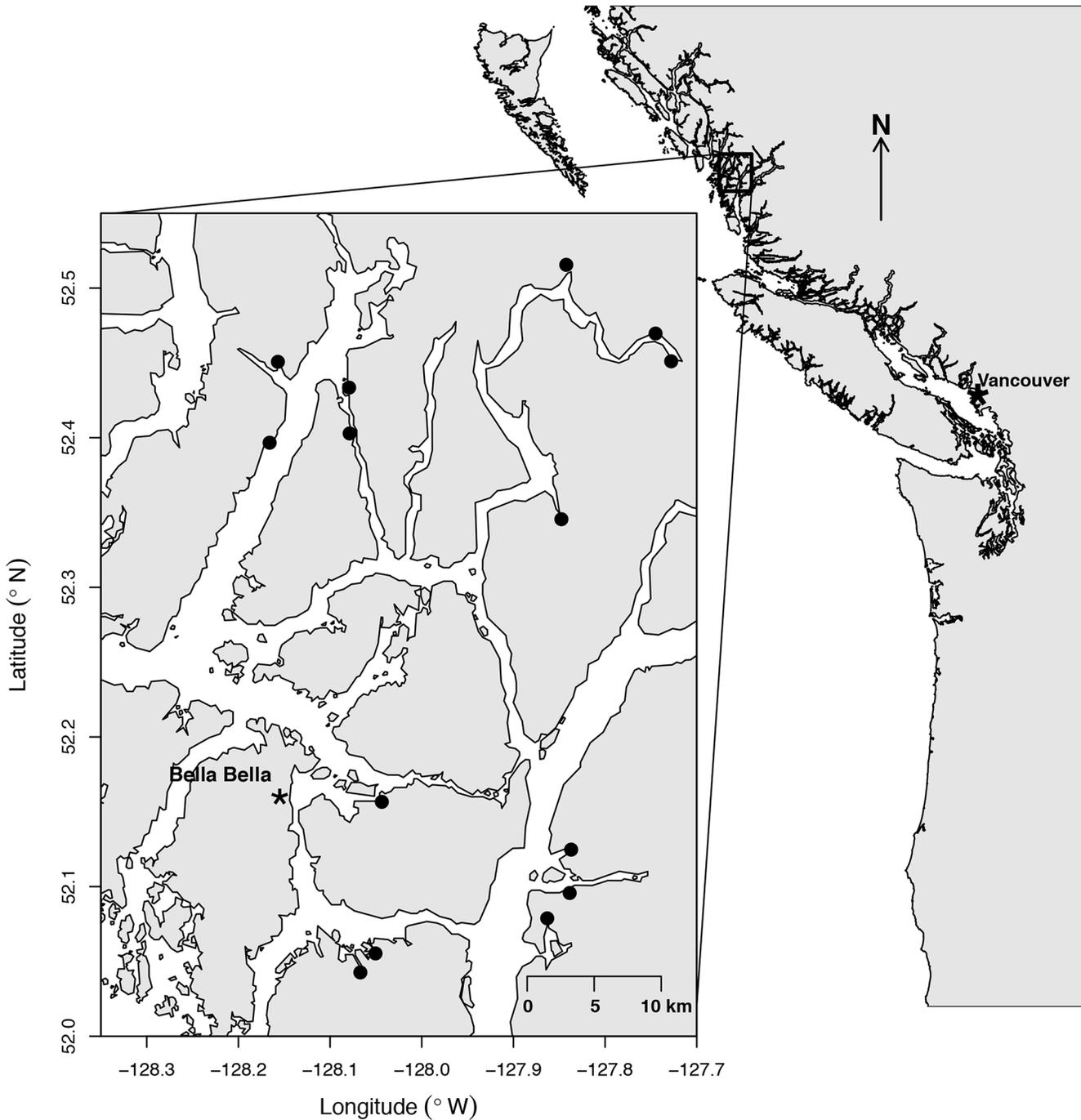


Fig 1. Study area in the vicinity of Bella Bella, on British Columbia's central coast.

doi:10.1371/journal.pone.0125167.g001

where %A is the percent alder for each site, B_{alder} is total basal area of all alder measured in a given site and B_{total} is the total basal area of all tree species measured in that site. Watershed catchment areas were estimated using the Government of British Columbia's mapping website *iMapBC* [57].

Principal components analysis (PCA) was used to generate a composite variable to describe watershed size to approximate the magnitude of stream discharge and amount of terrestrial-

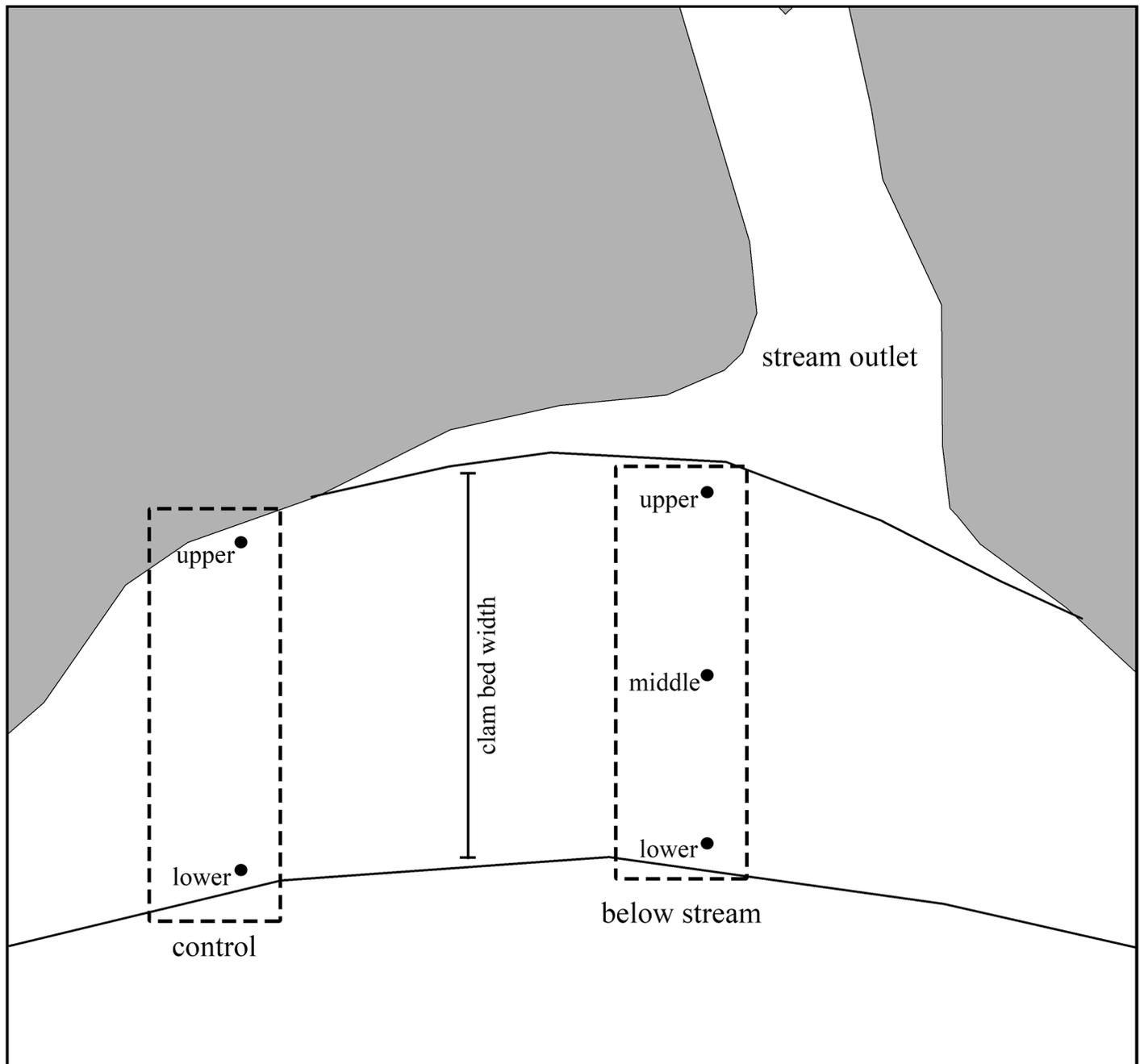


Fig 2. Sampling design. Upper, middle and lower clam bed zones were sampled below streams in 2008 and 2009, upper and lower zones were sampled in control locations in 2009.

doi:10.1371/journal.pone.0125167.g002

resource influx into estuaries. Component variables included total catchment area (km^2), mean stream bankfull width (width of the stream channel at its highest point before flooding), mean stream depth, and mean stream bank height (maximum stream depth before flooding). Pearson correlation coefficients of component variables ranged between 0.7 and 0.9. The first principal component axis (PC1) described 80% of component variable variances and variables all loaded positively on this axis ranging between 0.48 and 0.52. The PC1 axis values reflect both

the capacity of streams to transport nutrient subsidies into estuaries (stream channel measurements) and the amount of terrestrial-derived nutrient sources upstream (catchment area).

Salmon Population Data

The federal Department of Fisheries and Oceans, the Heiltsuk Integrated Resource Management Department, and Simon Fraser University cooperatively conducted all salmon enumeration and spawning channel measurements. We considered upstream salmon biomass density estimates between 2006 and 2009 as potential proxies for salmon carcass availability in estuaries downstream [26]. We determined this from data limitations (data collection began in 2006 and we did not want to consider years beyond 2009). Salmon biomass density indices were calculated for chum salmon, pink salmon and chum and pink salmon combined, for year combinations 2006–2007, 2006–2008 and 2006–2009 for each site:

$$SBD_{ij} = \frac{\sum (N_{ij} \times W_j)}{A}$$

where SBD_{ij} = average kg of salmon biomass per m^2 of spawning area per stream for year combination i and species j , N_{ij} = the mean number of returning adult salmon for year combination i and species j , W_j = average salmon mass for each species j , and A = the estimate of spawning area (m^2) within each stream. Spawning area was estimated by multiplying the mean bankfull width by the total spawning channel length for each stream. We accounted for variation in salmon body mass among populations by measuring the weight of 5 dead adult salmon of each sex for each species in a subset of study streams covering our study area. These average salmon masses were applied to the remaining study sites sharing island groups, channels or mainland inlets. We limited our analyses to chum and pink salmon because these species account for 90–100% of total adult salmon in our study region.

