

Serum γ -glutamyl Transferase Levels, Insulin Resistance and Liver Fibrosis in Patients with Chronic Liver Diseases

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Abstract

Background and Aims: Serum levels of γ -glutamyl-transpeptidase(γ -GT) were associated with liver disease severity and metabolic alterations, which in turn are able to affect hepatic damage. In patients with nonalcoholic fatty liver disease (NAFLD), genotype 1 chronic hepatitis C (G1CHC) and chronic hepatitis B (CHB), we assessed the link between liver fibrosis and γ -GT serum levels, and we evaluated if normal or high γ -GT serum levels affect the association between insulin resistance (IR) and severity of liver fibrosis.

Methods: 843 consecutive patients with chronic liver disease (CLD)(193 NAFLD, 481 G1CHC, 169 CHB) were evaluated by liver biopsy (Kleiner and Scheuer scores) and clinical and metabolic measurements. IR was diagnosed if HOMA>3. A serum γ-GT concentration of >36 IU/L in females and >61 IU/L in males was considered the threshold value for identifying high levels of γ-GT.

Results: By multivariate logistic regression analysis, abnormal γ-GT serum levels were independently linked to severe liver fibrosis in patients with NAFLD (OR2.711,Cl1.120–6.564,p=0.02), G1CHC (OR3.461,Cl2.138–5.603,p<0.001) and CHB (OR2.778,Cl1.042–7.414,p=0.04), together with IR and liver necroinflammation, and with a negative predictive value>80%. Interestingly, among patients with high or normal γ-GT values, even if IR prevalence was significantly higher in patients with severe fibrosis compared to those without, IR remained significantly associated with severe fibrosis in patients with abnormal γ-GT values only (OR4.150,Cl1.079–15.970,p=0.03 for NAFLD; OR2.250,Cl1.211–4.181,p=0.01 for G1CHC; OR3.096,Cl2.050–34.220,p=0.01 for CHB).

Conclusions: In patients with CLD, IR is independently linked to liver fibrosis only in patients with abnormal γ -GT values, without differences according to liver disease etiology, and suggesting a role of γ -GT as a marker of metabolic-induced liver damage. These data could be useful for the clinical and pharmacologic management of patients with CLD.

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Introduction

The prognosis of patients with chronic liver diseases (CLD), independently of the underlying etiology, is ultimately decided by the amount of liver fibrosis that will accumulate over the years as a consequence of several mechanisms of tissue injury, with the ultimate development of cirrhosis and its complicances [1,2]. Other than conventional host and viral factors known to affect liver fibrosis progression, metabolic factors, and especially insulin resistance (IR) act as major disease modifiers associated with the severity of hepatic damage. Specifically, IR is the key determinant of nonalcoholic fatty liver disease (NAFLD) and its severity [3], is common in western patients with chronic hepatitis C (CHC) [4], due to both viral and host factors [5], and associated with the severity of fibrosis and its progression independently of steatosis and visceral obesity [6,7], and finally seems also associated with the severity of liver fibrosis in patients with chronic hepatitis B (CHB) [8].

However, nevertheless these data, it is probably that the impact of IR on the severity of liver fibrosis is different according to the clinical setting. These considerations in fact arise from contrasting evidences about the association between liver fibrosis and IR observed in some studies on patients with CHC [4,9] or CHB [10], and from the evidences that insulin-sensitizer therapies obtain an improvement in histological outcomes in a proportion of NAFLD patients only [11,12].

 γ -glutamyl transpeptidase (γ -GT) is a hepatic and biliary enzyme synthesized by hepatocytes as well as epithelial cells of intra-hepatic bile ducts [13,14]. Several cross-sectional and prospective studies showed that plasma γ -GT is associated with metabolic syndrome risk factors and with markers of inflammation [15–18], also representing in large population studies an independent predictor of all-cause, liver-related, diabetes-related, and cancer-related death [13]. In this line, recent studies suggest that plasma γ -GT is a significant predictor of NAFLD, independently of alcohol intake [14,15,19], and that more specif-

ically elevated γ -GT levels are associated with a more severe histological spectrum of NAFLD, namely the presence of NASH and fibrosis [16], whereas reductions in γ -GT predict histological improvement in NAFLD following bariatric surgery [15,20]. Similarly, different cross-sectional studies also showed that higher γ -GT serum levels are predictive of the severity of liver fibrosis in patients with both CHC and CHB, being this test also included in non-invasive scores for hepatic fibrosis assessment [21,22].

All these data therefore could suggest that γ -GT is expression of an environmental and probably genetic state of inflammation and metabolic dysfunction, affecting the spectrum of the severity of liver disease.

In light of these facts, we aimed to confirm the association between severity of liver disease and γ -GT serum levels in patients with an histological diagnosis of CLD, due to NAFLD, CHC and CHB, and to evaluate, in these patients, if the presence of normal or high γ -GT serum levels affects the association between IR and severity of liver fibrosis.

Materials and Methods

Patients

Eight-hundred and forty-three consecutive patients with CLD, due to viral or metabolic etiology (193 with NAFLD, 481 with genotype 1 CHC, and 169 with CHB), recruited at the Gastrointestinal & Liver Unit at the University Hospital in Palermo and fulfilling all inclusion and exclusion criteria detailed below, were assessed. (a) histological diagnosis of NAFLD, or G1 CHC, or CHB on a liver biopsy performed less than 6 months before enrollment; (b) NAFLD diagnosed on the basis of chronically elevated ALT for at least 6 months, alcohol consumption of <20 g/day, steatosis (>5% of hepatocytes) at histology with necroinflammation and/or fibrosis, and negative anti-HCV and HBsAG; (c) G1 CHC characterized by the presence of anti-HCV and HCVRNA, with persistently abnormal alanine aminotransferase (ALT), alcohol consumption of <20 g/ day in the last 12 months or more, naive to antiviral therapy, and negative HBsAg; (d) CHB characterized by the presence of HBsAg and HBVDNA >2000 IU/ml, with persistently abnormal alanine aminotransferase (ALT), alcohol consumption of <20 g/day in the last 12 months or more, naive to antiviral therapy, and negative anti-HCV. Exclusion criteria were: (1) advanced cirrhosis; (2) hepatocellular carcinoma; (3) other causes of liver disease or mixed aetiologies; (4) human immunodeficiency virus infection; (5) previous treatment with antiviral therapy, immunosuppressive drugs and/or regular use of steatosis-inducing drugs, evaluated by interview; (6) active intravenous drug addiction.

The study was carried out in accordance with the principles of the Declaration of Helsinki and its appendices, and with local and national laws. Approval was obtained from the hospital's Internal Review Board and its Ethics Committee, and written informed consent was obtained from all patients and controls.

Clinical and Laboratory Assessment

Clinical and anthropometric data were collected at the time of liver biopsy. Body mass index (BMI) was calculated on the basis of weight in kilograms and height in meters, and patients were classified as normal weight (BMI, 18.5–24.9 kg/m²), overweight (BMI, 25–29.9), or obese (BMI ≥30). A diagnosis of arterial hypertension was based on the following criteria: systolic blood pressure ≥130 mm Hg and/or diastolic blood pressure ≥85 mm Hg (measured three times in 30 minutes, in the sitting position and using a brachial sphygmomanometer), or use of blood-pressure-lowering agents. A diagnosis of type 2 diabetes was based on the

revised criteria of the American Diabetes Association, using a value of fasting blood glucose ≥126 mg/dL on at least two occasions [23]. In patients with a previous diagnosis of type 2 diabetes, current therapy with insulin or oral hypoglycemic agents was documented.

A 12-hour overnight fasting blood sample was drawn at the time of biopsy to determine serum levels of ALT, total cholesterol, triglycerides, plasma glucose concentration, insulin, and platelet count. Insulin resistance (IR) was determined by the homeostasis model assessment (HOMA) method, using the following equation [24]: insulin resistance (HOMA-IR) = fasting insulin (μ U/mL) x fasting glucose (mmol/L)/22.5. HOMA-IR has been validated in comparison with the euglycemic/hyperinsulinemic clamp technique in both diabetic and non-diabetic patients [25]. HOMA-IR values >3 were considered to indicate IR [26].

Commercially available enzymatic colorimetric test was used for serum γ -GT assays. In accordance with the kit's instructions, a serum γ -GT concentration of >36 IU/L in females, and >61 IU/L in males was considered the threshold value for identifying high levels of γ -GT.

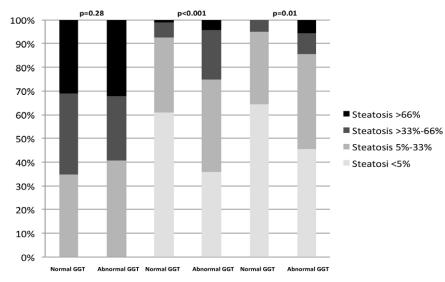
HCV-infected individuals were tested at the time of biopsy for HCV-RNA using qualitative PCR (Cobas Amplicor HCV Test version 2.0; limit of detection: 50 IU/ml). HCVRNA-positive samples were quantified by Versant HCV-RNA 3.0 bDNA (Bayer Co. Tarrytown, NY, USA) expressed in IU/ml. Genotyping was performed by INNOLiPA (HCV II, Bayer HealthCare, Berkeley, CA, USA). Hepatitis B virus-infected patients were tested at the time of biopsy for HBs, HBeAg, anti-HBe and anti-HDV IgG, using commercial enzyme immunoassays (Dia Sorin, Saluggia, Italy). HBV-DNA was quantified by bDNA (Versant HBV 3.0, Siemens Medical Solutions Diagnostics Europe, Dublin, Ireland; range 357–17 857 000 IU/ml).

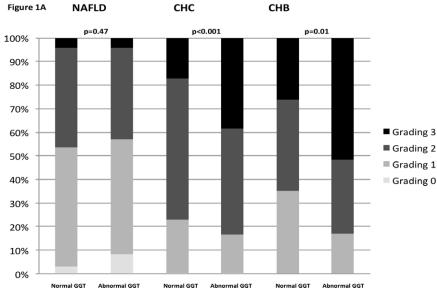
Histology

Slides were coded and read by one pathologist (D.C.), who was unaware of the patient's identity and history. Steatosis was assessed as the percentage of hepatocytes containing fat droplets (minimum 5%), and evaluated as a continuous variable. Steatosis was classified according to Kleiner classification as: absent <5%; mild 5%–33%; moderate >33%–66%; and severe >66%. In the NAFLD group, the Kleiner classification [27] was used to compute the NAFLD activity score (from 0 to 8, on a scale including separate scores for steatosis, lobular inflammation, and hepatocellular ballooning) and to stage fibrosis from 0 to 4. Biopsies of CHC and CHB patients were classified for grade and stage according to Scheuer's system [28].

Statistics

Continuous variables were summarized as mean \pm SD, and categorical variables as frequency and percentage. The Student's ttest and chi-square test were used when appropriate. Multiple logistic regression analyses were done to identify factors independently associated with severe fibrosis in NAFLD, CHC and CHB patients considered separately, and in the sub-groups with high or normal γ -GT serum levels. As candidate risk factors, we selected age, gender, BMI, baseline ALT, normal or high γ -GT (only in the entire populations), triglycerides, total cholesterol, blood glucose, insulin, HOMA score, IR, diabetes, arterial hypertension, steatosis grade, lobular inflammation, ballooning, NAS score, and fibrosis. In these models, the dependent variable was severe fibrosis coded as 1 = present (F3–F4) versus 0 = absent (F0–F2).





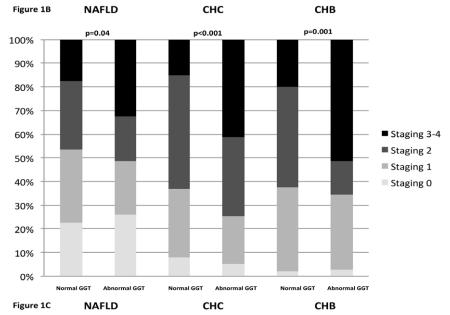


Figure 1. Histological characteristics of patients. (A) Steatosis grade according to liver disease etiology and normal/abnormal γ -GT values; (B) Grade of liver inflammation, according to liver disease etiology (Kleiner score for NAFLD, Scheuer score for CHC and CHB) and normal/abnormal γ -GT values; (C) Stage of liver fibrosis, according to liver disease etiology (Kleiner score for NAFLD, Scheuer score for CHC and CHB) and normal/abnormal γ -GT values.