We conducted an initial exploratory analysis to identify the best salmon density metric that explained stable isotope ratios of clam foot muscle tissue. We constructed univariate linear models with chum, pink or total (chum and pink) salmon density for each selected year combination explaining $\delta^{15}N$ or $\delta^{13}C$. We competed these models using Akaike Information Criterion corrected for small sample sizes (AICc) that selects for the most parsimonious model given the data. We log transformed all salmon density metrics in all analyses to reduce the leverage of high salmon density values on slope estimates.

Stable Isotope Analysis

Foot muscle tissue samples for isotope analysis were removed from thawed samples and placed in a drying oven at 58°C for up to 96 hours. Each sample was homogenized into a fine powder using a heavy duty Wig-L-Bug grinder (Pike Technologies Ltd). Sample weights ranging between 0.8–1.2µg were packaged in standard pressed tin capsules (3.5 x 5 mm) and sent to the UC Davis Stable Isotope Facility for analysis of nitrogen and carbon abundance using a PDZ Europa ANCA-GSL elemental analyzer interfaced to a PDZ Europa 20–20 isotope ratio mass spectrometer (Sercon Ltd., Cheshire, UK). Stable isotopes are expressed as the difference between the sample and a known standard, or δ , in parts per thousand (‰):

$$\delta^{15}N \text{ or } \delta^{13}C = \left(\frac{R_{sample}}{R_{standard}} - 1 \right) \times 1000$$

where R is the ratio of the heavy isotope to the light isotope ($^{15}N/^{14}N$ or $^{13}C/^{12}C$).

Standards for nitrogen and carbon analysis are derived from N_2 in air and Pee-Dee Belemnite (PDB) limestone, respectively.

Percent nitrogen of soft-shell clam muscle tissue was calculated as:

$$\%N = \frac{N}{T} \times 100$$

where N is the mass of nitrogen in the sample and T is the total mass of the sample.

Statistical Analyses

Bivalve mass was chosen as the most ecologically meaningful metric representing an individual's size [58]. We used the open source statistical software R for all analyses [59]. Variance inflation factors (VIF) of all covariates were less than 2.2 and thus indicated low multicollinearity [60]. Pearson correlation coefficients between individual covariates were 0.6 or less and not of great concern [61]. The only exception was % alder, which had a VIF of 4.3 and Pearson correlation coefficients of approximately 0.8 with both watershed size and temperature. Due to this high collinearity % alder was removed from all analyses.

For all analyses (isotopes, mass and %N) we used linear mixed-effects modeling to account for the hierarchical structure of the data [62]. This method allowed regression intercepts to vary by site (site as random intercept), accounted for potential correlation of individuals from the same site between sample years (correlation structure of site within year for all analyses), and accounted for heterogeneity in the residual variance structure [60]. Correlation and variance structures were established from residuals of the global models, or models including all variables considered, and AICc selection of the most parsimonious structures with the global model using restricted maximum likelihood (REML) estimation [60]. Variance structures on datum depth and control/below stream covariates improved the likelihood of the global models and satisfied the assumptions of residual normality and equal variance for mass and %N analyses respectively [60]. No variance structures were required for isotope analyses as the assumptions of equal variance were already met. We include a pseudo- R^2 value for the averaged model from each analysis. This is the R^2 value for a linear model between the fitted values of the averaged model and the observed data. We conducted an additional analysis on an approximation of clam growth that we calculated as individual clam mass divided by age, or the average mass acquired per year. Results were very similar to our analysis of clam mass so we chose to not include it to avoid redundancy.

We wanted to test how the effects of salmon and watershed subsidies could vary by distance from stream outlets (upper, middle and lower clam bed zones) and by location (control vs. below stream). We therefore constructed our models to include the following interactions in all analyses; salmon and zone, salmon and location, watershed size and zone and watershed size and location. We competed models of all combinations of covariates in addition to the specified interactions because we did not have any *a priori* reason to exclude any models from the analyses [63]. For each analysis we conducted two model competitions, first using centered covariates (subtracting the mean) and again using scaled covariates (centering and dividing by 2 standard deviations). All covariates were centered to avoid inaccuracies in slope estimates for main effects as they can vary considerably depending on the presence of interaction terms [64]. We also analyzed models with scaled covariates to enable direct comparison of effect sizes amongst variables [65,66]. In all analyses, $k-1$ binary dummy variables were created for the three-level zone factor (upper, middle, lower) and 2-level location factor (control, below stream), where k is the number of levels in a factor following the methods of Schielzeth [64]. In standardized models, dummy variables were not divided by 2 standard deviations as slope estimates from binary variables already relate to 2 standard deviations (comparisons of 0 and 1) [66]. Model competition using AICc revealed that top model weights in all analyses were less

than 0.22. We accounted for this model uncertainty using a multi-model approach [67]. Candidate models used in multi-model inference were limited to the subset of models with a ΔAIC_c less than 4 [63] and estimates for each covariate and interaction term were averaged across candidate model sets using the natural average method. Intercepts, slopes, scaled coefficients and standard errors for the combined effects of salmon and watershed size at each zone and location level were calculated from averaged model outputs. The equations used to calculate these combined effects from interactions are presented in the supplemental information.

Results

Sampling

A total of 154 and 243 soft-shell clams were sampled in 2008 and 2009 respectively, from 14 sites each year. Clam mass ranged between 1.3–116.9 g and 3.2–126.6 g in 2008 and 2009, respectively. In 2009, control samples were collected from 9 of the 14 sites (Fig 2). Bedrock and small estuary sizes prevented control sampling from the remaining sites. Table 3 provides a summary of site-level covariates and distances between sampling locations.

Salmon Metric Pre-selection

The 2006–07 mean pink salmon density explained the most variation in both $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ of soft-shell muscle tissue with model weights exceeding 0.8. This salmon metric was used in all subsequent analyses. Model rankings are presented in S1 Table.

Clam Isotopes

$\delta^{15}\text{N}$. The pseudo- R^2 of the averaged model was 0.66. Following our predictions, clams in estuaries with higher upstream salmon densities had enriched $\delta^{15}\text{N}$. In addition, the effect of salmon decreased going from upper to lower clam beds (Fig 3A). Clams below streams with large watersheds were more depleted in $\delta^{15}\text{N}$, but this was only detected in the lower zones (Fig 3B). Clams that were higher on shore (higher above chart datum), and those that were larger and older had enriched $\delta^{15}\text{N}$, following our predictions (Fig 3C–3E). Temperature did not describe $\delta^{15}\text{N}$. The standardized effects of salmon were positive at all zone and location levels with confidence intervals well above 0 (Fig 4A). The effects of watershed size were more variable, with confidence intervals crossing 0 with the exception of lower clam beds (Fig 4A). Age, mass and height above chart datum were all positive and highly certain while the effects of temperature were small and high a higher degree of uncertainty (Fig 4A). The averaged $\delta^{15}\text{N}$ model and candidate set are presented in S2 and S3 Tables.

$\delta^{13}\text{C}$. The pseudo- R^2 of the averaged model was 0.23. Clams below large watersheds were more depleted in $\delta^{13}\text{C}$ and, as we predicted, this depletion was strongest in the locations below streams (Fig 5A) but also in the lower zones compared to the upper and middle zones (Fig 5B), which did not support our predictions. Clams were enriched in $\delta^{13}\text{C}$ below streams with higher pink salmon densities and this effect was stronger in the upper and middle zones (Fig 5C). Contrary to our predictions, warmer estuaries had clams with more depleted $\delta^{13}\text{C}$ (Fig 5D). Larger and older clams were enriched in $\delta^{13}\text{C}$ (Fig 5E and 5F) but height above chart datum did not have any effect (Fig 4B). Similar to $\delta^{15}\text{N}$, the standardized effects of salmon on $\delta^{13}\text{C}$ were positive at all zone and location levels with confidence intervals above 0 (Fig 4B). The standardized effects of watershed size were strongest in the upper and lower zones, and below stream locations but less certain in middle zones and control locations (Fig 4B). The effects of age, mass and temperature had a high degree of certainty around coefficient estimates (Fig 4B). The averaged $\delta^{13}\text{C}$ model and candidate set are presented in S4 and S5 Tables.

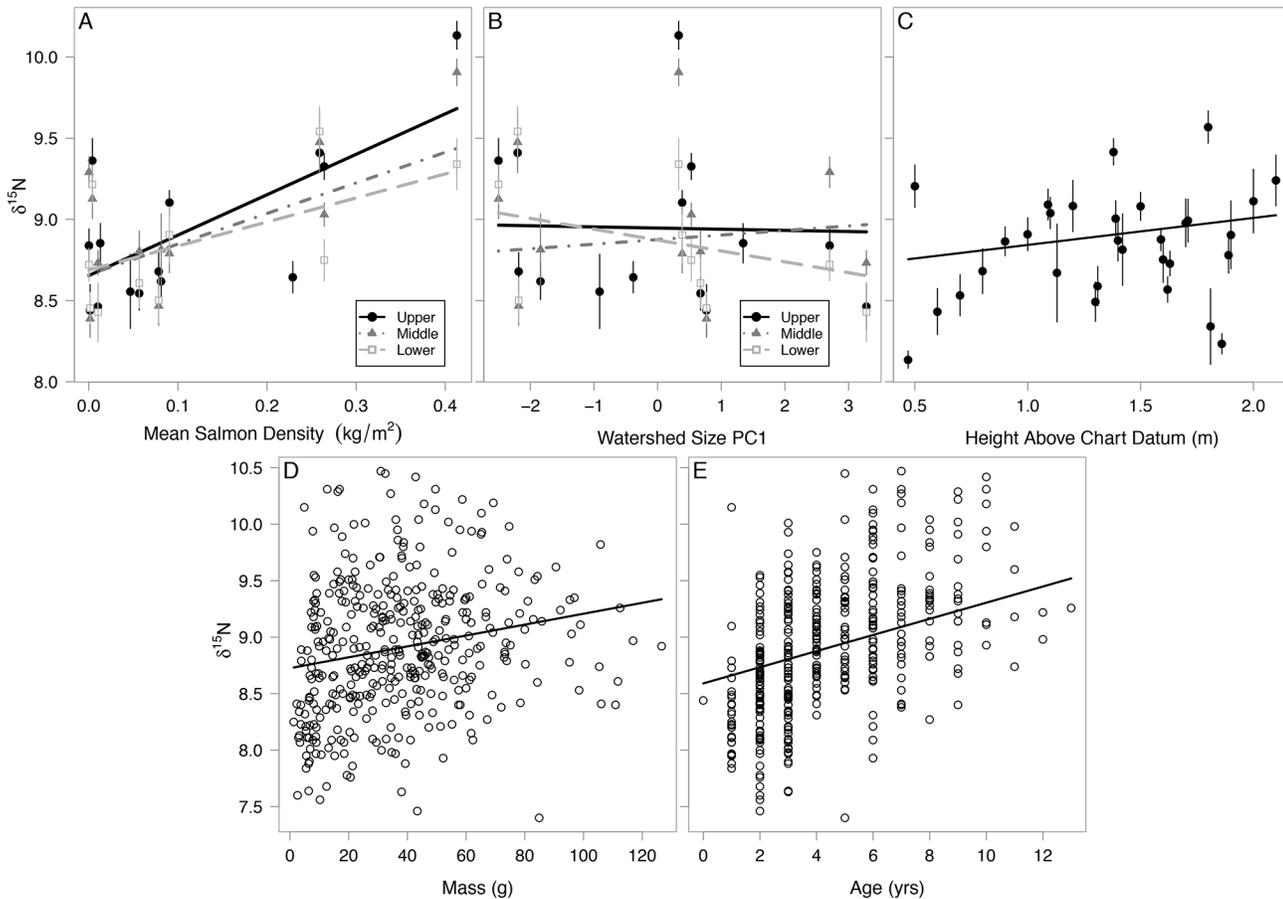


Fig 3. Correlates of soft-shell clam muscle tissue $\delta^{15}\text{N}$. (A) Pink salmon density at each clam bed zone, (B) Watershed size PC1 at each clam bed zone, (C) Height above datum depth, (D) Clam mass, and (E) Clam age. Each data point in panels A-C represents mean values with standard error bars. Data points in panels D-E represent individual clams. All trend lines represent relationships using intercept and coefficients from multi-model output; thus they represent the relationships for the x-axis variable that accounts for the effects of other variables, rather than fitting the univariate data shown in each graph.

doi:10.1371/journal.pone.0125167.g003

Size

The pseudo- R^2 of the averaged model was 0.62. There were larger clams below larger watersheds but this effect was restricted to below stream locations (Fig 6A). Watershed size showed the opposite, and negative, correlation with size in control locations (Figs 6A and 7A). The positive correlation between clam size and watershed size was strongest in lower clam beds, and below stream locations where confidence intervals did not cross 0 (Fig 7A). The effect of location (below stream vs. control) was not an important descriptor of clam mass on its own. Surprisingly, salmon correlated negatively with clam size, opposite to our predictions, though the correlation with salmon in the below stream locations was less negative (Figs 6B and 7A). This negative relationship was observed at all zone and location levels, where most of the confidence intervals did not cross 0 (Fig 7A). Clams were also slightly smaller in the upper, compared to middle and lower clam beds (Fig 6C), and clams that were higher above chart datum and younger were smaller (Fig 6D and 6E). The effects of age and height above chart datum were positive and negative respectively, with a higher degrees of certainty, while the effects of temperature were negligible. The averaged clam size model and candidate set are presented in S6 and S7 Tables.

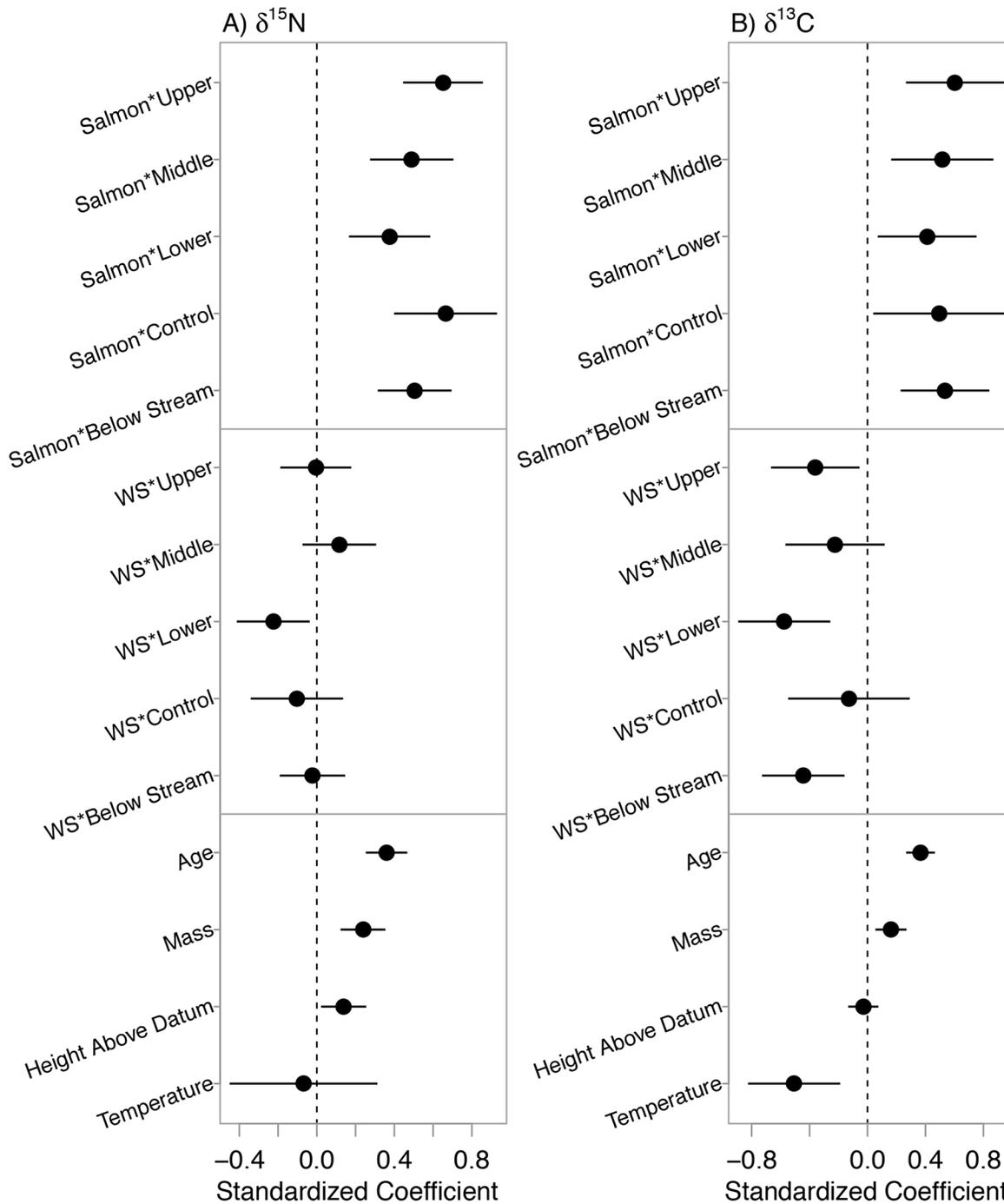


Fig 4. Standardized coefficients (mean = 0, standard deviation = 2) with 95% confidence intervals for covariates considered in the (A) $\delta^{15}\text{N}$ candidate model set and (B) $\delta^{13}\text{C}$ candidate model set. Salmon = 2006–07 mean pink salmon density; WS = watershed size PC1. Coefficient values indicate the change, on average, in $\delta^{15}\text{N}$ or $\delta^{13}\text{C}$ as the associated covariates increase by 2 standard deviations.

doi:10.1371/journal.pone.0125167.g004

Percent N

The pseudo- R^2 of the averaged model was 0.21. Clams below larger watersheds had higher percentages of N in their muscle tissues (Fig 8A). Contrary to our predictions, clams in upper