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Receiver operating characteristic (ROC) curves were applied to identify the area under ROC curve (AUC) of γ -GT and other variables to discriminate the presence of severe liver fibrosis.

Variables associated with the dependent variable on univariate analysis (probability threshold, p≤0.10) were included in the multivariate regression models. To avoid the effect of colinearity, diabetes, IR, HOMA score, blood glucose levels, and insulin levels, as well as NAS score and its components, were not included in the same multivariate model. Regression analyses were done using Proc Logistic, Proc Reg and subroutine in SAS (SAS Institute, Inc., Cary, North Carolina, U.S.A.) [29].

Results

Patient Characteristics and Histology

Abnormal values of serum γ -GT levels were observed in 49.7%, 43.2% and 20.7% of NAFLD, G1 CHC and CHB patients, respectively. The baseline characteristics of the 843 patients according to the etiology of liver disease, and to the presence of normal or high γ -GT serum levels are shown in Table 1 and in Supplemental Table S1. Overall, patients with abnormal γ -GT serum levels had higher ALT serum levels and, considering liver histological features, a more severe liver disease, in term of fibrosis in NAFLD, and of steatosis, inflammatory activity and fibrosis in both CHC and CHB (Figure 1). In addition, patients with abnormal γ -GT had more frequent metabolic alterations, namely higher cholesterol and triglycerides levels in NAFLD, higher triglycerides levels and a higher prevalence of IR and diabetes in CHC, and higher cholesterol and triglycerides and a higher prevalence of IR and hypertension in CHB.

Due to the evidence of a higher prevalence of IR in CHC and CHB patients with abnormal \gamma-GT values compared to their counterpart, in both CHC and CHB groups we tested if this association was maintained at multivariate analysis. We found that, after confirming older age (OR 1.025, CI 1.006-1.045, p = 0.01 for CHC), and higher BMI (OR 1.171, CI 1.105–1.242, p < 0.001 for CHC; OR 1.211, CI 1.071–1.369, p = 0.002 for CHB) as independent predictors of IR, abnormal y-GT values remained independently associated with IR in CHC patients only (OR 1.861, CI 1.228–2.821, p = 0.003). In this group of patients the AUC of γ-GT values for IR prediction was 0.652 (C.I. 0.603– 0.705), compared to 0.609 (C.I. 0.603-0.705) (p = 0.22) of age, and 0.689 (C.I. 0.642-0.736) (p = 0.27) of BMI. Finally, the positive predictive value (PPV) of abnormal γ-GT for IR prediction was 55.2%, while the negative predictive value (NPV) of normal γ -GT was 64.1%.

Factors Associated with Severe Liver Fibrosis in the Entire Cohort of Patients with CLD

Severe fibrosis (F2–F4) was independently associated with abnormal γ GT values in NAFLD (OR 2.711, CI 1.120–6.564, p=0.02), G1 CHC (OR 3.461, CI 2.138–5.603, p<0.001), and CHB (OR 2.778, CI 1.042–7.414, p=0.04) patients, together with other well known risk factors for fibrosis like older age, IR and lobular inflammation in NAFLD (Table 2 up), older age, lower cholesterol, IR and moderate-severe necroinflammatory activity in G1 CHC (Table 2 medium), and older age, virological status, IR and moderate-severe necroinflammatory activity in CHB (Table 2

bottom) patients. The AUC of the entire models were 0.830 (C.I. 0.769–0.880) for NAFLD, 0.798 (C.I. 0.759–0.833) for CHC, and 0.855 (C.I. 0.793–0.904) for CHB. The PPV and NPV of abnormal and normal γGT values, respectively, for severe fibrosis prediction, were reported in table 3.

Finally, when replacing in the model γGT as categorical variable, with γGT as continuous variable, the latter remained significantly associated with severe fibrosis in NAFLD (OR 1.003, CI 1.001–1.006, p=0.04), G1 CHC (OR 1.004, CI 1.001–1.007, p=0.004), and CHB patients (OR 1.022, CI 1.005–1.039, p=0.01). The AUC of γGT values for severe fibrosis prediction, compared to those of the other variables, were reported in table 3.

Factors Associated with Liver Fibrosis in Patients with CLD and with Normal γ GT Serum Levels

In patients with normal γGT values, severe fibrosis was independently associated with older age (OR 1.070, CI 1.013–1.129, p=0.01) and lobular inflammation (OR 3.215, CI 1.150–8.987, p=0.02) in NAFLD (Table 4 up), older age (OR 1.034, CI 1.001–1.070, p=0.04), no male gender (OR 0.295, CI 0.129–0.673, p=0.004), and lower cholesterol levels (OR 0.981, CI 0.969–0.993, p=0.003) in G1 CHC (Table 4 medium), and with only moderate-severe liver necroinflammatroy activity (OR 15.985, CI 1.894–134.893, p=0.01) in CHB (Table 4 bottom) patients.

Factors Associated with Liver Fibrosis in Patients with CLD and with Abnormal γGT Serum Levels

In patients with abnormal γGT values, severe fibrosis was independently associated with older age (OR 1.061, CI 1.015–1.111, p=0.01), lower cholesterol levels (OR 0.982, CI 0.968–0.996, p=0.01), IR (OR 4.150, CI 1.079–15.970, p=0.03), and lobular inflammation (OR 6.824, CI 2.228–20.904, p=0.001) in NAFLD (Table 5 up), with older age (OR 1.049, CI 1.020–1.079, p=0.001), IR (OR 2.250, CI 1.211–4.181, p=0.01), and moderate-severe liver necroinflammatory activity (OR 8.970, CI 2.559–31.447, p=0.001) in G1 CHC (Table 5 medium), and with virological status (OR 13.757, CI 1.013–189.931, p=0.04), and IR (OR 3.096, CI 2.050–34.220, p=0.01) in CHB (Table 5 bottom) patients.