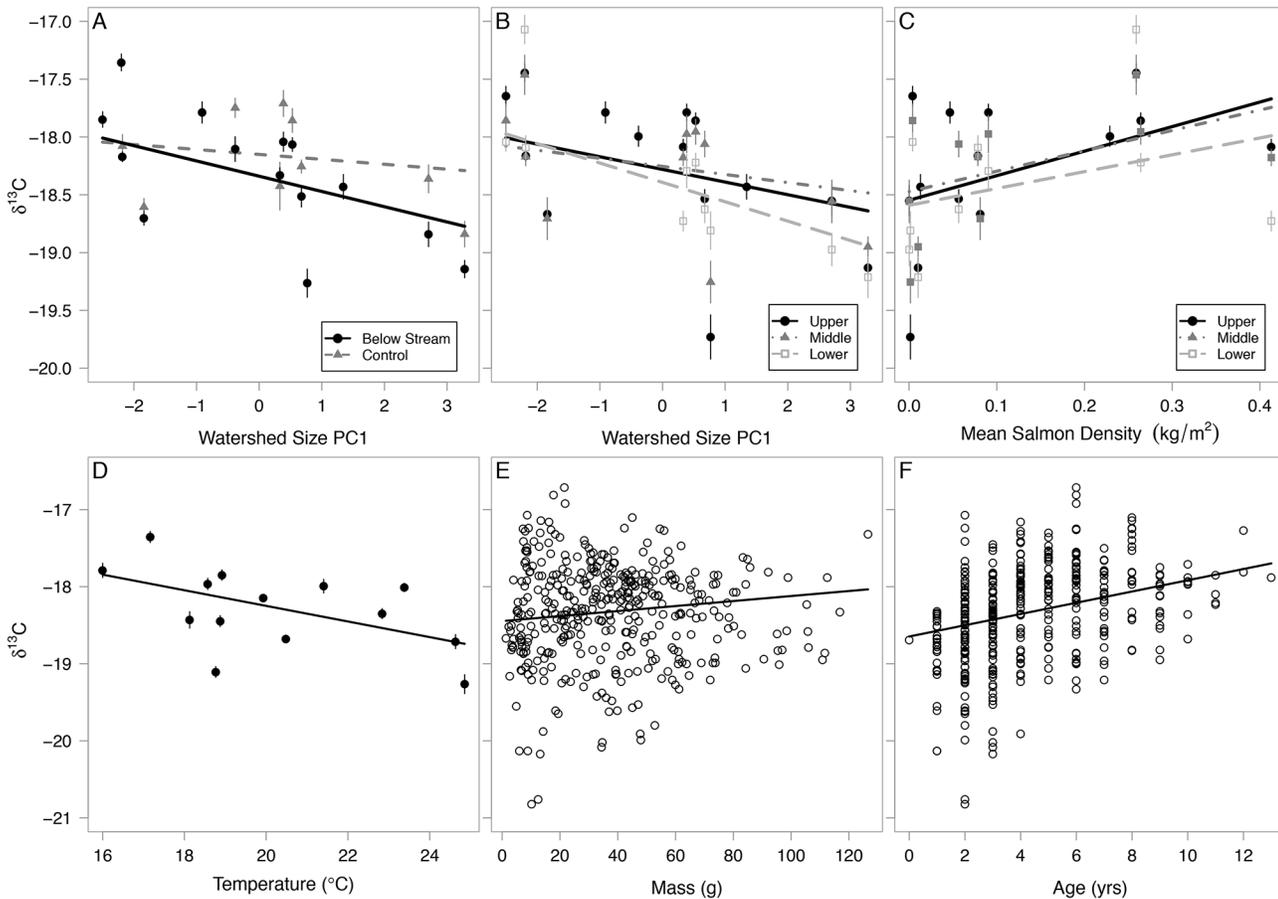


Fig 5. Correlates of soft-shell clam muscle tissue $\delta^{13}\text{C}$. (A) Watershed size PC1 at below stream vs. control locations, (B) Watershed size PC1 at each clam bed zone, (C) Pink salmon density at each clam bed zone, (D) Temperature (maximum weekly average temperature), (E) Clam mass, and (F) Clam age. Each data point in panels A-D represents mean values with standard error bars. Data points in panels E-F represent individual clams. All trend lines represent relationships using intercept and coefficients from multi-model output; thus they represent the relationships for the x-axis variable that accounts for the effects of other variables, rather than fitting the univariate data shown in each graph.

doi:10.1371/journal.pone.0125167.g005

zones contained higher %N in their tissues than their counterparts (Fig 8B). Analysis did not detect an influence from any other covariates including salmon density, temperature, clam size and age. Although zone and location did not have any interaction effects with salmon density or watershed size, clams below streams had higher %N than those in control locations (Fig 8C) and this disparity was most apparent in lower clam beds (Fig 8D). The standardized effects of salmon on %N were negative but highly uncertain at all zone and location levels with confidence intervals crossing 0 (Fig 7B). The effects of watershed size were positive, particularly in upper clam beds and below streams and confidence intervals did not cross 0, with the exception of control locations. Clams higher above chart datum had elevated %N with confidence intervals well above 0 while all remaining covariates had undetectable effects (Fig 7B). The averaged %N model and candidate set are presented in S8 and S9 Tables.

Discussion

This study demonstrates the complex relationships between terrestrial and marine resource subsidies and traits of sedentary estuarine consumers. It highlights how cross-ecosystem resource linkages can vary both within and across landscapes. Watershed size and salmon

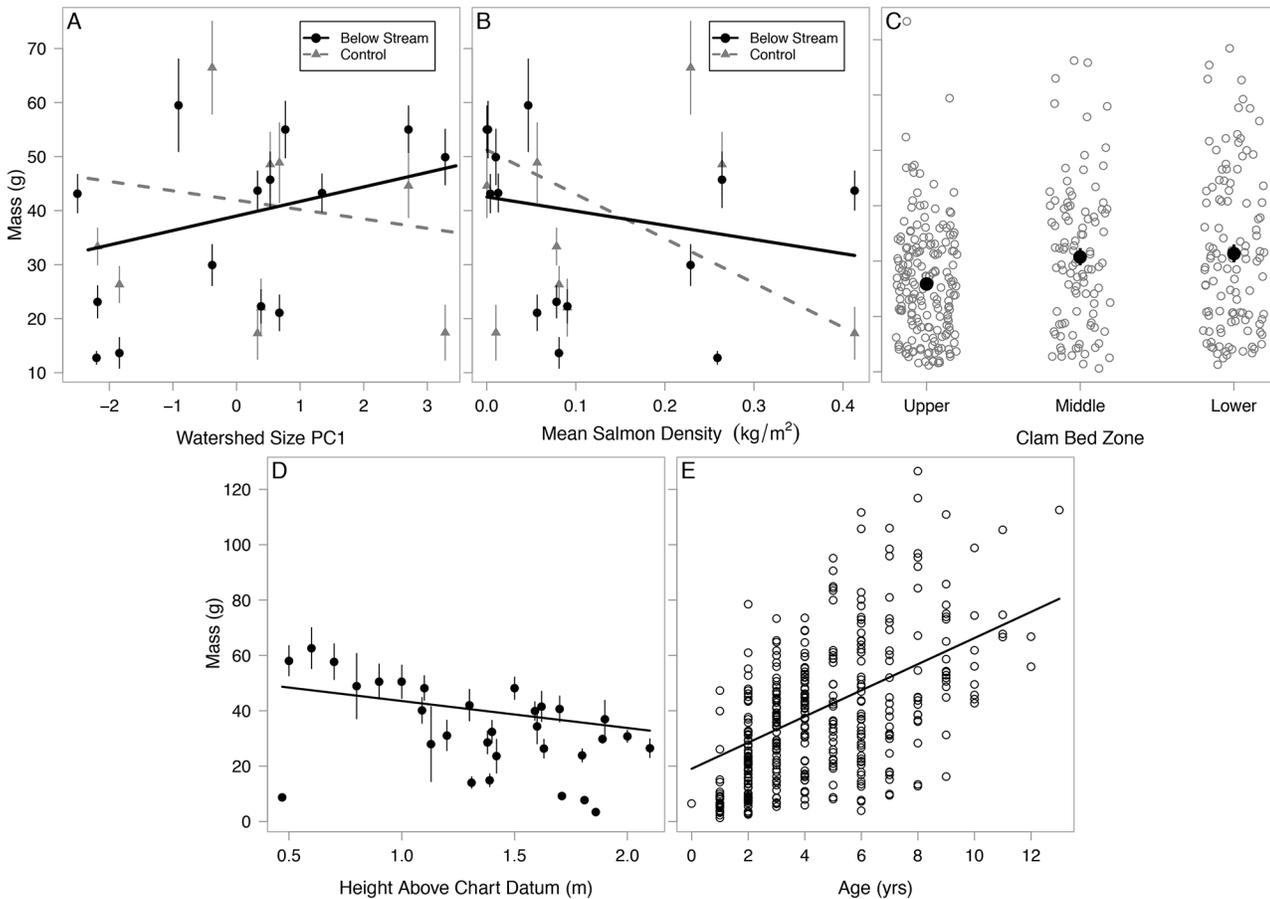


Fig 6. Correlates of soft-shell clam size. (A) Watershed size PC1 at below stream vs. control locations, (B) Pink salmon density at below stream vs. control locations, (C) Clam bed zone, (D) Height above chart datum, and (E) Clam age. Each data point in panels A, B and D represents mean values with standard error bars. Data points in panels C and E represent individual clams. Solid circles in panel C indicate mean mass for each zone with standard error bars. A jitter function was used in panel C for better visualization. All trend lines represent relationships using intercept and coefficients from multi-model output; thus they represent the relationships for the x-axis variable that accounts for the effects of other variables, rather than fitting the univariate data shown in each graph.

doi:10.1371/journal.pone.0125167.g006

density, individual traits, and habitat characteristics described, to varying degrees, stable isotope ratios of soft-shell clam muscle tissue. The effects of watershed size and salmon subsidies on isotope ratios generally decreased from upper to lower zones of clam beds. Clams size and percent nitrogen increased as the size of upstream watersheds increased. However the effect on clam size was only observed in tidal flats below streams and not in control locations. We were surprised to find that upstream salmon density had a negative relationship with clam size, though this negative effect was weaker below streams compared to control locations. To our knowledge, this is the first study to simultaneously test for terrestrial- and salmon-derived subsidies between and within sites across a broad spatial scale.

Watershed size explained isotope ratios and correlated positively with both size and %N of softshell clams. These results support other studies that have demonstrated the importance of terrestrial-derived resource subsidies to estuaries [23,24,50], which can scale directly with watershed size [19,26]. In this case the quantity of terrestrial resource influx into estuaries, as explained by watershed size, appears to be more important than higher-quality pulsed inputs such as salmon-nutrients. Although watershed nutrient exports may be lower quality than

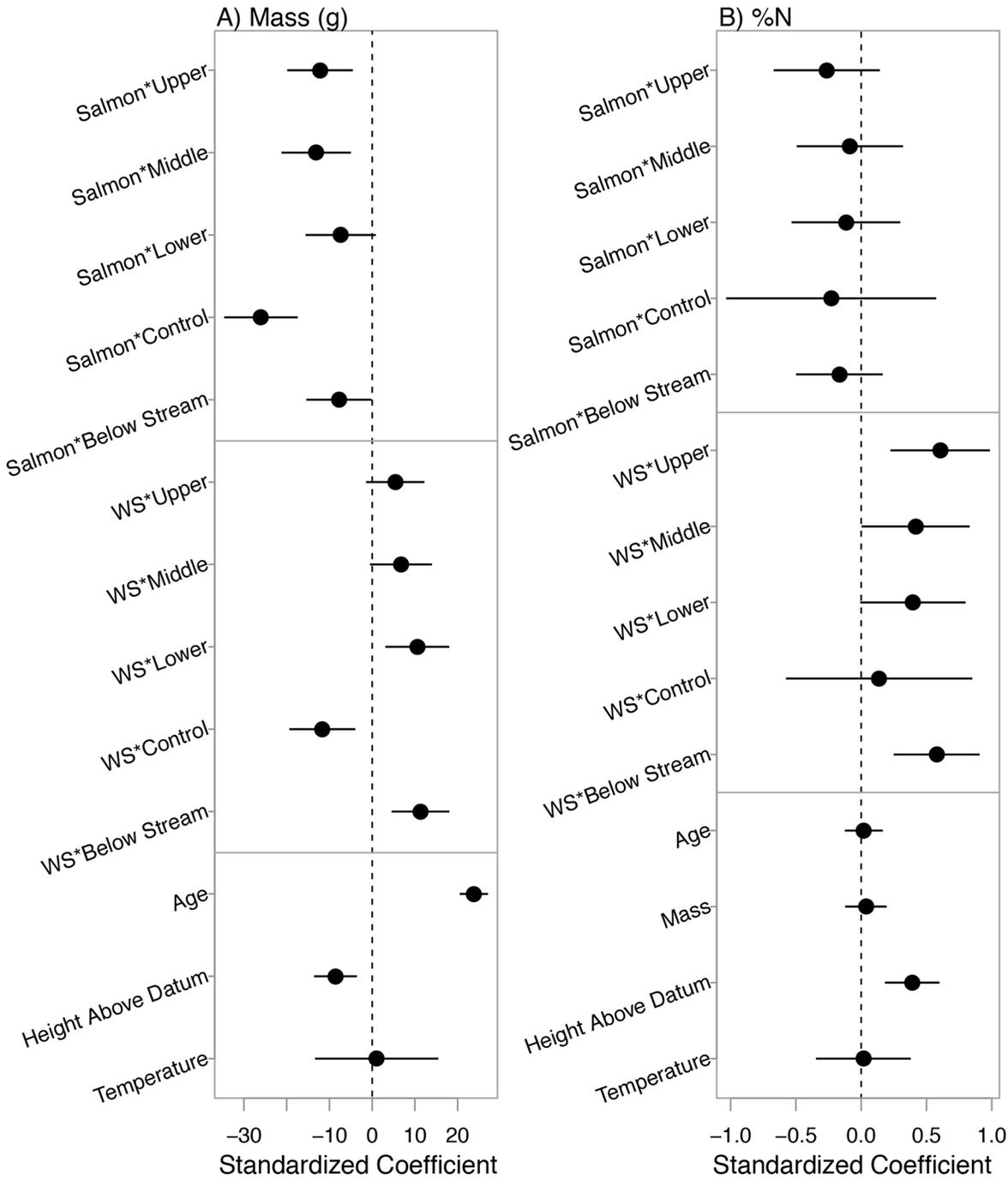


Fig 7. Standardized coefficients (mean = 0, standard deviation = 2) with 95% confidence intervals for all covariates considered in the (A) Soft-shell clam size candidate model set and (B) Soft-shell clam %N candidate model set. Salmon = 2006–07 mean pink salmon density; WS = watershed size PC1. Coefficient values indicate the change, on average, in clam size or %N as the associated covariates increase by 2 standard deviations.

doi:10.1371/journal.pone.0125167.g007

salmon or estuarine resources, total energy export from upstream may overwhelm other sources and make it a more influential resource [8,68].

The considerable depletion effect of watershed size on stable isotope ratios in soft-shell clams suggests that terrestrial-derived resources are consumed in proportion to their

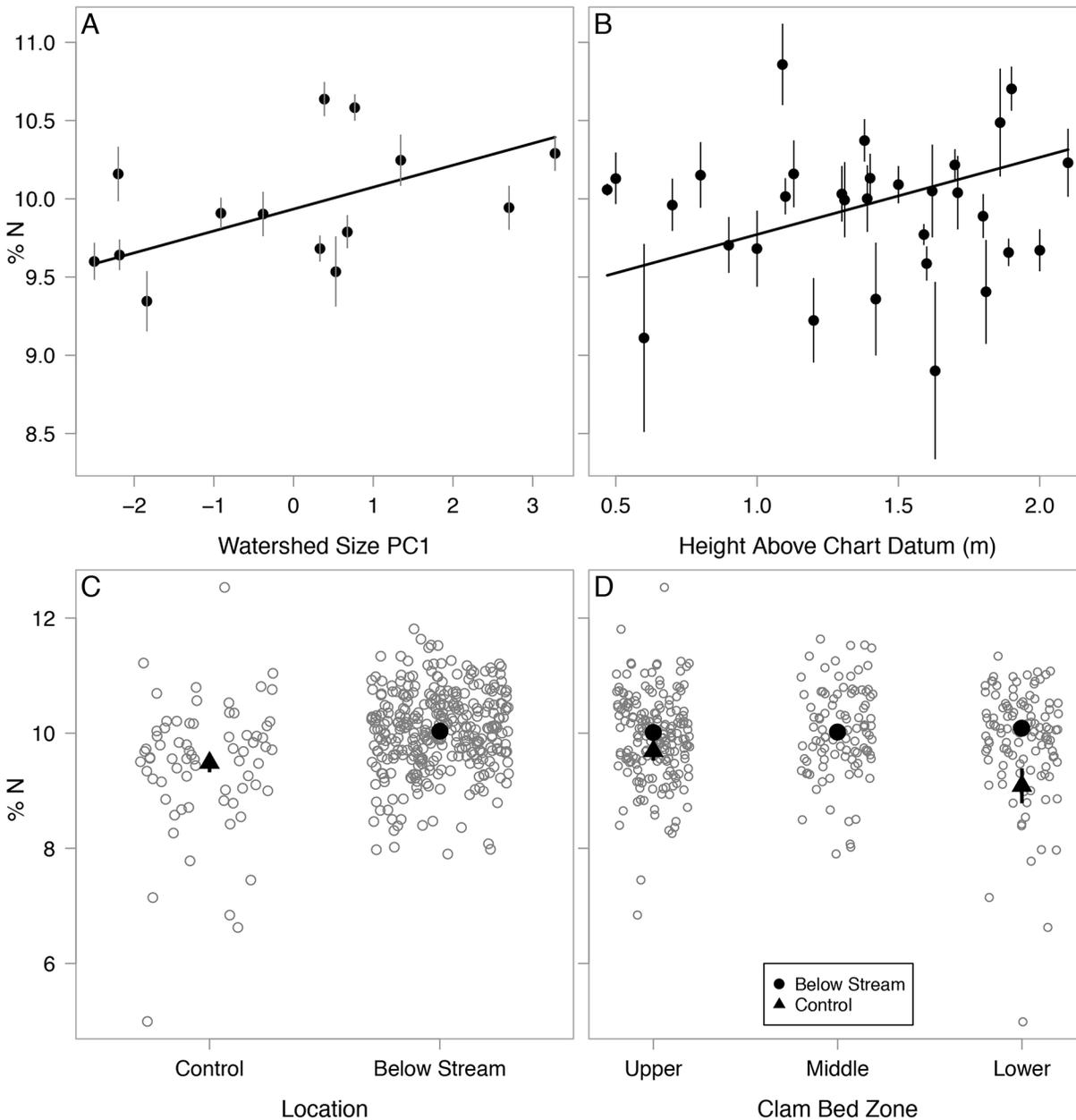


Fig 8. Correlates of soft-shell muscle tissue %N. (A) Watershed size PC1, (B) Height above chart datum, (C) Below stream vs. control locations, and (D) Clam bed zones. Data points in panels A and B represent mean values with standard error bars. Hollow data points in panels C and D represent individual clams; points were dispersed using a jitter function for better visualization. Solid symbols in panels C and D indicate mean values with standard error bars (control locations were not sampled at middle zones). All trend lines represent relationships using intercept and coefficients from multi-model output; thus they represent the relationships for the x-axis variable that accounts for the effects of other variables, rather than fitting the univariate data shown in each graph.

doi:10.1371/journal.pone.0125167.g008

availability, as also found in a study of Dungeness crabs, *Metacarcinus magister*, [26]. Stream exports are dominated by terrestrial-derived organic material [69], which have low proportions of nitrogen and carbon heavy isotopes. These inputs into estuaries also increase in proportion to watershed size [19,70]. Thus as influx of terrestrial-derived resources increases, soft-shell clams become more 'terrestrial' in their isotopic ratios. Although the effects of watershed size

on $\delta^{13}\text{C}$ were clear, the effects on $\delta^{15}\text{N}$ were less so, with a strong effect only in lower clam beds. This could be a result of settlement dynamics of particulate organic matter, which could favour deeper individuals [24]. However, as watershed sizes increase, there can also be a shift from heterotrophic nitrogen inputs to autotrophic production in streams, while maintaining a reliance on terrestrial-derived carbon [71]. Therefore, terrestrial-derived nitrogen subsidies could be displaced by freshwater algal nitrogen exports as watershed size increases [51]. Because $\delta^{15}\text{N}$ in stream algae enriches with watershed size [26], and thus becomes more similar to enriched estuarine sources [21], any relationships between watershed size and $\delta^{15}\text{N}$ could be masked.

Clams in estuaries below large watersheds were also larger, with higher percentages of nitrogen in their tissues. Bivalves have the ability to consume terrestrial-derived particulate organic matter directly [49,50]. They may also benefit indirectly, through subsidized abundances of diatoms, bacteria and microphytobenthos [72]. Because growth and %N of soft-shell clams are known to increase with nutrient loading and water flow [73,74], elevated resource imports into estuaries from larger watersheds [19] could allow individuals to grow larger, faster and with higher nitrogen content in tissues [73]. Soft-shell clams are well-suited to these types of resource subsidies, relative to other species, as they can maintain growth at higher nutrient and particulate matter concentrations [75,76]. Although watershed size appears to increase soft-shell %N throughout estuaries, the positive effect on size appears to be limited to the depositional zones below streams. A possible explanation could be sub-optimal habitat limitations to clam growth in control locations as we observed more coarse substrates such as cobble and gravel in these areas. Observed trends in clam size could also be influenced by differences in sediment grain sizes between sampling locations and estuaries, which did not measure in this study. However, clam bed locations below river outlets consisted of sand, mud and fine crushed shell mixtures which do not inhibit growth to the degree of larger substrate sizes [77]. Because growth in bivalves is known to be density dependent, lower clam densities below larger watersheds could also explain the positive correlation between watershed size and clam size [78]. Unfortunately we were unable to properly assess clam densities due to time constraints with low tides, which is a limitation to this study. However qualitative observations did not reveal any noticeable correlations between clam availability and watershed size. It is also possible that more established and mature populations and higher stream flows below large watersheds could hinder larval recruitment success and bias size distributions towards larger individuals [79,80].