Discussion

In a large cohort of patients with an histological diagnosis of CLD, other than to confirm the independent association between higher $\gamma\text{-}GT$ serum levels and severity of liver fibrosis, we showed that, IR is independently associated with severe fibrosis only in patients with abnormal $\gamma\text{-}GT$ values, without differences according to the metabolic (NAFLD) or viral (HCV or HBV) etiology of the underlying liver disease.

In patients with CLD the severity of liver fibrosis is the strongest predictor of liver related morbidity and mortality, being, in a context of genetic predisposition, IR and liver necroinflammation two of the most important factors associated with the severity and the progression of liver disease [3–8]. Specifically, IR has been clearly associated with fibrosis severity in NAFLD and CHC patients, while contrasting data exist in CHB. In our study, we confirmed both IR and liver necroinflammation as independently

Table 1. Demographic, Clinical, Biochemical, and Histological Features of 843 Patients with Metabolic and Viral Chronic Liver Diseases, According to the Etiology and to γ -glutamiltranspherase Serum Levels.

| Variable | Non-alcoholic N=193 | Non-alcoholic Fatty Liver Disease N = 193 | | Genotype 1 Chronic Hepatitis C N=481 | epatitis C N=481 | | Chronic Hepatitis B N=169 | itis B N=169 | |
|--------------------------------------|-----------------------------|--|---------|--------------------------------------|-------------------------|---------|---------------------------|----------------------|---------|
| | normal γ GT (n = 97) | abnormal yGT (n=96) P value normal yGT n=273 | P value | normal γ GT n=273 | abnormal γGT n = 208 | P value | normal γGT n = 134 | abnormal γGT n=35 | P value |
| Mean age – yrs | 44.7±13.6 | 46.7±13.2 | 0.31 | 51.9±12.1 | 54.2±11.8 | 0.03 | 38.6±13.7 | 46.7±13.1 | 0.002 |
| Male Gender | 78 | 50 | <0.001 | 131 | 111 | 0.24 | 102 | 21 | 0.05 |
| Mean body mass index – kg/m² | 30.0±4.5 | 29.5 ± 4.8 | 0.42 | 26.2 ± 4.2 | 26.7±4.1 | 0.22 | 25.4±3.2 | 26.0±3.3 | 0.37 |
| Alanine aminotransferase – IU/L | 70.0 ± 56.7 | 89.2±52.9 | 0.01 | 61.7±38.2 | 128.4±128.8 | <0.001 | 93.7±94.9 | 138.5±99.5 | 0.01 |
| γ -glutamiltranspherase –IU/L | 32.8 ± 13.7 | 155.9±143.2 | <0.001 | 27.5±13.4 | 135.4 ± 155.5 | <0.001 | 28.5±13.3 | 83.5 ± 46.9 | <0.001 |
| Cholesterol – mg/Dl | 192.8±42.0 | 217.0±49.9 | <0.001 | 175.9±36.0 | 173.8±36.4 | 0.53 | 185.2±39.1 | 211.7±57.7 | 0.01 |
| Triglycerides – mg/dL | 136.4 ± 69.8 | 161.9±83.1 | 0.02 | 88.2 ± 40.0 | 107.2±49.1 | <0.001 | 96.7±33.2 | 117.2±66.6 | 0.01 |
| Blood glucose – mg/dL | 97.4±23.0 | 101.1±37.8 | 0.41 | 92.9±25.4 | 103.7 ± 42.3 | 0.001 | 88.5±16.8 | 98.0±28.5 | 0.01 |
| Insulin – μU/mL | 16.3 ± 18.5 | 18.2±11.8 | 0.20 | 12.3±8.9 | 14.1 ± 7.5 | 0.02 | 10.0±6.9 | 12.0 ± 8.0 | 0.15 |
| HOMA-score | 4.05 ± 2.95 | 4.73±3.93 | 0.18 | 2.97±2.23 | 3.56±2.64 | 0.001 | 2.21±1.60 | 3.02±2.37 | 0.01 |
| Insulin Resistance | 44 | 49 | 0.43 | 86 | 115 | <0.001 | 29 | 13 | 0.05 |
| Type 2 diabetes | 17 | 16 | 0.84 | 29 | 40 | 0.008 | Э | 8 | 0.07 |
| Stage of Fibrosis* | | | | | | | | | |
| 0 | 22 | 25 | | 22 | 11 | | 7 | - | |
| - | 30 | 22 | | 62 | 42 | | 46 | 11 | |
| 2 | 28 | 18 | | 131 | 69 | | 55 | 5 | |
| 8 | 15 | 20 | | 29 | 48 | | 18 | 8 | |
| 4 | 2 | 11 | 0.04 | 12 | 38 | <0.001 | 8 | 10 | 0.001 |

Data are given as mean \pm SD or as number of cases. Yrs, indicates years; HOMA, homeostasis model assessment. * Kleiner score for non-alcoholic fatty liver disease patients, and Scheuer score for genotype 1 chronic hepatitis C, and chronic hepatitis B patients. doi:10.1371/journal.pone.0051165.t001

Table 2. Univariate and Multivariate Analysis of Risk Factors Associated with Severe Fibrosis (F3–F4) in 843 Patients with Metabolic and Viral Chronic Liver Diseases by Logistic Regression Analysis.