Pink salmon density was a strong correlate with soft-shell clam isotope ratios compared to chum salmon. Pink salmon tend to spawn closer to estuaries than chum salmon in our study region, which increases habitat overlap with bivalves. This effect also decreased moving from upper to lower clam bed zones.

Much to our surprise, salmon density had a negative correlation with clam size. One possible explanation is that bivalves require smaller particle sizes such as sand while salmon require coarser gravel for spawning [34], so sites favorable for pink salmon could be less favourable to clams [de 81]. The timing of salmon resource subsidies, just before dormant winter periods, could also result in the routing of any energetic benefits from salmon nutrients to metabolic maintenance instead of tissue growth. Salmon can also play a dual role in stream ecosystems as sources of both nutrient subsidies and disturbance [26,82]. Pink salmon spawning in upper reaches of estuaries may exert similar disturbances to bivalves as they disrupt the substrate while digging and defending nests [83,84].

Both size and age of soft-shell clams were strongly correlated with stable isotopes. Few studies consider individual-level traits when using isotopes as an ecological tool and this study underscores the importance of their consideration [26,85,86]. In addition, local habitat conditions

can influence isotope ratios, particularly for sedentary organisms such as bivalves. Clams higher in the intertidal were enriched in $\delta^{15}\text{N}$, likely reflecting a reduction in isotope discrimination as a result of more limited feeding opportunities. Higher estuary temperatures also correlated with more depleted $\delta^{13}\text{C}$, which was contrary to our expectations. Higher water temperatures upstream could elevate exports of isotopically depleted terrestrial detritus from watersheds as a result of faster decomposition of organic matter. Alternately, this relationship could reflect the positive correlation of temperature with the percentage of alder trees upstream, which we dropped from our analyses (see [Methods](#)). Alder trees can provide substantial inputs of isotopically depleted detritus [87,88], which could also deplete soft-shell clam isotope ratios.

As expected, larger clams were found deeper in the intertidal, suggesting higher survival or growth. These clams also had reduced %N in their tissues compared to shallower individuals, contrary to our prediction. Terrestrial-derived nitrogen subsidies could be more concentrated higher in the intertidal and diluted lower down where clams are tidally submerged for longer periods of time. However, because %N of clam tissues is known to increase with growth rates [89], this result may reflect the fact that larger, and thus slower growing clams are concentrated deeper in the intertidal while smaller and faster growing clams dominate shallower locations.

Our work demonstrates the importance of connectivity amongst coastal landscapes and that this connectivity can vary with landscape traits. Our results, and other work, also suggest that the effect of watershed size can broaden food web connectivity, through increased inputs of upstream resources [26,50]. Harding and Reynolds [26] observed increases in Dungeness crab size in response to terrestrial resource influx within the same region, implying these subsidies may have broader effects within estuarine food webs. Animal movement, such as spawning salmon migrations, also provides substantial resource inputs into these ecosystems. Due to the open nature of estuaries, resource subsidies have the potential to stabilize these communities, increase productivity, and increase resilience to disturbance and periods of resource scarcity [90,91]. Natural flow regimes are an essential component to the maintenance of subsidy dynamics [92], providing resource linkages and passage for animal movement between terrestrial, freshwater and marine landscapes [93]. These considerations have direct implications for estuarine productivity in intact ecosystems such as the central coast of British Columbia, which faces increasing industrial development pressures that can disrupt discharge regimes and alter resource dynamics [94,95]. Estuaries buffer coastlines and produce resources crucial to coastal First Nations and commercial and recreational fisheries. Recognizing the importance of cross-ecosystem resource linkages in maintaining ecosystems can better enable us to understand how they might respond to human-driven pressures such as resource extraction and climate change [50]. Broader-scale studies such as this can also shed light on how cross-ecosystem processes vary across space and thus can promote realistic resource management and conservation frameworks that acknowledge the inherent heterogeneity in natural systems.

Supporting Information

S1 Table. Model weights of salmon linear models predicting $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ in soft-shell clam foot muscle tissue.

(DOCX)

S2 Table. Average coefficient estimates from multi-model analysis of candidate model set for soft-shell clam foot muscle tissue $\delta^{15}\text{N}$.

(DOCX)

S3 Table. Candidate model set from multi-model inference of soft-shell clam foot muscle tissue $\delta^{15}\text{N}$.

(DOCX)

S4 Table. Average coefficient estimates from multi-model analysis of candidate model set for soft-shell clam foot muscle tissue $\delta^{13}\text{C}$.

(DOCX)

S5 Table. Candidate model set from multi-model inference of soft-shell clam foot muscle tissue $\delta^{13}\text{C}$.

(DOCX)

S6 Table. Average coefficient estimates from multi-model analysis of candidate model set for soft-shell clam mass.

(DOCX)

S7 Table. Candidate model set from multi-model inference of soft-shell clam mass.

(DOCX)

S8 Table. Average coefficient estimates from multi-model analysis of candidate model set for soft-shell clam %N.

(DOCX)

S9 Table. Candidate model set from multi-model inference of soft-shell clam %N.

(DOCX)

S1 Fig. Bivariate plot of individual soft-shell clam mass versus shell length.

(TIF)

Acknowledgments

This research was supported by the Natural Sciences and Engineering Research Council and the Tom Buell BC Leadership Chair endowment funded by the Pacific Salmon Foundation and the BC Leading Edge Endowment Fund. Support was also received from the Tula Foundation, including a scholarship to Joel Harding through The Hakai Institute. We are grateful to everyone who provided countless hours of field and lab support including Khadijah Ali, Mike Chung, Rachel Field, Johanna Gordon-Walker, Jennifer Harding, Morgan Hocking, Leah Honka, Cherie Ko, Jason Lawrence, Michelle Nelson, Ralph Nelson, Heather Recker, Morgan Stubbs, Dan Wagner, and Alan Wu. We thank the Heiltsuk First Nation and the Department of Fisheries and Oceans (DFO) for providing salmon count data, valuable input, and logistical support. We are much indebted to Barbara Campbell and Shayne MacLellan at DFO's Pacific Biological Station for valuable support with shell ageing. Finally, we thank Sean Anderson, Michael Beakes, Doug Braun, Nick Dulvy, Jennifer Harding, Rick Routledge, the Earth to Ocean Research Group, and two anonymous reviewers for help with statistical analyses and insightful comments.

Author Contributions

Conceived and designed the experiments: JMSH MRS JDR. Performed the experiments: JMSH MRS. Analyzed the data: JMSH. Contributed reagents/materials/analysis tools: JMSH JDR. Wrote the paper: JMSH MRS JDR.

References

1. Polis GA, Anderson WB, Holt RD. Toward an Integration of Landscape and Food Web Ecology: The Dynamics of Spatially Subsidized Food Webs. *Annu Rev Ecol Syst.* 1997; 28: 289–316.
2. Polis GA, Power ME, Huxel GR. *Food Webs at the Landscape Level.* University of Chicago Press; 2004. pp. 1–548.
3. Lovelock CE, Feller IC, Adame MF, Reef R, Penrose HM, Wei L, et al. Intense storms and the delivery of materials that relieve nutrient limitations in mangroves of an arid zone estuary. *Funct Plant Biol.* 2011; 38: 514–522.
4. Spiller DA, Piovia-Scott J, Wright AN, Yang LH, Takimoto G, Schoener TW, et al. Marine subsidies have multiple effects on coastal food webs. *Ecology.* 2010; 91: 1424–1434. PMID: [20503874](#)
5. Sánchez-Piñero F, Polis GA. Bottom-up dynamics of allochthonous input: Direct and indirect effects of seabirds on islands. *Ecology.* 2000; 81: 3117–3132.
6. Stapp P, Polis GA. Marine resources subsidize insular rodent populations in the Gulf of California, Mexico. *Oecologia.* 2003; 134: 496–504. PMID: [12647121](#)
7. Nowlin WH, Vanni MJ, Yang LH. Comparing resource pulses in aquatic and terrestrial ecosystems. *Ecology.* 2008; 89: 647–659. PMID: [18459329](#)
8. Marczak LB, Thompson RM, Richardson JS. Meta-analysis: Trophic level, habitat, and productivity shape the food web effects of resource subsidies. *Ecology.* 2007; 88: 140–148. PMID: [17489462](#)
9. Anderson WB, Wait DA, Stapp P. Resources from another place and time: Responses to pulses in a spatially subsidized system. *Ecology.* 2008; 89: 660–670. PMID: [18459330](#)
10. Yang LH, Edwards KF, Byrnes JE, Bastow JL, Wright AN, Spence KO. A meta-analysis of resource pulse–consumer interactions. *Ecol Monogr.* 2010; 80: 125–151.
11. Marcarelli AM, Baxter CV, Mineau MM, Hall RO Jr. Quantity and quality: unifying food web and ecosystem perspectives on the role of resource subsidies in freshwaters. *Ecology.* 2011; 92: 1215–1225. PMID: [21797150](#)
12. Correll DL, Jordan TE, Weller DE. Nutrient Flux in a Landscape—Effects of Coastal Land-Use and Terrestrial Community Mosaic on Nutrient Transport to Coastal Waters. *Estuaries.* 1992; 15: 431–442.
13. Leroux SJ, Loreau M. Subsidy hypothesis and strength of trophic cascades across ecosystems. *Ecol Lett.* 2008; 11: 1147–1156. doi: [10.1111/j.1461-0248.2008.01235.x](#) PMID: [18713270](#)
14. Richardson JS, Zhang Y, Marczak LB. Resource subsidies across the land-freshwater interface and responses in recipient communities. Milner AM, Tockner K, editors. *River Res Applic.* 2009; 26: 55–66.
15. Carr MH, Neigel JE, Estes JA, Andelman S. Comparing marine and terrestrial ecosystems: implications for the design of coastal marine reserves. *Ecol Appl.* 2003; 13: S90–S107.
16. Elliott M, Whitfield AK. Challenging paradigms in estuarine ecology and management. *Estuar Coast Shelf Sci.* 2011; 94: 306–314.
17. Milliman JD, Syvitski JPM. *Geomorphic/Tectonic Control of Sediment Discharge to the Ocean: The Importance of Small Mountainous Rivers.* J Geol. The University of Chicago Press; 1992; 100: 525–544.
18. Vinagre C, Salgado J, Cabral HN, Costa MJ. Food Web Structure and Habitat Connectivity in Fish Estuarine Nurseries—Impact of River Flow. *Estuaries Coast.* 2010; 34: 663–674.
19. Sakamaki T, Shum JYT, Richardson JS. Watershed Effects on Chemical Properties of Sediment and Primary Consumption in Estuarine Tidal Flats: Importance of Watershed Size and Food Selectivity by Macrobenthos. *Ecosystems.* Springer-Verlag; 2010; 13: 328–337.
20. Chester AJ, Larrance JD. Composition and vertical flux of organic matter in a large Alaskan estuary. *Estuaries Coast.* 1981; 4: 42–52.
21. Deegan LA, Garritt RH. Evidence for spatial variability in estuarine food webs. *Mar Ecol-Prog Ser.* 1997; 147: 31–47.
22. Chanton JP, Lewis FG. Examination of Coupling between Primary and Secondary Production in a River-Dominated Estuary: Apalachicola Bay, Florida, USA. *Limnol Oceanogr.* JSTOR; 2002; 47: 683–697.
23. Connolly RM, Schlacher TA, Gaston TF. Stable isotope evidence for trophic subsidy of coastal benthic fisheries by river discharge plumes off small estuaries. *Mar Biol Res.* Taylor & Francis Group; 2009; 5: 164–171.

24. Darnaude AM, Salen-Picard C, Harmelin-Vivien ML. Depth variation in terrestrial particulate organic matter exploitation by marine coastal benthic communities off the Rhone River delta (NW Mediterranean). *Mar Ecol-Prog Ser. Inter-Research*; 2004; 275: 47–57.
25. B naru D, Harmelin-Vivien ML, Gomoiu M-T, Onciu T-M. Influence of the Danube River inputs on C and N stable isotope ratios of the Romanian coastal waters and sediment (Black Sea). *Mar Pollut Bull.* 2007; 54: 1385–1394. PMID: [17631911](#)
26. Harding JMS, Reynolds JD. From earth and ocean: investigating the importance of cross-ecosystem resource linkages to a mobile estuarine consumer. *Ecosphere.* 2014; 5: art54.
27. Peterson BJ, Fry B. Stable isotopes in ecosystem studies. *Annu Rev Ecol Syst.* JSTOR; 1987; 18: 293–320.
28. Fry B, Scalan RS, Parker PL. Stable carbon isotope evidence for two sources of organic matter in coastal sediments: seagrasses and plankton. *Geochim Cosmochim Acta.* 1977; 41: 1875–1877.
29. Alliot E, Younesa WAN, Romano JC, Rebouillon P, Massé H. Biogeochemical impact of a dilution plume (Rhone River) on coastal sediments. *Estuar Coast Shelf Sci.* 2003; 57: 357–367.
30. Hopkinson CS, Giblin AE, Tucker J, Garritt RH. Benthic metabolism and nutrient cycling along an estuarine salinity gradient. *Estuaries Coast.* 1999; 22: 863–881.
31. Oczkowski AJ, Lewis FG, Nixon SW, Edmiston HL, Robinson RS, Chanton JP. Fresh Water Inflow and Oyster Productivity in Apalachicola Bay, FL (USA). *Estuaries Coast.* 2011; 34: 993–1005.
32. Hoffman JC, Bronk DA, Olney JE. Contribution of allochthonous carbon to American shad production in the Mattaponi River, Virginia, using stable isotopes. *Estuaries Coast.* Springer; 2007; 30: 1034–1048.
33. B naru D, Harmelin-Vivien ML. Trophic links and riverine effects on food webs of pelagic fish of the north-western Black Sea. *Mar Freshw Res.* CSIRO; 2009; 60: 529–540.
34. Groot C, Margolis L. *Pacific Salmon Life Histories.* Vancouver, British Columbia, Canada: UBC Press; 1991. pp. 1–576.
35. Hocking MD, Reynolds JD. Impacts of salmon on riparian plant diversity. *Science.* 2011; 331: 1609–1612. PMID: [21442794](#)
36. Janetski DJ, Chaloner DT, Tiegs SD, Lamberti GA. Pacific salmon effects on stream ecosystems: a quantitative synthesis. *Oecologia.* 2009; 159: 583–595. doi: [10.1007/s00442-008-1249-x](#) PMID: [19132408](#)
37. Gende SM, Edwards RT, Willson MF, Wipfli MS. Pacific salmon in aquatic and terrestrial ecosystems. *BioScience.* 2002; 52: 917–928.
38. Naiman RJ, Bilby RE, Schindler DE, Helfield JM. Pacific salmon, nutrients, and the dynamics of freshwater and riparian ecosystems. *Ecosystems.* 2002; 5: 399–417.
39. Moore JW, Schindler DE, Carter JL, Fox J, Griffiths J, Holtgrieve GW. Biotic control of stream fluxes: spawning salmon drive nutrient and matter export. *Ecology.* 2007; 88: 1278–1291. PMID: [17536413](#)
40. Kohler AE, Kusnierz PC, Copeland T, Venditti DA, Denny L, Gable J, et al. Salmon-mediated nutrient flux in selected streams of the Columbia River basin, USA. *Can J Fish Aquat Sci.* 2013; 70: 502–512.
41. Tiegs SD, Levi PS, Rüegg J, Chaloner DT, Tank JL, Lamberti GA. Ecological Effects of Live Salmon Exceed Those of Carcasses During an Annual Spawning Migration. *Ecosystems.* 2011; 14: 598–614.
42. Gende SM, Quinn TP, Willson MF, Heintz R, Scott TM. Magnitude and Fate of Salmon-Derived Nutrients and Energy in a Coastal Stream Ecosystem. *J of Freshwater Ecol.* 2004; 19: 149–160.
43. Mitchell NL, Lamberti GA. Responses in dissolved nutrients and epilithon abundance to spawning salmon in southeast Alaska streams. *Limnol Oceanogr.* 2005; 50: 217–227.
44. Cak AD, Chaloner DT, Lamberti GA. Effects of spawning salmon on dissolved nutrients and epilithon in coupled stream-estuary systems of southeastern Alaska. *Aquat Sci.* 2008; 70: 169–178.
45. Jauquet J, Pittman N, Heinis JA, Thompson S, Tatyama N, Cederholm CJ. Observations of chum salmon consumption by wildlife and changes in water chemistry at Kennedy Creek during 1997–2000. In: Stockner J, editor. *Nutrients in Salmonid Ecosystems: Sustaining Production and Biodiversity,* American Fisheries Society Symposium, Eugene, Oregon, April 24–26, 2001. American Fisheries Society, Bethesda, Maryland, USA; 2003. pp. 71–88.
46. Reimchen TE. Further studies of predator and scavenger use of chum salmon in stream and estuarine habitats at Bag Harbour, Gwaii Haanas [Internet]. Queen Charlotte City, British Columbia, Canada: Canadian Parks Service; 1994. Available: <http://web.uvic.ca/~reimlab/FURTHER%20STUDIES%20BAG%20HARBOUR0001.pdf>. Accessed: 2013 Jul 27.
47. Fujiwara M, Highsmith RC. Harpacticoid copepods: potential link between inbound adult salmon and outbound juvenile salmon. *Mar Ecol-Prog Ser.* 1997; 158: 205–216.

48. Fry B. Conservative mixing of stable isotopes across estuarine salinity gradients: A conceptual framework for monitoring watershed influences on downstream fisheries production. *Estuaries*. 2002; 25: 264–271.
49. Kasai A, Nakata A. Utilization of terrestrial organic matter by the bivalve *Corbicula japonica* estimated from stable isotope analysis. *Fisheries Sci.* Springer-Verlag; 2005; 71: 151–158.
50. Sakamaki T, Richardson JS. Effects of small rivers on chemical properties of sediment and diets for primary consumers in estuarine tidal flats. *Mar Ecol-Prog Ser.* 2008; 360: 13–24.
51. Vannote RL, Minshall GW, Cummins KW, Sedell JR, Cushing CE. The River Continuum Concept. *Can J Fish Aquat Sci.* 1980; 37: 130–137.
52. Pojar J, Klinka K, Demarchi DA. Coastal western hemlock zone. In: Meidinger DV, Pojar J, editors. *Ecosystems of British Columbia*. Victoria, British Columbia, Canada: Research Branch, Ministry of Forests; 1991. pp. 95–111.
53. Price K, Roburn A, MacKinnon A. Ecosystem-based management in the Great Bear Rainforest. *For Ecol Manage.* 2009; 258: 495–503.
54. Slaney TL, Hyatt KD, Northcote TG, Fielden RJ. Status of anadromous salmon and trout in British Columbia and Yukon. *Fisheries*. Taylor & Francis; 1996; 21: 20–35.
55. MacDonald BA, Thomas MLH. Age determination of the soft-shell clam *Mya arenaria* using shell internal growth lines. *Mar Biol.* 1980; 58: 105–109.
56. Bain MB, Stevenson NJ. *Aquatic Habitat Assessment*. Bethesda, Maryland: American Fisheries Society; 1999. pp. 1–216.
57. Government of British Columbia. *iMapBC* [Internet]. 1st ed. 2006. Available: http://www.data.gov.bc.ca/dbc/geographic/view_and_analyze/imapbc/index.page Accessed: 2014 Jan 10.
58. Peters RH. *The Ecological Implications of Body Size*. Cambridge, United Kingdom: Cambridge University Press; 1983. pp. 1–329.
59. R Core Team. *R: A language and environment for statistical computing*. [Internet]. 2nd ed. Vienna, Austria: R Foundation for Statistical Computing; 2012. Available: <http://www.R-project.org/>
60. Zuur AF, Ieno EN, Walker NJ, Saveliev AA, Smith GM. *Mixed Effects Models and Extensions in Ecology with R*. New York, New York, USA: Springer; 2009. pp. 1–574.
61. Zuur AF, Ieno EN, Elphick CS. A protocol for data exploration to avoid common statistical problems. *Methods in Ecology and Evolution*. 2010; 1: 3–14.
62. Pinheiro J, Bates D, DebRoy S, Sarkar D, R Core Team. nlme: Linear and Nonlinear Mixed Effects Models. R package version 3.1–111. [Internet]. 3rd ed. 2012. Available: <http://cran.r-project.org/web/packages/nlme/index.html>
63. Burnham KP, Anderson DR. *Model selection and multimodel inference: a practical information-theoretic approach*. 2nd ed. New York, New York, USA: Springer; 2002. pp. 1–488.
64. Schielzeth H. Simple means to improve the interpretability of regression coefficients. *Methods in Ecology and Evolution*. 2010; 1: 103–113.
65. Grueber CE, Nakagawa S, Laws RJ, Jamieson IG. Multimodel inference in ecology and evolution: challenges and solutions. *Journal of Evolutionary Biology*. 2011; 24: 699–711. doi: [10.1111/j.1420-9101.2010.02210.x](https://doi.org/10.1111/j.1420-9101.2010.02210.x) PMID: [21272107](https://pubmed.ncbi.nlm.nih.gov/21272107/)
66. Gelman A. Scaling regression inputs by dividing by two standard deviations. *Statist Med.* 2008; 27: 2865–2873.
67. Barton K. MuMIn: Multi-model inference. R package version 1.7.11. [Internet]. 1st ed. 2012. p. 37. Available: <http://CRAN.R-project.org/package=MuMIn>
68. Jonsson B, Jonsson N. Migratory Atlantic salmon as vectors for the transfer of energy and nutrients between freshwater and marine environments. *Freshw Biol.* 2002; 48: 21–27.
69. Harmelin-Vivien ML, Dierking J, Barnaud D, Fontaine MF, Arlhac D. Seasonal variation in stable C and N isotope ratios of the Rhone River inputs to the Mediterranean Sea (2004–2005). *Biogeochemistry*. Springer; 2010; 100: 139–150.
70. Darnaude AM. Fish ecology and terrestrial carbon use in coastal areas: implications for marine fish production. *J Anim Ecol.* Wiley Online Library; 2005; 74: 864–876.
71. Sakamaki T, Richardson JS. Nonlinear variation of stream-forest linkage along a stream-size gradient: an assessment using biogeochemical proxies of in-stream fine particulate organic matter. *Arnot S, editor. J Appl Ecol.* 2013; 50: 1019–1027.
72. Yamanaka T, Mizota C, Maki Y, Matsumasa M. *Estuarine, Coastal and Shelf Science*. Estuar Coast Shelf Sci. Elsevier Ltd; 2013; 126: 87–92.