| Variable | No Severe Fibrosis (F0-F2) | Severe Fibrosis (F3-F4) | Univariate Analysis <i>p</i> value | Multivariate Analysis OR (95% CI) <i>p</i> value |
|-----------------------------------|-------------------------------|----------------------------|--|--|
| Non-alcoholic Fatty Liver Disease | | | | |
| | n = 144 | n = 49 | | |
| Age – yrs | 42.9±12.6 | 54.0±12.3 | <0.001 | 1.063 (1.028–1.100) <0.001 |
| Male Gender | 104 | 40 | 0.003 | 1.079 (0.457–2.546) 0.86 |
| Body Mass Index – Kg/m2 | 29.3±4.7 | 31.1±4.2 | 0.01 | 1.044 (0.948–1.150) 0.37 |
| Abnormal γ-GT | 65 | 31 | 0.02 | 2.711 (1.120–6.564) 0.02 |
| Insulin Resistance | 57 | 36 | < 0.001 | 2.749 (1.153–6.556) 0.02 |
| Histology at Biopsy | | | | |
| Lobular Inflammation 0/1/2/3 | 11/84/46/3 | 0/12/32/5 | <0.001 | 4.531 (2.244–9.147) < 0.001 |
| Genotype 1 Chronic Hepatitis C | | | | |
| | n = 354 | n = 127 | | |
| Age – yrs | 51.1±12.3 | 57.8±9.5 | <0.001 | 1.044 (1.022–1.067) <0.001 |
| Male Gender | 185 | 54 | 0.06 | 0.639 (0.397–1.028) 0.07 |
| Abnormal γ -GT | 122 | 86 | < 0.001 | 3.461 (2.138–5.603) < 0.001 |
| Cholesterol – mg/Dl | 178.1±34.9 | 166.5±38.2 | < 0.001 | 0.991 (0.984–0.998) 0.01 |
| Insulin Resistance | 133 | 80 | < 0.001 | 1.688 (1.043–2.731) 0.03 |
| Histology at Biopsy | | | | |
| Steatosis grade 0/1/2/3 | 198/118/32/6 | 43/50/28/6 | < 0.001 | 1.275 (0.950–1.711) 0.10 |
| Grading 1 vs 2-3 | 92/262 | 6/121 | <0.001 | 5.767 (2.327–14.289) < 0.001 |
| Chronic Hepatitis B | | | | |
| | n = 125 | n = 44 | | |
| Age – yrs | 37.0±13.6 | 49.4±10.8 | <0.001 | 1.044 (1.003–1.088) 0.03 |
| Abnormal γ-GT | 17 | 18 | < 0.001 | 2.778 (1.041–7.414) 0.04 |
| Virological status | | | | |
| HBe+/HBe-/HBe-Anti-HDV+ | 26/94/5 | 2/33/9 | < 0.001 | 3.303 (1.229–8.877) 0.01 |
| Insulin Resistance | 105/20 | 22/22 | < 0.001 | 2.405 (1.038–6.197) 0.04 |
| Histology at Biopsy | | | | |
| Steatosis grade 0/1/2/3 | 88/30/7/0 | 14/25/3/2 | <0.001 | 1.929 (0.978–3.804) 0.07 |
| Grading 1 vs 2–3 | 50/75 | 3/41 | < 0.001 | 7.326 (1.690–31.754) 0.008 |

Data are given as mean \pm SD or as number of cases. Yrs, indicates years; γ -GT, γ -glutamiltranspherase. doi:10.1371/journal.pone.0051165.t002

related to the severity of liver fibrosis in patients with NAFLD and CHC, adding data pros this association in CHB.

In our study, we identified in high γ -GT serum levels a risk factor for severe fibrosis. This feature is in accord not only to when highlighted by other authors in NASH [19], and to the evidences about an association between γ -GT serum levels reduction and regression of liver damage in NAFLD patients after bariatric surgery [20], but also to different studies showing an independent link between higher γ -GT serum levels and severity of fibrosis in both CHC [21] and CHB patients [22], and potentially to the role of γ -GT serum levels as predictor of all-cause deaths, including liver-related death [20]. Interestingly, in our cohort, the presence of abnormal γ -GT, had a NPV of more than 80% for rule-out severe fibrosis in all patients, independently of the aetiology of liver

disease. This data is encouraging, even if, in clinical practice, we are not confident to suggest to use γ -GT as a single test to predict or rule-out fibrosis severity, but as a potential useful component of noninvasive scores.

Considering therefore the association between γ -GT and liver disease severity, the evidences about a link between γ -GT values and metabolic alterations, like visceral obesity, diabetes, etc, in both cross-sectional and prospective studies [15,16], and also observed in our population, and the sometime contrasting evidences on the role of IR in liver disease severity [4,9,10] and in NAFLD treatment [11,12], we speculated that discriminating NAFLD patients according to normal or abnormal γ -GT values, we can identify different patterns of liver disease where metabolic alterations, and in particular IR, could have a different

Table 3. Diagnostic performance of GGT in predicting severe liver fibrosis.

| | AUC | C.I. 95% | p* | | | |
|--|--------------------------|-------------|-------|--|--|--|
| | Non-alcoholic fatty live | er disease | | | | |
| γ-GT | 0.574 | 0.484-0.664 | - | | | |
| Age | 0.739 | 0.654-0.834 | 0.009 | | | |
| нома | 0.752 | 0.670-0.834 | 0.004 | | | |
| | Chronic Hepatitis C | | | | | |
| ү-GТ | 0.712 | 0.661-0.764 | - | | | |
| Age | 0.659 | 0.606–0.712 | 0.16 | | | |
| нома | 0.615 | 0.556-0.673 | 0.007 | | | |
| Total cholesterol | 0.607 | 0.562-0.651 | 0.01 | | | |
| | Chronic Hepatitis B | | | | | |
| γ-GT | 0.770 | 0.696-0.845 | - | | | |
| Age | 0.759 | 0.683-0.836 | 0.81 | | | |
| НОМА | 0.693 | 0.598-0.787 | 0.007 | | | |
| PPV and NPV of abnormal/normal γ -GT for predicting severe liver fibrosis | | | | | | |
| | PPV (%) | NPV (%) | | | | |
| NAFLD | 32.2 | 81.4 | | | | |
| СНС | 41.3 | 84.9 | | | | |
| СНВ | 51.4 | 80.5 | | | | |

HOMA, homeostasis model assessment; γ -GT, γ -glutamiltranspherase; PPV, positive predictive value; NPV, negative predictive value. * p value of corresponding AUC compared to γ -GT AUC (pairwise comparison). doi:10.1371/journal.pone.0051165.t003

impact. In this line, for the first time to our knowledge, we found that, while classical host and viral risk factors were confirmed as independently linked to the severity of liver fibrosis in NAFLD, CHC and CHB patients with high or normal γ -GT values, by contrast IR remained significantly associated with liver fibrosis only in patients with abnormal γ -GT levels nevertheless similar HOMA values were observed in those with normal or high γ -GT levels, and independently of the etiology of the underlying liver disease.

Although this study was not designed to clarify this interesting finding, a few hypotheses may be put forward according to the literature. A German cohort study found that hepatic steatosis is a significant predictor of all-cause and cardiac-related mortality in men only when it was associated with high γ -GT levels [30]. Another population Italian study also showed that, considering a non-invasive index for steatosis diagnosis (fatty liver index) composed by different metabolic traits including γ -GT, only γ -GT was predictive of all-cause mortality in a context of presence of other metabolic alterations [31]. Finally it has recently been described that serum γ-GT levels have shared genetic determinants which may include the b-2 adrenergic receptor gene [32] and loci interacting with cardiometabolic disease alterations [33]. Overall these data could suggest that γ -GT, in a context of metabolic disturbances, may reflect genetic and metabolic determinants which are associated with poorer outcomes. In this line, according to these data and to our findings on patients with CLD due to metabolic or viral ones, it should be possible to speculate that high γ -GT levels identify a group of liver patients whit a metabolic milieu probably also influenced by genetic background, and able to participate in

liver damage affecting the severity of disease. Instead normal γ -GT levels probably identify another group of patients with CLD where metabolic alterations, and in particular IR, even if present, have not a dominant role in affecting disease severity. By contrast in these patients conventional risk factors, like liver inflammation, are the main determinant of disease severity, suggesting a potential dominant role of inflammatory genetic background. In this line we recently reported that IL28B polymorphisms are able to affect severity of liver inflammation in NAFLD patients independently of metabolic alterations [34].

From a clinical standpoint, the identification of these two different patterns of CLD according to \gamma-GT values, could be useful in the management of these patients. In fact overall our results might also suggest to evaluate if γ-GT can identify NAFLD, CHC and CHB patients at different liver disease progression risk and with different natural history, or where the correction of IR is of further help. In this line our data could help to better understand because: 1) IR is associated with fibrosis severity in a great proportion but not in all studies on CHC patients; and 2) very contrasting results exist about the association between IR and severity of fibrosis in CHB. In addition, considering NAFLD, even if is well known that lifestyle correction is the gold standard treatment for these patients [35], conflicting results exist about pharmacologic therapy. In particular literature data showed that insulin sensitizer [11] and antioxidant drugs [12] could improve liver histological damage, but only in a proportion of NAFLD patients. The discrimination of NAFLD patients according to γ -GT values could therefore help to distinguish patients more sensitive to an approach (lifestyle and/or pharmacological)

Table 4. Univariate and Multivariate Analysis of Risk Factors Associated with Severe Fibrosis (F3–F4) in 504 Patients with Metabolic and Viral Chronic Liver Disease and normal GGT values, by Logistic Regression Analysis.

| Variable | No Severe Fibrosis (F0-F2) | Severe Fibrosis (F3–F4) | Univariate Analysis <i>p</i> value | Multivariate Analysis OR (95% CI) <i>p</i> value |
|-----------------------------------|-------------------------------|----------------------------|--|--|
| Non-alcoholic Fatty Liver Disease | | | | |
| | n=79 | n = 18 | | |
| Mean age – yrs | 42.5±13.1 | 54.6±11.1 | 0.01 | 1.070 (1.013–1.129) 0.01 |
| Male Gender | 12 | 7 | 0.02 | 2.132 (0.538-8.458) 0.28 |
| Mean body mass index – kg/m² | 29.6±4.5 | 31.8±3.9 | 0.05 | 1.028 (0.881–1.199) 0.72 |
| Insulin Resistance | 32 | 12 | 0.04 | 2.555 (0.671–9.727) 0.16 |
| Histology | | | | |
| Lobular inflammation 0/1/2/3 | 3/45/28/3 | 0/4/13/1 | 0.02 | 3.215 (1.150–8.987) 0.02 |
| Genotype 1 Chronic Hepatitis C | | | | |
| | n = 232 | n = 41 | | |
| Age – yrs | 50.9±12.3 | 57.3±8.5 | 0.002 | 1.034 (1.001–1.070) 0.04 |
| Male Gender | 100 | 31 | < 0.001 | 0.295 (0.129–0.673) 0.004 |
| Cholesterol – mg/Dl | 179.5±35.5 | 155.6±31.8 | <0.001 | 0.981 (0.969–0.993) 0.003 |
| Insulin Resistance | 76 | 22 | 0.01 | 1.331 (0.613–2.889) 0.46 |
| Histology at Biopsy | | | | |
| Steatosis grade 0/1/2/3 | 148/71/11/2 | 18/16/6/1 | 0.02 | 1.605 (0.910-2.830) 0.10 |
| Grading 1 vs 2–3 | 60/172 | 3/38 | 0.009 | 3.143 (0.873–11.316) 0.08 |
| Chronic Hepatitis B | | | | |
| | n = 108 | n = 26 | | |
| Age – yrs | 36.5±13.5 | 47.1±11.6 | <0.001 | 1.037 (0.992–1.085) 0.10 |
| Virological status | | | | |
| HBe+/HBe-/HBe-Anti-HDV+ | 22/81/5 | 1/20/5 | 0.009 | 2.666 (0.879–8.087) 0.08 |
| Insulin Resistance | 19 | 10 | 0.02 | 1.383 (0.460–4.155) 0.56 |
| Histology at Biopsy | | | | |
| Steatosis grade 0/1/2/3 | 76/27/5/0 | 10/14/2/0 | 0.009 | 1.917 (0.831–4.425) 0.12 |
| Grading 1 vs 2-3 | 46/62 | 1/25 | < 0.001 | 15.985 (1.894–134.892) 0.01 |

Data are given as mean \pm SD or as number of cases. Yrs, indicates years; γ -GT, γ -glutamiltranspherase. doi:10.1371/journal.pone.0051165.t004

aimed to correct metabolic dysfunctions, to patients more sensitive to drugs acting on oxidative stress and liver inflammation. Our hypotheses would seem to suggest using γ -GT as a way of stratify NAFLD patients in intervention trials, even if, clearly, further studies must be done, in diverse settings and with larger cohorts of patients, before applying this method in clinical practice.