73. Carmichael RH, Shriver AC, Valiela I. Bivalve Response to Estuarine Eutrophication: The Balance between Enhanced Food Supply and Habitat Alterations. *J Shellfish Res.* 2012; 31: 1–11.
74. Emerson CW. Influence of Sediment Disturbance and Water Flow on the Growth of the Soft-Shell Clam, *Mya arenaria* L. *Can J Fish Aquat Sci.* 1990; 47: 1655–1663.
75. MacDonald BA, Bacon GS, Ward JE. Physiological responses of infaunal (*Mya arenaria*) and epifaunal (*Placopecten magellanicus*) bivalves to variations in the concentration and quality of suspended particles II. Absorption efficiency and scope for growth. *J Exp Mar Bio Ecol.* 1998; 219: 127–141.
76. Weiss ET, Carmichael RH, Valiela I. The effect of nitrogen loading on the growth rates of quahogs (*Mercenaria mercenaria*) and soft-shell clams (*Mya arenaria*) through changes in food supply. *Aquaculture.* Elsevier; 2002; 211: 275–289.
77. Newell CR, Hidu H. The effects of sediment type on growth rate and shell allometry in the soft shelled clam *Mya arenaria* L. *J Exp Mar Bio Ecol.* 1982; 65: 285–295.
78. Beal BF, Gayle Kraus M. Interactive effects of initial size, stocking density, and type of predator deterrent netting on survival and growth of cultured juveniles of the soft-shell clam, *Mya arenaria* L., in eastern Maine. *Aquaculture.* 2002; 208: 81–111.
79. André C, Rosenberg R. Adult-larval interactions in the suspension-feeding bivalves *Cerastoderma edule* and *Mya arenaria*. *Mar Ecol-Prog Ser.* 1991; 71: 227–234.
80. Morse BL, Hunt HL. Impact of settlement and early post-settlement events on the spatial distribution of juvenile *Mya arenaria* on an intertidal shore. *J Exp Mar Bio Ecol.* Elsevier B.V.; 2013; 448: 57–65.
81. de la Huz R, Lastra M, López J. The influence of sediment grain size on burrowing, growth and metabolism of *Donax trunculus* L. (*Bivalvia: Donacidae*). *J Sea Res.* 2002; 47: 85–95.
82. Moore JW, Schindler DE, Scheuerell MD. Disturbance of freshwater habitats by anadromous salmon in Alaska. *Oecologia.* 2004; 139: 298–308. PMID: [14997375](#)
83. Neave F. Salmon of the North Pacific. Part III. A review of the life history of North Pacific salmon. 5: Pink salmon in British Columbia. *Int North Pacif Fish Comm Bull.* 1966; 18: 71–79.
84. Hunter JG. Survival and Production of Pink and Chum Salmon in a Coastal Stream. *J Fish Res Bd Can.* 1959; 16: 835–886.
85. Martínez del Rio C, Wolf N, Carleton SA, Gannes LZ. Isotopic ecology ten years after a call for more laboratory experiments. *Biol Rev.* 2009; 84: 91–111. doi: [10.1111/j.1469-185X.2008.00064.x](#) PMID: [19046398](#)
86. Swain NR, Hocking MD, Harding JN, Reynolds JD. Effects of salmon on the diet and condition of stream-resident sculpins. *Can J Fish Aquat Sci.* 2014; 71: 521–532.
87. Wipfli MS, Musslewhite J. Density of red alder (*Alnus rubra*) in headwaters influences invertebrate and detritus subsidies to downstream fish habitats in Alaska. *Hydrobiologia.* 2004; 520: 153–163.
88. Sakamaki T, Richardson JS. Retention, breakdown, and biological utilisation of deciduous tree leaves in an estuarine tidal flat of southwestern British Columbia, Canada. *Can J Fish Aquat Sci.* 2008; 65: 38–46.
89. Carmichael RH, Shriver AC, Valiela I. Changes in shell and soft tissue growth, tissue composition, and survival of quahogs, *Mercenaria mercenaria*, and softshell clams, *Mya arenaria*, in response to eutrophic-driven changes in food supply and habitat. *J Exp Mar Bio Ecol.* 2004; 313: 75–104.
90. Huxel GR, McCann K. Food Web Stability: The Influence of Trophic Flows across Habitats. *Am Nat.* 1998; 152: 460–469. doi: [10.1086/286182](#) PMID: [18811452](#)
91. Anderson WB, Polis GA. Allochthonous nutrient and food inputs: consequences for temporal stability. In: Power ME, Huxel GR, editors. *Food Webs at the Landscape Level.* University of Chicago Press; 2004. pp. 82–95.
92. Poff NL, Allan JD, Bain MB, Karr JR, Prestegard KL, Richter BD, et al. The Natural Flow Regime. *Bio-Science.* American Institute of Biological Sciences; 1997; 47: 769–784.
93. Palarly JE, Witman JD. Flow, recruitment limitation, and the maintenance of diversity in marine benthic communities. *Ecology.* 2014; 95: 286–297. PMID: [24669723](#)
94. Fulweiler RW, Nixon SW. Export of nitrogen, phosphorus, and suspended solids from a southern New England watershed to Little Narragansett Bay. *Biogeochemistry.* 2005; 76: 567–593.
95. Valiela I, Bartholomew M, Giblin A, Tucker J, Harris C, Martinetto P, et al. Watershed Deforestation and Down-Estuary Transformations Alter Sources, Transport, and Export of Suspended Particles in Panamanian Mangrove Estuaries. *Ecosystems.* 2014; 17: 96–111.
96. Kline TC, Goering JJ, Mathisen OA, Poe PH, Parker PL, Scalan RS. Recycling of Elements Transported Upstream by Runs of Pacific Salmon: II. $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ Evidence in the Kvichak River Watershed, Bristol Bay, Southwestern Alaska. *Can J Fish Aquat Sci.* 1993; 50: 2350–2365.

97. Galster JC. Natural and anthropogenic influences on the scaling of discharge with drainage area for multiple watersheds. *Geosphere*. 2007; 3: 260.
98. Rossi F, Herman PMJ, Middelburg JJ. Interspecific and intraspecific variation of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in deposit- and suspension-feeding bivalves (*Macoma balthica* and *Cerastoderma edule*): Evidence of ontogenetic changes in feeding mode of *Macoma balthica*. *Limnol Oceanogr*. 2004; 49: 408–414.
99. Carleton SA, Martínez del Río C. Growth and catabolism in isotopic incorporation: a new formulation and experimental data. *Funct Ecol*. 2010; 24: 805–812.
100. Bowes RE, Lafferty MH, Thorp JH. Less means more: nutrient stress leads to higher $\delta^{15}\text{N}$ ratios in fish. *Freshw Biol*. 2014; 59: 1926–1931.
101. Honkoop PJC, Beukema JJ. Loss of body mass in winter in three intertidal bivalve species: an experimental and observational study of the interacting effects between water temperature, feeding time and feeding behaviour. *J Exp Mar Bio Ecol*. 1997; 212: 277–297.
102. Bilby RE, Beach EW, Fransen BR, Walter JK, Bisson PA. Transfer of nutrients from spawning salmon to riparian vegetation in western Washington. *T Am Fish Soc*. 2003; 132: 733–745.
103. Brousseau DJ. Analysis of growth rate in *Mya arenaria* using the Von Bertalanffy equation. *Mar Biol*. Springer-Verlag; 1979; 51: 221–227.
104. Rip JMK, McCann KS. Cross-ecosystem differences in stability and the principle of energy flux. *Ecol Lett*. 2011; 14: 733–740. doi: [10.1111/j.1461-0248.2011.01636.x](https://doi.org/10.1111/j.1461-0248.2011.01636.x) PMID: [21627748](https://pubmed.ncbi.nlm.nih.gov/21627748/)
105. Shurin JB, Gruner DS, Hillebrand H. All wet or dried up? Real differences between aquatic and terrestrial food webs. *Proc R Soc B*. 2006; 273: 1–9. PMID: [16519227](https://pubmed.ncbi.nlm.nih.gov/16519227/)