The main limitation of this study lies in its cross-sectional nature, making it impossible to dissect the temporal relation among γ -GT values, liver fibrosis, and both metabolic and liver inflammatory findings in patients with CLD. A further methodological question is the potentially limited external validity of the results for different populations and settings. Our study included a cohort of Italian patients enrolled at a tertiary care center, who may be different from the majority of cases of CLD in the general population. In particular we enrolled patients with abnormal ALT levels and, for CHC and CHB, with a high viral load; therefore

our results could be applied only to this setting of patients and not to those with a milder liver disease. An hidden alcohol abuse, and lack of data on genetic polymorphisms affecting CLD severity and γ -GT serum levels may also have affected interpretation of the results.

In conclusion, we confirmed γ -GT serum levels as a marker of fibrosis in patients with CLD useful to be included in noninvasive scores, and observed that IR is independently linked to liver fibrosis only in patients with abnormal γ -GT values, without differences according to the metabolic (NAFLD) or viral (HCV or HBV) etiology of the underlying liver disease, and suggesting a role of γ -GT as a marker of metabolic-induced liver damage. These data need further validation in independent, large scale studies, potentially representing an useful tools for the clinical and pharmacologic management of patients with CLD.

Table 5. Univariate and Multivariate Analysis of Risk Factors Associated with Severe Fibrosis (F3–F4) in 339 Patients with chronic liver disease (Chronic Hepatitis C, Non-alcoholic Fatty Liver Disease and Chronic Hepatitis B) and abnormal GGT values, by Logistic Regression Analysis.

| Variable | No Severe Fibrosis (F0–F2) | Severe Fibrosis (F3–F4) | Univariate Analysis <i>p</i> value | Multivariate Analysis OR (95% CI) p value |
|-----------------------------------|-------------------------------|----------------------------|--|---|
| Non-alcoholic Fatty Liver Disease | | | | |
| | n=65 | n = 31 | | |
| Mean age – yrs | 43.6±11.5 | 49.7±14.2 | 0.02 | 1.061 (1.015–1.111) 0.01 |
| Mean body mass index – kg/m² | 28.9±4.9 | 30.7±4.3 | 0.08 | 1.046 (0.911–1.201) 0.52 |
| Cholesterol – mg/Dl | 225.6±49.3 | 200.4±47.6 | 0.02 | 0.982 (0.968–0.996) 0.01 |
| Insulin Resistance | 25 | 24 | < 0.001 | 4.150 (1.079–15.970) 0.03 |
| Histology | | | | |
| Lobular inflammation 0/1/2/3 | 8/39/18/0 | 0/8/19/4 | < 0.001 | 6.824 (2.228–20.904) 0.001 |
| Genotype 1 Chronic Hepatitis C | | | | |
| | n = 122 | n = 86 | | |
| Age – yrs | 51.6±12.3 | 58.0±10.0 | <0.001 | 1.049 (1.020–1.079) 0.001 |
| Insulin Resistance | 65/57 | 28/58 | 0.003 | 2.250 (1.211–4.181) 0.01 |
| Histology at Biopsy | | | | |
| Grading 1 vs 2–3 | 32/90 | 3/83 | < 0.001 | 8.970 (2.559–31.447) 0.001 |
| Chronic Hepatitis B | | | | |
| | n = 17 | n = 18 | | |
| Virological status | | | | |
| HBe+/HBe-/HBe-Anti-HDV+ | 4/13/0 | 1/13/4 | 0.05 | 13.757 (1.013–189.931) 0.04 |
| Insulin Resistance | 16/1 | 6/12 | <0.001 | 3.096 (2.050–34.220) 0.01 |
| Histology at Biopsy | | | | |
| Steatosis grade 0/1/2/3 | 12/13/2/0 | 4/11/1/2 | 0.01 | 1.706 (0.316-9.209) 0.53 |

Data are given as mean \pm SD or as number of cases. Yrs, indicates years; γ -GT, γ -glutamiltranspherase. doi:10.1371/journal.pone.0051165.t005

Supporting Information

Table S1 Demographic, Clinical, Biochemical, and Histological Features of 843 Patients with Metabolic and Viral Chronic Liver Diseases, According to the Etiology and to γ -glutamiltranspherase Serum Levels.

References

- Herzer K, Sprinzl MF, Galle PR (2007) Hepatitis viruses: Live and let die. Liver Int 27: 293–301.
- Petta S, Muratore C, Craxì A (2009) Non-alcoholic fatty liver disease pathogenesis: the present and the future. Dig Liver Dis 41: 615–25.
- Petta S, Craxì A (2010) Hepatocellular carcinoma and non-alcoholic fatty liver disease: from a clinical to a molecular association. Curr Pharm Des 16: 741–52.
- disease: from a clinical to a molecular association. Curr Finarth Des 10: 741–32.
 Petta S, Amato M, Cabibi D, Cammà C, Di Marco V, et al (2010) Visceral adiposity index is associated with histological findings and high viral load in
- patients with chronic hepatitis C due to genotype 1. Hepatology 52: 1543–52.

 5. Cammà C, Petta S (2009) Insulin resistance in HCV mono-infected and in HIV/HCV co-infected patients: Looking to the future. J Hepatol 50: 648–51.
- Petta S, Cammà C, Di Marco V, Alessi N, Cabibi D, et al (2008) Insulin resistance and diabetes increase fibrosis in the liver of patients with genotype 1 HCV infection. Am J Gastroenterol 103: 1136–44.
- Everhart JE, Lok AS, Kim HY, Morgan TR, Lindsay KL, et al (2009) HALT-C Trial Group. Weight-related effects on disease progression in the hepatitis C antiviral long-term treatment against cirrhosis trial. Gastroenterology 137: 549– 57.

(DOC)

Author Contributions

Conceived and designed the experiments: SP CC VD AC. Performed the experiments: FSM MRB DC. Analyzed the data: SP. Wrote the paper: SP.

- Petta S, Cammà C, Di Marco V, Macaluso FS, Maida M, et al (2011) Hepatic steatosis and insulin resistance are associated with severe fibrosis in patients with chronic hepatitis caused by HBV or HCV infection. Liver Int 31: 507–15.
- Cammà C, Bruno S, Di Marco V, Di Bona D, Rumi M, et al (2006) Insulin resistance is associated with steatosis in nondiabetic patients with genotype 1 chronic hepatitis C. Hepatology 43: 64–71.
- Park SH, Kim DJ, Lee HY (2009) Insulin resistance is not associated with histologic severity in nondiabetic, noncirrhotic patients with chronic hepatitis B virus infection. Am J Gastroenterol 104: 1135–9.
- Belfort R, Harrison SA, Brown K, Darland C, Finch J, et al (2006) A Placebo-Controlled Trial of Pioglitazone in Subjects with Nonalcoholic Steatohepatitis. NEJM 355;22997–2307.
- Sanyal AJ, Chalasani N, Kowdley KV, McCullough A, Diehl AM, et al (2010) Pioglitazone, Vitamin E, or Placebo for Nonalcoholic Steatohepatitis. NEJM 362: 1675–85.
- Nemesanszky E, Lott JA (1985) Gamma-glutamyltransferase and its isoenzymes: progress and problems. Clin Chem 31: 797–803.

- Whitfield JB. Gamma glutamyl transferase (2001) Crit Rev Clin Lab Sci 38: 263–355.
- Lee DS, Evans JC, Robins SJ, Wilson PW, Albano I, et al (2007) Gamma glutamyl transferase and metabolic syndrome, cardiovascular disease, and mortality risk: the Framingham Heart Study. Arterioscler Thromb Vasc Biol 27: 197–133
- Lee DH, Jacobs DR Jr, Gross M, Kiefe CI, Roseman J, et al (2003) Gammaglutamyltransferase is a predictor of incident diabetes and hypertension: the Coronary Artery Risk Development in Young Adults (CARDIA) Study. Clin Chem 49: 1358–1366.
- Kazemi-Shirazi L, Endler G, Winkler S, Schickbauer T, Wagner O, et al (2007) Gamma glutamyltransferase and long-term survival: is it just the liver? Clin Chem 53: 940–946.
- Jo SK, Lee WY, Rhee EJ, Won JC, Jung CH, et al (2009) Serum gammaglutamyl transferase activity predicts future development of metabolic syndrome defined by 2 different criteria. Clin Chim Acta 403: 234–40.
- Leite NC, Villela-Nogueira CA, Pannain VL, Bottino AC, Rezende GF, et al (2011) Histopathological stages of nonalcoholic fatty liver disease in type 2 diabetes: prevalences and correlated factors. Liver Int 31: 700–6.
- Dixon JB, Bhathal PS, O'Brien PE (2006) Weight loss and non-alcoholic fatty liver disease: falls in gamma-glutamyl transferase concentrations are associated with histologic improvement. Obes Surg 16: 1278–1286.
 Poynard T, Imbert-Bismut F, Ratziu V, Chevret S, Jardel C, et al (2002)
- Poynard T, Imbert-Bismut F, Ratziu V, Chevret S, Jardel C, et al (2002) Biochemical markers of liver fibrosis in patients infected by hepatitis C virus: longitudinal validation in a randomized trial. J Viral Hepat 9: 128–33.
- Seto WK, Lee CF, Lai CL, Ip PP, Fong DY, et al (2011) A new model using routinely available clinical parameters to predict significant liver fibrosis in chronic hepatitis B. PLoS One 6(8): e23077.
- American Diabetes Association. Report of the Expert Committee on the diagnosis and classification of diabetes mellitus (2000) American Diabetes Association: Clinical Practice Recommendations 2000 Committee Report. Diabetes Care 23: S4–19.
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, et al (1985) Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia 28: 412– 410

- Ikeda Y, Suehiro T, Nakamura T, Kumon Y, Hashimoto K (2001) Clinical significance of the insulin resistance index as assessed by homeostasis model assessment. Endocr J 48: 81–86.
- Moucari R, Asselah T, Cazals-Hatem D, Voitot H, Boyer N, et al (2008) Insulin resistance in chronic hepatitis C: association with genotypes 1 and 4, serum HCV RNA level, and liver fibrosis. Gastroenterology 134: 416–23.
- Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, et al (2005) Design and validation of a histological scoring system for nonalcoholic fatty liver disease. Hepatology 411: 313–321.
- Scheuer PJ (1991) Classification of chronic viral hepatitis: a need for reassessment. J Hepatol 13: 372–4.
- SAS Technical Report (1992) SAS/STAT software: changes and enhancement, Release 6.07. Cary, NC: SAS Institute, Inc.
- Haring R, Wallaschofski H, Nauck M, Dörr M, Baumeister SE, et al (2009) Ultrasonographic hepatic steatosis increases prediction of mortality risk from elevated serum gamma-glutamyl transpeptidase levels. HEPATOLOGY 50: 1403–1411.
- Calori G, Lattuada G, Ragogna F, Garancini MP, Crosignani P, et al (2011) Fatty liver index (FLI) and mortality: The cremona study at the 15(th) year of follow up. HEPATOLOGY 54: 145–152.
- Loomba R, Rao F, Zhang L, Khandrika S, Ziegler MG, et al (2010) Genetic covariance between gamma-glutamyl transpeptidase and fatty liver risk factors: role of beta2-adrenergic receptor genetic variation in twins. Gastroenterology 139: 836–845.
- Middelberg RP, Benyamin B, de Moor MH, Warrington NM, Gordon S, et al (2011) Loci affecting gamma-glutamyl transferase in adults and adolescents show age × SNP interaction and cardiometabolic disease associations. Hum Mol Genet 21: 446–55.
- Petta S, Grimaudo S, Cammà C, Cabibi D, Di Marco V, et al (2011) IL28B and PNPLA3 Polymorphisms Affect Histological Liver Damage in Patients with Non-alcoholic Fatty Liver Disease. J Hep 2012 56: 1356–62.
- Promrat K, Kleiner DE, Niemeier HM, Jackvony E, Kearns M, et al (2010)
 Randomized controlled trial testing the effects of weight loss on nonalcoholic steatohepatitis. Hepatology 51: 121–9.